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BIOLOGICAL DEPOSITS

The following viral strains have been deposited under the Budapest Treaty with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852:

VIRUS	/2	ACCESSION NO.	DATE OF DEPOSIT
Wild type A/Ann Arbor/6 (H2N2) egg passage 2			June 10, 1993
Cold-adapted "Master Sti A/Ann Arbor/6/60 7PI (H2			June 10, 1993

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## FIELD OF THE INVENTION

The present invention relates generally to cold-adapted influenza virus and, more particularly, to a cold-adapted influenza virus vaccine and methods of preventing and treating influenza by employing the vaccine.

## **BACKGROUND OF THE INVENTION**

The tremendous impact of influenza virus infections on the public health is widely recognized. Control of influenza has relied primarily on the use of inactivated influenza vaccines. More current approaches, however, have moved towards the use of live attenuated vaccine. Kilbourne, E.D. "Influenza" (Plenum Publishing Corp. New York), p. 291-332 (1987). The most promising efforts in the development of an effective live vaccine have centered on adapting the virus to grow at suboptimal temperatures. Maassab, H.F., et al., *Vaccine* 3:355-369 (1985). Using this approach, cold-adapted attenuated influenza viruses have been developed in both the former Soviet Union and the United States. Alexandrova, G.I., et al., *Rev. Roum. Inframicrobil*. 2:179-189 (1965); Maassab, H.F. *Nature (London)* 213: 612-614 (1967).

In particular, cold adaptation (*ca*) has permitted the A/Ann Arbor/6/60 (H2N2) (A/AA/6/60) virus of the present invention to grow as well at 25°C as it does at 33°C. Maassab, H.F. *Nature (London)* 213:612-614 (1967); Maassab, H.F. "Biology of Large RNA Viruses" (Academic Press, New York), p. 542-565 (1970). The *ca* A/AA/6/60 virus is also temperature-sensitive (*ts*), a property that impedes replication at higher temperatures in the lungs and thus is highly desirable for live vaccines. Maassab, H.F., "Biology of Large RNA Viruses" (Academic Press, New York), p. 542-565 (1970); Mulder, J., et al., "Influenza" (Wolters-Noordhoff, Amsterdam), 1-6:78-80 (1972). Singlegene studies of this cold-adapted virus in a background of A/Korea/1/82 (H3N2) have identified the genes responsible for the *ca* and *ts* phenotypes and for attenuation in that gene constellation. Snyder, M.H., et al., *J. Virol.* 62(2):488-495 (1988).

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Live attenuated vaccines are produced by reassorting the six internal genes of the cold-adapted A/Ann Arbor/6/60 influenza virus with the two surface genes of the currently circulating wild type (wt) virus, thereby producing a reassortant strain. Maassab, H.F. "Negative Strand Viruses" (Academic Press, New York), p. 755-763 (1975); Davenport, F.M., et al., *J. Infect. Dis.* 136:17-25 (1977). Vaccines prepared from *ca* A/AA/6/60 have proven both non-reactogenic and non-transmissible in preliminary field trials at six different medical centers involving over 20,000 people. Couch, R.B., et al., "Options for the Control of Influenza" (Alan R. Liss, New York), p. 223-241 (1986); Wright, P.F., et al., "Options for the Control of Influenza" (Alan R. Liss, New York), p. 243-253 (1986). These vaccines also provide higher IgA levels than the killed vaccines and afford longer-lasting protection in children. Murphy, B.R., et al., *Infect. Immun.* 36(3):1102-1108 (1982); Johnson, P.R., et al., *J. Infect. Dis.* 154(1):121-127 (1986). Currently, the *ca* A/AA/6/60 7PI (plaque-purified seven times) master strain preparation is under development for use as a live vaccine in children and other live virus vaccines are being developed using the live *ca* influenza vaccine as a model.

Cold-adapted reassortant vaccines have thus been shown to have the proper level of attenuation, immunogenicity, and non-transmissibility combined with proven genetic stability and are produced in acceptable tissue culture substrates. In general, live cold-adapted reassortant vaccines offer several advantages over the existing inactivated vaccine. These include the possible use of a single dose, and administration by the natural route of infection, *i.e.* intranasally. In addition, *ca* vaccines stimulate a wide range of antibody responses, and result in induction of both local and humoral immunity. Furthermore, these vaccines are cost-effective and can be rapidly produced and updated in the event of antigenic changes. In addition, laboratory guidelines are available for the assessment of virulence (reactogenicity in

ferrets) and attenuation can be reproducibly achieved. Moreover, the presence of two phenotypic markers (the temperature-sensitive and cold-adapted phenotypes) allows for the evaluation of virulence and monitoring of the vaccine in the field.

However, despite the above-described advantages, until now virtually nothing has been known about the molecular basis of cold adaptation. Published information indicates that cold adaptation has produced one or more mutations in each of the genes encoding the internal proteins of the A/AA/6/60 master strain. Cox, N.J., et al., "Genetic Variation Among Influenza Viruses" (Academic Press), p. 639-652 (1981). However, all of the work has been done on viruses passaged 28 to 32 times in eggs in parallel with the virus passaged in primary chick kidney cells during cold adaptation. Cox, N.J., et al., *Virol.* 167:554-567 (1988). Studies, however, have shown a gradual buildup of mutations in the RNA1 of sequential 35°C egg passages 2 through 28 of wild type viruses, and recent findings have shown the influence of host cell variation on influenza viruses passaged in chicken eggs. Katz, J.M., et al., *Virol.* 156:386-395 (1987). Thus, the mutations leading to cold adaptation and attenuation have heretofore been unknown.

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It would thus be desirable to isolate and provide the wild type A/Ann Arbor/6/60 progenitor virus and determine the accurate nucleic acid sequence of its genome. It would further be desirable to identify the mutations leading to cold adaptation, thus accurately characterizing the nucleic acid sequence of the *ca* master strain. It would also be desirable to produce and provide cold-adapted influenza strains through reassortment with currently circulating wild type strains. It would also be desirable to produce and use a cold-adapted influenza vaccine to prevent and/or treat influenza.

### SUMMARY OF THE INVENTION

The cold-adapted A/Ann Arbor/6/60 7PI (H2N2) influenza strain ("master strain") has been isolated and deposited, and its genome accurately sequenced and compared to its progenitor temperature-sensitive wild type E2(3) (wt 2(3)) virus. The A/Ann Arbor/6/60 virus is a single-stranded RNA virus having eight gene segments. During investigation of the virus leading to the vaccines of the present invention, unexpected deviations from previously reported sequences of the ca and wt were also identified. In particular, in the ca master strain sequences, seven nucleotide differences were found, occurring in the nucleoprotein gene (NP), the gene encoding an acidic polymerase protein (PA) and the gene encoding a basic polymerase polypeptide (PB2). The wt progenitor strain and ca master strain have both been deposited with the American Type Culture Collection, as set forth above.

In comparing the cold-adapted master strain to the *wt* progenitor strain, four nucleotide differences encoding two amino acid differences were found in three gene segments. Computer-predicted RNA folds projected different secondary structures between the cold-adapted and wild type molecules based on the two silent differences between them. Genes coding for the PA, matrix (M), and non-structural (NS) proteins were identical between the two viruses. The differences suggest that cold adaptation may serve to provide conformational changes in the RNA structure advantageous to growth at 25°C and provide a new form of genetic stability to the highly variable RNA genome.

With the identification of the correct nucleotide sequence of the *ca* master strain and its deposit, reassortant strains can now be produced which can be used as vaccines, to prophylactically and therapeutically treat influenza. Reassortant strains are produced by genetically combining the *ca* master strain with a variety of epidemic wild type viruses to yield reassortants which contain the hemagglutinin (HA) and neuraminidase (NA) gene segments of the wild type virus and the other six genome segments of the *ca* master strain. The reassortants thus contain the epidemic wild type strain genes that code for immunizing antigens found on the surface of the virus particle and the *ca* master strain genes that are responsible for the attenuated phenotype in humans and animals. To produce the vaccines of the present invention, a cold-adapted reassortant vaccine strain is passed once to prepare a virus seed lot which is used to produce vaccine pools.

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In practicing the present invention, the amount of vaccine to be used or administered, alone or in combination with other agents, may vary with the patient being treated and may be monitored on a patient-by-patient basis by the physician. The vaccines of the present invention may also be administered in combination with other vaccines. Generally, a therapeutically effective amount of the vaccine will be administered for a therapeutically effective duration. By "therapeutically effective amount" and "therapeutically effective duration" is meant an amount and duration to achieve the desired therapeutic or prophylactic result in accordance with the present invention with medically acceptable side effects, which can be determined by those skilled in the medical arts.

The vaccines of the present invention may comprise the reassortant virus as well as a pharmaceutical formulation, together with a pharmaceutically acceptable carrier therefor. Each carrier must be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not injurious to the patient. Formulations include those suitable for oral, nasal, topical (including transdermal,

buccal and sublingual), parenteral (including subcutaneous) and pulmonary administration. The formulations may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy.

It will be appreciated that administration of the vaccines of the present invention will also be by procedures well-established in the pharmaceutical arts, e.g. preferably intranasally or orally, and most preferably intranasally. Intramuscular, intravenous and intradermal administration is also contemplated by the present invention, either alone or in combination.

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The present invention thus comprises isolated nucleic and amino acids with sequences corresponding to the *ca* master and wild type strain sequences set forth in Sequence ID Listings 1-40. By "isolated" is meant substantially purified from the natural state through chemical, biochemical, immunological or other means, or obtained in substantially pure form by other methods known to those skilled in the art. By "substantially pure" is meant substantially free from undesirable contaminants such as other proteins. Thus, these terms are not meant to exclude synthetic and recombinant nucleic and amino acids which are contemplated within the scope of the present invention. These terms are also not meant to exclude nucleic and amino acids which are linked, bound or intentionally combined with other moieties such as transgenes, labels, flanking amino acid sequences and the like. It will also be appreciated that although the viruses of the present invention are RNA viruses, the present invention further includes DNA sequences corresponding and complementary thereto.

The present invention further comprises isolated or substantially pure *ca* master strain and wild type E2(3) A/AA/6/60 virus. By "isolated" or "substantially pure strain" is meant the viral strain substantially free from other contaminants such as other viruses, bacteria, and the like.

The present invention further comprises reassortant viruses produced by combining the cold-adapted master strain with a variety of epidemic wild type viruses. The two surface protein genes of an epidemic wild type virus are operatively-linked to the six internal genes of the cold-adapted master strain. By "operatively-linked" is meant attached or assembled in a manner which allows for expression of the surface and internal genes. In the context of reassortant viruses, operative linkage will allow for the packaging of the reassorted RNA into virions. It will also be appreciated that the term "gene" is used comprehensively to include all polynucleotide sequences coding for the gene product or protein, and is not limited to naturally occurring coding and regulating elements.

In addition, the present invention comprises the production and use of coldadapted influenza vaccines to prevent and/or treat influenza.

Additional objects, advantages, and features of the present invention will become apparent from the following description and appended claims, taken in conjunction with the accompanying drawings and Sequence ID Listings.

## **BRIEF DESCRIPTION OF THE DRAWINGS**

The various advantages of the present invention will become apparent to one skilled in the art by reading the following specification and subjoined claims and by referencing the following drawings in which:

Figure 1 shows the derivation of the progenitor wild type and cold-adapted master strain A/AA/6/60 in PCK cells; and

Figure 2 shows the computer-projected RNA fold of cold-adapted and wild type 2(3) RNA1's (PB2's).

## DETAILED DESCRIPTION OF THE INVENTION

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## **OVERVIEW**

The nucleic and amino acid sequences for the eight genes of the cold-adapted master strain A/Ann Arbor/6/60 7PI (H2N2) are set forth in Sequence ID Listings 1-20. The nucleic and amino acid sequences for the eight genes of the wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3) are set forth in Sequence ID Listings 21-40. Table 1 summarizes the gene products of influenza A and B virus genes. The cold-adapted master strain and wild type 2(3) progenitor have been deposited with the ATCC, as described above.

TABLE 1 Gene Products of Influenza A and B Viruses

5	RNA	Gene Product(s)	Function
	1	PB2	Viral polymerase component involved in RNA transcription
	2	PB1	Viral polymerase component with RNA transcription and replication activities
	3	PA	Viral polymerase component involved in RNA replication
	4	HA	Virion surface attachment and fusion glycoprotein, major antigenic determinant
10	5	NA	Virion surface glycoprotein with receptor- destroying enzyme activity, major antigenic determinant
	6	NP	Major nucleocapsid structural component and type-specific antigen
		NB	Glycoprotein pututive membrane ion channel found only in type B
	7 M1		Membrane matrix protein and type-specific antigen
		M2	Nonglycosylated membrane ion channel, found only in type A
15	8	NS1	RNA-binding non-structural protein of transport function
		NS2	Cellular and virion protein of unknown function

The A/Ann Arbor/6/60 virus contains six internal genes, NS, M, NP, PA, basic polymerases (PB1 and PB2), and two surface genes, HA and NA. Seven nucleotide differences were found between the sequences of the present invention and those previously published for cold-adapted A/Ann Arbor/6/60: three in the NP gene, one in the PA gene and three in the PB2 gene. The eight viral genes and the discrepancies in the previously published sequences can be summarized as follows:

NS. The non-structural (NS) gene is the smallest RNA segment of influenza virus, 890 nucleotides long, and codes for the two non-structural proteins (NS1 and NS2) (nucleic acid Sequence ID Listing 1 and 3; amino acid Sequence ID Listings 2 and 4). There were no errors in the previously published sequences for the ca A/AA/6/60 NS1 and NS2 genes.

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M. The matrix gene (M) is a 1,027 base nucleic acid sequence (nucleic acid Sequence ID Listings 5 and 7; amino acid Sequence ID Listings 6 and 8). There were also no errors in the previously published sequences for the ca A/AA/6/60 M gene.

NP. The nucleoprotein gene (NP) (nucleic acid Sequence ID Listing 9) is 1566 nucleotides in length and encodes a basic structural protein of 498 amino acid residues (amino acid Sequence ID Listing 10) which specifically interacts with RNA molecules to form ribonucleoprotein complexes and has sequences that direct its migration into the nuclei of infected cells. Despite previous reports, nucleotide 627 of NP is actually cytosine not adenine, and nucleotide 909 is guanine, not cytosine. In addition, nucleotide 113 was previously published as an adenine, although in GenBank it is reported as a cytosine. Cox, N.J. et al., Virol. 167:554-567 (1988). Regardless of this discrepancy, it is now known that nucleotide 113 is actually a cytosine.

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**PA**. The polymerase acidic protein gene (PA) RNA sequence (nucleic acid Sequence ID Listing 11) is 2233 nucleotides in length and encodes an acidic polymerase protein 716 amino acids in length (amino acid Sequence ID Listing 12). Although previous publications indicate thymine at nucleotide 75 of PA, guanine is actually present at that position.

**PB1**. The polymerase basic 1 gene (PB1) RNA sequence (nucleic acid Sequence ID Listing 13) is 2341 nucleotides in length and encodes a basic polymerase protein 757 amino acids in length (amino acid Sequence ID Listing 14). No errors in the previously published sequence were found.

PB2. The polymerase basic 2 gene (PB2) RNA sequence (nucleic acid Sequence ID Listing 15) is 2341 nucleotides in length and encodes a basic polymerase polypeptide of 759 amino acids (amino acid Sequence ID Listing 16). There are three errors in the previously published sequence for PB2: thymine at 714 instead of the previously published cytosine at that position; guanine at 936 instead of adenine; and cytosine instead of thymine is the predominant base at 1933, with thymine as the secondary base.

HA and NA. The hemagglutinin gene (HA) and neuraminidase gene (NA) code for surface receptors. HA is 1773 nucleotides long and codes for a 562 amino acid sequence (nucleic acid Sequence ID Listing 17; amino acid Sequence ID Listing 18). See Schäfer, J.R. et al. Virol. 194:781-788 (1993). NA is 1467 nucleotides long and codes for a 469 amino acid sequence (nucleic acid Sequence ID Listing 19; amino acid Sequence ID Listing 20).

Results from previous studies indicate that cold adaptation causes mutations in every gene of the A/AA/6/60 master strain, thus ensuring the genetic stability of the virus. There are actually, however, four base differences in three of the internal genes of A/AA/6/60 after 28 passages in primary chicken kidney (PCK) cells and four passages in eggs. Two of the substituted bases are silent and two result in single amino acid differences in two of the genes. Moreover, the *wt* 2(3) progenitor virus is attenuated in ferrets. Hence, the stability and immunogenicity of the *ca* A/AA/6/60 vaccine appears to reflect inherent properties of the *wt* A/AA/6/60 E2(3) virus selected as the progenitor for the master strain. This interpretation is supported by the large number of amino acids unique to both *wt* 2(3) and *ca* viruses (see Table 3), some of which may be attenuating.

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By attempting to identify changes arising from cold adaptation using the ca master strain and the wt 2(3) virus, there is at least one further critical variable passage of the virus in different host tissues. It has been shown that the host cell influences the selection of antigenic variants of influenza viruses. Katz, J.M., et al., Virol. 156:386-395 (1987). In studies of the HA gene of H3N2 viruses, passage in Madin-Darby canine kidney (MDCK) cells and in primary chick kidney (PCK) cells selected populations that were homogeneous and true to the original isolate for this gene whereas passage in eggs selected heterogeneous populations. Katz, J.M., et al., J. Gen. Virol. 73:1159-1165 (1992). Thus, the changes observed could relate to the number of passages of each virus. The wild type 2(3) virus, with only two egg passages, is the only virus among all of those listed in GenBank to have isoleucine encoded by base 1276 of RNA2 and asparagine encoded by base 113 of NP. The positions of those two amino acids in the cold-adapted virus, with 29 PCK passages and 4 egg passages, are the same as those of all other viruses listed in GenBank. This finding suggests that the valine encoded by base 1276 in the cold-adapted PB1 is a host adaptation change rather than a cold adaptation change; the same holds for the threonine encoded by base 113 of the cold-adapted NP gene.

Differences between the *wt* 2(3) sequence as set forth herein and the *wt* 28-32 previously sequenced reflect mutations acquired during high passage in eggs at 35°C. Cox, N.J., et al., *Virol.* 167:554-567 (1988). These mutations may be the result of host adaptation in the egg or simply selection of the highly variable RNA population with the highest relative fitness. Clarke, D.K., et al., *J. Virol.* 67:222-228 (1993).

Since only the *ca* RNA1 has guanine (G) at position 141 and cytosine (C) at position 1933, by comparison with all other human RNA1's in GenBank, the two base changes between the *wt* 2(3) and *ca* RNA1's may in fact be cold-adapted changes.

No wild type human viruses, including the *wt* 2(3) progenitor, have G at 141 or C at 1933. This suggests that cold adaptation may operate at the RNA level. Recent findings indicate that unique RNA structures in influenza viruses may have common regulatory functions. Parvin, J.D., et al., *J. Virol.* 63:5142-5149 (1989). The more stable conformation of the *ca* molecule predicted by base pairing might provide a growth advantage over the predicted conformation of the *wt* 2(3) molecule. The importance of RNA structure to biological function has been well documented for poliovirus. Racaniello, V.R., et al., *Virol.* 155:498-507 (1986). The presence of a hairpin structure at the 5' noncoding end has been shown to be necessary for the *ts* phenotype of the virus.

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Although RNA viruses have notoriously high mutation rates and have been referred to as "quasi-species," Holland, J.J., et al., Cur. Topics Microbiol. Immunol. (Springer-Verlag) 176:1-20 (1992), the ca A/AA/6/60 virus showed unusual stability after cold adaptation in PCK cells. In 33 passages there were only four sequence changes in the six internal genes, yielding a mutation rate of 2 x 10<sup>-6</sup>. Compared to expected chance mutation rates calculated for the NS gene in MDCK cells, one would have expected 21 sequence changes. Parvin, J.D., et al., J. Virol. 59(2):377-383 (1986). Since RNA viruses have not been shown to have proofreading functions, this low mutation rate may be an inherent property of the wild type polymerases or a result of the cold adaptation process, or both. Suarez has shown that wild type viruses comprise subgroups with different mutation rates. Suarez, P., et al., J. Virol. 66(4):2491-2494 (1992). The wt A/AA/6/60 may have a dominant population with a more error-free polymerase. In addition, certain positions may simply be difficult for the polymerase to read, owing to conformation of the RNA molecule. Lowering the growth temperature by 10°C slows the whole replicative process including the speed at which the polymerase unit is moving. Thermus aquaticus (Taq) polymerase is notorious for its high error rate due in part to the high temperature of its use, and it has been shown that a 5°C reduction in temperature increases the fidelity of Tub polymerase. Kainz, P., et al., Anal. Biochem. 202:46-49 (1992). The lower temperature may provide a slowed-down environment conducive to faithful copying even in areas with conformational bends and twists. Thus the A/AA/6/60 polymerase might exhibit greater fidelity at 25°C than at 35°C.

Single gene cold-adapted reassortants, constructed to identify the genetic basis of the *ca* and *ts* phenotypes and of attenuation, should be interpreted with care. For instance, in the study by Snyder et al., conducted in a background of A/Korea/1/82 genes, both PA and M were implicated in attenuation. Snyder, M.H., et

al., J. Virol. 62(2):488-495 (1988). Neither gene showed sequence differences from its wt 2(3) counterpart in the present analysis. This would suggest that single gene wt 2(3) PA or wt 2(3) M in an A/Korea background would react similarly to the ca PA and M single genes. From the sequence data, one would also expect that RNA1 5 encoding PB2 would contribute to the ca phenotype in single gene studies and yet only PA was involved. Snyder, M.H., et al., J. Virol. 62(2):488-495 (1988). Gene constellation studies suggest that single gene studies in one wild type may be applicable to only that wild type. Subbarao, E.K., et al., Virus Res. 25:37-50 (1992). In a different wild type background, the assignment of phenotype to specific ca genes might change because other wild type genes might be dominant or carry natural extragenic suppressor mutations. This emphasizes the need for the presence of six genes from the ca virus rather than five in ca reassortants to ensure maximum stability. Maassab, H.F., et al., J. Infect. Dis. 146(6):780-790 (1982).

### SPECIFIC EXAMPLE 1 - SEQUENCING

#### **A. MATERIALS AND METHODS** 15

Viruses. All viruses were supplied by Professor H.F. Maassab at the University of Michigan and the ca master strain and wild type progenitor strain viruses have now been deposited with the ATCC as previously set forth. Steps in the preparation of the ca master strain A/AA/6/60 7PI (H2N2) live influenza virus and the wt A/AA/6/60 (H2N2) egg passage 2(3) virus are shown in Figure 1. In Figure 1, PCK cells refers to primary chick kidney cells, SPAFAS refers to specific pathogen-free eggs and PI refers to plaque-purified. To guard against any possibility of mix-up in the two viruses, the passage history of both viruses was carefully traced and their separate identities were verified. Moreover, the two viruses were grown in different institutions and sequenced separately. The authenticity of the wt A/AA/6/60 E2(3) virus is supported by sequence differences between the HA's and NA's of the cold-adapted and wild type viruses. Viruses grown in 11-day old embryonated chicken eggs and virion RNA were prepared as previously described. Bean, W.J., et al., Anal. Biochem. 102:228-232 (1980).

Growth and Infectivity of Viruses. Plaque titrations were performed with both viruses in PCK cells at 25°C, 33°C, and 39°C, and in MDCK cells at 33°C and 39°C. Mills, J., et al., J. Infect. Dis. 123:145-157 (1971). Plaque counts obtained at each of the three temperatures were compared to assess the ca and ts phenotypes of both viruses.

Ferret Studies. One week before infection with virus, 4 female ferrets were bled and screened for influenza antibody against A/Taiwan/1/86 (H1N1),

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A/Beijing/353/89 (H3N2), *wt* A/AA/6/60 (H2N2) E2(3) and B/Victoria/2/87. The animals' temperatures were taken twice a day for 1 week preceding their inoculation with 1 x 10<sup>9</sup> EID<sub>50</sub> of *wt* E2(3), and then until they were sacrificed at either 3 or 8 days after infection. Lungs and turbinates of the ferrets were examined by previously reported methods. Maassab, H.F., et al., *J. Infect. Dis.* 146(6):780-790 (1982).

**Gene Cloning.** Double-stranded cDNA was prepared as previously described. Huddleston, J.A., et al., *Nucleic Acids Res.* 10:1029-1039 (1982). Full-length double-stranded copies of genes 4 through 8 (HA, NA, NP, M, NS) were blunt-end ligated into the Pvu II site of vector Pvu II, obtained from C. Naeve at St. Jude Children's Research Hospital.

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For the polymerase genes (PB1, PB2, PA), the first-strand cDNA was amplified by the polymerase chain reaction (PCR) using phosphorylated primers. "Genecleaned" PCR product was blunt-end ligated into the Pvu II site of pATX.

Nucleic Acid Sequencing. Nucleotides of all eight cloned genes of each virus were sequenced by the method of Chen and Seeburg using alkali-denatured DNA templates. Chen, E.Y., et al., DNA 4:165-170 (1985). Due to the extreme heterogeneity of RNA viruses, several clones of each gene were sequenced to avoid reporting the sequence of a minor mutant population. Clones of each orientation were sequenced for each gene. If the two clones differed at any position, as many as 7 clones of each gene were sequenced and the consensus sequence was reported. Compressions were resolved by the addition of 42% formamide to the gels.

Differences between the cold-adapted virus and the wild type E2(3) virus were confirmed by direct sequencing of the virion RNA, a method which would expose any mutations introduced by use of the *Taq* polymerase. Air, G.M. *Virol.* 97:468-472 (1979).

Sequence Analysis. The IntelliGenetics software package (Palo Alto, CA) was used to analyze nucleotide sequence data. Chou-Fasman two-dimensional protein structure predictions were made with programs available at the St. Jude Molecular Biology Computing Center. The reliability of protein folding by this method is predicted to be approximately 60%. Fasman, G.D. "Prediction of Protein Structure and the Principals of Protein Confirmation" (Plenum, New York), p. 417-467 (1986).

The Zuker Fold program on the Cray Y-MP supercomputer at the Pittsburgh Supercomputing Center was used to study the folding of RNA molecules. Optimal foldings were obtained using the Zuker algorithm which calculates the structure exhibiting minimal free energy. Zuker, M., et al., *Nucleic Acids Res.* 9:133-148 (1981). This program calculates the structure that is energetically most favorable and has a

predicted accuracy of 80%, although the structure with the lowest free energy may not represent all biologically active structures. Zuker, M., et al., *Nucleic Acids Res.* 9:133-148 (1981).

## **B. RESULTS**

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Biological Properties. The ca and ts characteristics of the viruses in PCK cells was first examined. The ca master strain reached essentially the same titer at 25°C (3.0 x 10<sup>8</sup>) as it did at 33°C, but failed to grow at 39°C (see Table 2), fulfilling accepted criteria for cold adaptation and temperature sensitivity. By contrast, on day 6, the wt E2(3) virus had produced fewer than 1.0 x 10<sup>5</sup> plaques at 25°C, although by day 8 it had generated 5.0 x 10<sup>6</sup> plaques, indicating a subpopulation of virus capable of growth at low temperatures. The 4-log reduction in growth at 39°C compared with that at 33°C demonstrates the ts phenotype of the wt 2(3) virus. Similar results were obtained in MDCK cells at 33°C and 30°C (data not shown).

The pathogenicity of the wild type 2(3) virus was studied in ferrets. The virus was not recovered from lung tissue in any of the 4 animals examined, and it was recovered from turbinates in only the 2 animals sacrificed on day 3 (data not shown). None of the ferrets showed physical signs of illness, such as coryza, lethargy or sneezing. Rises in temperature ranging from 1°C to 1.5°C were observed, but they persisted for only several hours and were not considered significant since normal temperatures fluctuated by 1°C. These results, which correspond to findings with the ca virus, indicate that the wt 2(3) virus was attenuated before cold adaptation. Maassab, H.F., et al., J. Infect. Dis. 146(6):780-790 (1982).

TABLE 2 Infectivity Titers of A/AA/6/60 (H2N2)

		Number of Plaques in Primary Chick Kidney Cells <sup>a</sup>					
5	Virus	33°Cp	39₀C <sub>p</sub>	25°C			
	ca Master Strain	6.0 x 10 <sup>8</sup>	<1.0 x 10 <sup>4</sup>	5.0 x 10 <sup>7</sup> on day 6°			
	A/AA/6/60 (H2N2)			8.0 x 10 <sup>7</sup> on day 7			
	7PI (SE4)			3.0 x 10 <sup>8</sup> on day 8			
	wt A/AA/6/60	1.5 x 10 <sup>8</sup>	2.0 x 10 <sup>4</sup>	<1.0 x 10 <sup>5</sup> on day 6			
10	(H2N2) E2(3)			8.0 x 10 <sup>5</sup> on day 7			
				5.0 x 10 <sup>6</sup> on day 8			

<sup>a</sup>Similar results were obtained in MDCK cells at 33°C and 39°C.

blnfectivity titers at 33°C and 39°C were determined on post-infection 15 day 4.

<sup>c</sup>Post-infection days.

Tests were also performed employing ferrets to determine whether the coldadapted vaccine would interfere with or block growth of the influenza virus. The experimental protocol and results of this study are set forth in U.S. Patent No. 5,149,531, issued September 22, 1992 to Younger et al., hereby incorporated by reference.

Sequencing. Table 3 compares sequencing results of the ca master strain with wt E2(3) virus. The data represent consensus DNA sequencing of multiple clones. If the clone consensus indicated a difference between the two viruses, RNA sequence data were used to support the findings. Positions reported as mixed populations in Table 3 show the distribution of the clones.

Between the internal genes of the ca and the wt 2(3) viruses, no differences were found in the genes coding for PA, M or NS, even though PA and M were previously reported to be important for attenuation of the ca master strain and cold adaptation was attributed to PA. Snyder, M.H., et al., J. Virol. 62(2):488-495 (1988). Differences were found in the genes coding for PB2, PB1 and NP.

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TABLE 3 Sequence Differences between wt 2(3) and ca A/Ann Arbor/6/60 Viruses

		<u>-</u>		wt A/AA/	wt A/AA/6/60 E2		4/6/60
5	Gene	Base No.	Amino Acid No.	Base	Amino Acid	Base	Amino Acid
	PB2	141		A/g(4/2)		G (5)	
		1933		T/c(4/2)		C/t(4/1)	
	PB1	1276	418	A (5)	lle	G/a(4/3)	Val
	PA			-	-	-	-
10	НА	144	34	A (2)	Asn	T (2)	lle
		455	138	C (2)	Ala	A (2)	Thr
		729	229	A (2)	Lys	C (2)	Thr
	NA	394		C (2)		T (4)	
		604		A (2)		T (4)	
15	NP	113	23	A/c(2/1)	Asn	C/a(3/1)	Thr
	М			-	-	-	-
	NS			-	-	-	_

In Table 3 above, in positions with mixed bases, the capital letter represents the dominant base. The distribution of the clones representing the positions with differences between the wt 2(3) and the ca internal genes are shown next to the bases.

RNA1 (PB2). Two nucleotide differences, in bases 141 and 1933, were found between the ca and wt 2(3) RNA1 genes, which encode a basic polymerase protein 759 amino acids in length. Called PB2, this protein is part of the transcriptase complex and has been identified as recognizing and binding the cap structure of the host-cell primer RNA. Plotch, S.J., et al., Cell 23:847-858 (1981). Both changes are in the coding region but are silent. Moreover, bases 141 and 1933 of the ca RNA1 are unique among all other human RNA1 sequences in GenBank. Position 1933 in the wt 2(3) and ca RNA1 segments is a mixed population of two bases; however, the darker band in the RNA sequence (thymine (T) in wt 2(3) and cytosine (C) in ca) conforms with the consensus DNA sequence reported in Table 3.

To assess the potential functional significance of the two nucleotide sequence differences between the ca and the wt 2(3) viruses, the Zuker RNA-fold algorithm and computer modeling techniques were used to predict RNA secondary structures. As

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shown in Figure 2, the difference at base 141 does not impinge on the predicted structure of RNA1 because it is part of an unpaired loop in both molecules; however, the change at nucleotide 1933, T in *wt* 2(3) to C in *ca* (shown by arrows in Figure 2), does affect the predicted fold of RNA1. The RNA fold of the *ca* virus has greater stability than the analogous fold of *wt* 2(3), as judged by its lower free energy of -736.2 compared to -733.6 for the *wt* 2(3) molecule. Both folds were pivoted -25° at pair 1068/1381 and 180° at pair 1675/1861 to better visualize the area of difference between the two molecules. The single base change at 1933 causes a cascade of 163 pairing differences, from base 1888 to base 2151, and thus might constitute a true cold adaptation. Similar RNA1 sequencing results were obtained for a *wt* A/AA/6/60 E3(4) passage virus.

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RNA2 (PB1). The only nucleotide change found between the RNA2 genes of the ca and wt 2(3) viruses occurred at base 1276, resulting in a substitution of valine (ca) for isoleucine (wt 2(3)), both of which are hydrophobic and uncharged. RNA2 encodes a basic polymerase (PB1) that mediates transcription and elongation of the mRNA chain. Braam, J., et al., Cell 34:609-618 (1983). Analysis of protein secondary structures predicted by Chou-Fasman and Garnier-Osguthorpe methods, as well as computer-predicted RNA structures, failed to reveal differences between the ca and wt 2(3) RNA2's. Valine is not an amino acid unique to the ca virus because later passages of the wt A/AA/6/60 virus (both wt E6 and wt E28) also have valine at this position, as do all other RNA2's in GenBank. Both DNA clones and RNA sequencing show that base 1276 comprises a mixed population of adenine (A) and guanine (G) in the ca RNA2; however, the G predominates.

**RNA6** (NP). The nucleoprotein gene (RNA6) encodes a basic protein 498 amino acids in length which specifically interacts with RNA molecules to form ribonucleoprotein complexes. Huddleston, J.A., et al., *Nucleic Acids Res.* 10:1029-1039 (1982). NP is necessary for transcription and is a major determinant of host range. Huang, T.S., et al., *J. Virol.* 64:5669-5 673 (1990); Scholtissek, C., et al., *Virol.* 147:287-294 (1985). There was one difference between the *wt* 2(3) and the *ca* NP molecules, at base 113 leading to substitution of threonine for asparagine, neither of which is hydrophobic or charged. The reverse change was reported in Cox, N.J., et al., *Virol.* 167:554-567 (1988).

Although having similar protein secondary structures by Chou-Fasman and Garnier-Osguthorpe predictions, the two RNA molecules showed a distinct difference in their predicted RNA structures. In *wt* 2(3) RNA6, base 113 creates a larger unpaired loop making the molecule less stable than *ca* RNA6 (structure not shown). DNA

cloning and RNA sequencing revealed that base 113 is a mixed population of A and C in both the *wt* 2(3) and the *ca* RNA6's; however, in the *wt* 2(3) the consensus base is A and in the *ca* the consensus base is C.

The asparagine in the *wt* 2(3) virus is unique among all reported NP molecules (see Table 3), but not the threonine of the *ca* virus. The A/AA/6/60 (*wt* and *ca*) viruses are the only viruses in 54 GenBank sequences with an inserted A at base 1550 near the putative polyadenlyation signal.

RNA4 (HA) and RNA5 (NA). The sequences of ca RNA4 (HA) and ca RNA5 (NA) have not been previously reported, as neither molecule is included in ca reassortant vaccines. RNA4 encodes the hemagglutinin (HA) surface glycoprotein (562 amino acids in length), while RNA5, encodes the neuraminidase (NA) surface glycoprotein (469 amino acids in length). Two silent nucleotide differences were observed between ca RNA5 and wt 2(3) RNA5 at bases 394 and 604. Three additional differences seen at bases 144, 455, and 729 of ca RNA4 and wt 2(3) RNA4 coded for amino acid changes: asparagine to isoleucine (position 34), alanine to threonine (position 138) and lysine to threonine (position 229). The presence of clear differences in these two surface genes underscores the different passage histories of the two viruses and provides additional evidence for their separate identities.

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## **SPECIFIC EXAMPLE 2 - SEQUENCE COMPARISONS**

Sequence of Wild Type Progenitor. Table 4 presents positions for each gene where the *ca* and *wt* 2(3) viruses have unique amino acids, by comparison to previous GenBank sequences. Webster, R.G., et al., *Microbiol. Rev.* 56(1):152-179 (1992). In Table 5, a comparison to data previously published is shown and differences between the *wt* 2(3) and *ca* sequences as set forth herein, and the previously published sequences, are shown in bold type and bracketed. In positions with mixed bases, the capital letter represents the predominant base. Some of these amino acids found only in the two *ts* A/AA/6/60 viruses may be attenuating. However, many of the viruses reported in GenBank have been extensively passaged in the laboratory and will have accumulated mutations related to high relative fitness and host adaptation. Comparison to the A/AA/6/60 *wt* 28 virus previously sequenced provides further insight into attenuating lesions. Cox, N.J., et al., *Virol.* 167:554-567 (1988).

TABLE 4
Unique Amino Acid Differences between Temperature-sensitive and Attenuated wt 2(3) and ca A/AA/6/60 Viruses and Other Influenza Viruses in GenBank

Gene	No. in GenBank	Base No.	A/AA/	6/60	GenBank Viruses <sup>b</sup>
			ca/wt 2(3)	wt 28	
PB2 <sup>a</sup>	27	821	Ser	Asn	Asn
		954	Glu	Glu	Asp
PB1	23	215	His	His	Pro
		1096	Lys	Lys	Glu
		1276	Val/IIe	Val	Val
		1395	Asp	Glu	Glu
		1660	Leu	Leu	Met
PA	21	599	His	His	Arg
		2167/8	Pro	Leu	Leu
NP	54	113	Thr/Asn	Thr	Thr
		1550	Α	-	-
M1	44	453	Val	Val	Ala
		457	Leu	Leu	Phe
		678/9	Val	Val	Ala
M2	44	847	His	His	Arg
		969	Ser	Ala	Ala
NS1	73	35	Pro	Pro	Ser
		483	Thr	Ala	Glu

<sup>&</sup>lt;sup>a</sup> Five other silent differences.

<sup>&</sup>lt;sup>b</sup> Sources of GenBank viruses for each gene used in phylogenetic analysis are reported in Webster R.G., et al., *Microbiol. Rev.* 56(1):152-179 (1992).

**TABLE 5** 

# Summary of Comparative Sequence Data for A/Ann Arbor/6/60 Wild Type and Cold-Adapted Viruses

Data fr	om Stud	у				Data P Publish		sly	
wt A/AA/6/60 E2(3)		ca A/AA/6/60		ca A/AA/6/60		wt A/AA/6/60 E28			
Gene	Base No.	Base	AA	Base	AA	Base	AA	Base	AA
PB2	141 <sup>+</sup>	A/g		<u>G</u>		G		Α	"
	426	С		С		С		Т	
	714	Т		[T]		[C]		[C]	
	821	G 265	ser	G	ser	G	ser	Α	as p
	963	G		[G]		[A]		[A]	
	1182	Т		Т		Т		Α	
	1212	T		T		Т		С	
	1353	G		G		G		Т	
	1923	G		G		G		Α	
	<u>1933</u> ~	<u>T/c</u>		[C]/t		[17]		Т	
PB1	123	G		G		G		Α	
	486	Т		Т		Т		С	
	1195	G 391	glu	G	glu	G	glu	Α	lys
	1276^	<u>A/g</u> 418	<u>ile</u>	<u>G/a</u>	<u>val</u>	G	val	G	val
	1395	T 457	asp	Т	asp	Т	asp	G	glu
	1766	G 581	gly	G	gly	G	gly	Α	glu
	2005	A 661	thr	Α	thr	Α	thr	G	ala
	2019	Т		Т		Т		<u> </u>	
PA	20	С		С		С		T	
	75	G		[G]		m		[17]	
	1861	G 613	glu	G	glu	G	glu	Α	lys
	2167	C 715	pro	С	pro	С	pro	T	leu
	2168	С		С		С		Т	
HA	144	A 34	asn	T	ile				

Data fr	Data from Study						Data Previously Published <sup>a</sup>			
wt A/AA/6/60 E2(3)			ca A/AA/6/	ca A/AA/6/60		ca A/AA/6/60		6/60		
Gene	Base No.	Base	AA	Base	AA	Base	AA	Base	AA	
	455	G 138	ala	Α	thr					
	729	A 229	lys	С	thr					
NA	394	С		Т						
	604	Α		Т						
NP	113<	A/c 23	asn	C/[a]	thr	[A]	asn	С	thr	
	146	G 34	gly	G	gly	G	gly	Α	as p	
	627	С		[C]		[A]		Α		
	909	G		[G]		[C]		С		
	1550	Α		Α		Α		*		
М	969	Т	ser	Т	ser	Т	ser	G	ala	
NS	483	A 153	thr	Α	thr	Α	thr	G	ala	
	813	G		G		G		Α		

<sup>&</sup>lt;sup>a</sup> Cox, N.J., et al., *Virol.* 167: 554-567 (1988).

The distribution of the clones representing the positions with the differences between the wt 2(3) and the ca viruses are listed below:

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<sup>+</sup> wt 2(3) PB2 141 four clones A, two clones G ca PB2 141 five clones G 20

<sup>~</sup> wt 2(3) PB2 1933 four clones T, two clones C ca PB2 1933 four clones C, one clone T

wt 2(3) PB1 1276 five clones A 25 ca PB1 1276 four clones G, three clones A

 $<sup>^{&</sup>lt;}$  wt 2(3) NP 113 two clones A, one clone C ca NP 113 three clones C, one clone A

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## **SPECIFIC EXAMPLE 3 - REASSORTANT SCHEMES**

## A. TYPE A REASSORTANTS

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The following is a procedure for developing Type A 6/2 cold-adapted influenza virus vaccine (CAIV) reassortants.

Materials

*Media.* The media used in this sample were prepared using the following components: a) HBSS - 500 ml HBSS (BioWhitaker 10-508); 0.5 ml gentamicin sulfate 50 mg/ml (BioWhitaker 17-518); and adjust pH to 7.0 using 0.5N NaOH; b) 2 x Eagle's - 500 ml HBSS (BioWhitaker 10-508); 10 ml BME amino acids (GIBCO 320-1051); 10 ml BME vitamins (GIBCO 320-1040); 10 ml L-glutamine (GIBCO 320-5030); and 0.5 ml gentamicin sulfate 50 mg/ml (BioWhitaker 17-518); adjust pH to 7.0 using 0.5N NaOH; c) 0.5N NaOH - 2 g NaOH; 100 ml Type I deionized water; sterilize by autoclaving 250°C for 15 min, liquid cycle.

Inoculum. Inocula were prepared as follows: Cold-adapted Master Strain Parent (A/Ann Arbor/6/60 - 7PI) - make a 10<sup>-2</sup> dilution in 2 x Eagle's. Wild Type Parent - make a 10<sup>-1</sup> dilution in 2 x Eagle's. Combine equal volumes of the two diluted parents (1:1 dilution) and use this as the inoculum.

Cells. Use SPAFAS-derived primary chick kidney (SPF-PCK) cells grown in 16 x 125 mm tissue culture tubes on the fifth day after seeding.

20 Passages

SPF-CK1 Passage. SPF-CK1 passages were performed as follows: 1) remove growth media from ten SPF-PCK tubes; 2) wash SPF-PCK tubes with 1 ml of HBSS media; 3) inoculate with 0.3 ml of inoculum per tube; 4) adsorb at room temperature for 90 min while continuously rocking at low speed; 5) remove inoculum; 6) wash SPF-PCK tubes with 1 ml of HBSS media; 7) add 1 ml of 2 x Eagle's media and incubate at 33°C; 8) after 24 hr feed tubes with 0.3 ml of 2 x Eagle's media; and 9) observe cells daily for cytopathic effect (CPE). When CPE is >75%, pass the tubes to CK2 (usually 48-72 hr).

SPF-CK2 Passage. SPF-CK2 passages were performed as follows: 1) remove growth media from the SPF-PCK tubes; 2) wash SPF-PCK tubes with 1 ml of HBSS media; 3) serially pass the CK1 passage with 0.3 ml of inoculum per tube; 4) adsorb at room temperature for 90 min while continuously rocking at low speed; 5) remove inoculum; 6) wash SPF-PCK tubes with 1 ml of HBSS media; 7) add 0.3 ml of ferret antisera against A/AA/6/60-7PI which has been treated by the trypsin-periodate method to remove nonspecific inhibitors which has been filter sterilized (0.22μ). Use a 1:32 - 1:56 final dilution of sera (note that the treated sera is a 1:8 dilution); 8)

adsorb at room temperature for 15 min while continuously rocking at low speed; 9) add 1 ml of 2 x Eagle's media and incubate at 33°C; and 10) observe cells daily for CPE. When CPE is >75%, pass the tubes to CK3 (usually 48-72 hr).

**SPF-CK3 Passage.** The procedure for this passage was identical to the CK2 passage. When the CPE of this passage is >75%, plaque-purify the material in SPF-PCK cells.

## Plaque Purification/Genotype Screening

1PI (1st) Plaque Purification. First plaque purification and genotype screening were performed as follows: 1) serially dilute the CK3 passage in 2 x Eagle's media through a 10<sup>-4</sup> dilution, one ml of each dilution is needed per flask infected; 2) plaque the 10<sup>-3</sup> and 10<sup>-4</sup> dilution of each tube at 33°C following the procedure for plaquing in PCK cells; 3) pick several plaques for each tube. Using a sterile cotton plugged Pasteur pipet which has been bent to a 90° angle remove the agar and cells surrounding a well-isolated plaque. Draw a small volume of HBSS into the Pasteur pipet prior to picking the plaque to facilitate the expulsion of the plaque from the Pasteur pipet. Transfer the plaque material to a sterile capped tube containing 0.5 ml of 2 x Eagle's media. One plaque from each tube is passed in SPAFAS eggs and the other plaques should be frozen at -70°C as backup material; 4) pass one plaque in two SPAFAS eggs (0.2 ml of inoculum per egg) at 33°C for 72 hr. Refrigerate eggs at 4°C for at least one hr prior to harvesting the allantoic fluid. Determine the hemagglutinin titer (HA) of the egg pool to confirm the presence of virus and determine plaquing dilutions for the next purification. Two eggs will provide all the virus needed; and 5) genotype the 1PI egg material following the genotype procedure to identify potential 6/2 candidates.

**2PI (2nd) Plaque Purification.** Second plaque purification and genotype screening were performed as follows: 1) plaque the 1PI egg material in SPF-PCK cells at 33°C following the procedure for plaquing in PCK cells, using the following appropriate dilutions to obtain well-isolated plaques:

**TABLE 6** 

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HA Titers	Approximate Dilutions
< 1:32	10 <sup>-3</sup> and 10 <sup>-4</sup>
<u>&lt;</u> 1:128	10 <sup>-4</sup> and 10 <sup>-5</sup>
<u>&lt;</u> 1:512	10 <sup>-5</sup> and 10 <sup>-6</sup>
> 1:512	10 <sup>-5</sup> , 10 <sup>-6</sup> and 10 <sup>-7</sup>

2PI plaques should be derived from the same material which is genotyped since the egg passage may exert selective pressure on the plaques; and 2) pick several plaques following the procedure previously described. One plaque from each tube will be replaqued in SPF-PCK cells and the other plaques should be frozen at -70°C as backup material.

*3PI (3rd) Plaque Purification.* Third plaque purification and genotype screening were performed as follows: 1) plaque the 2PI plaques in SPF-PCK cells at 33°C following the procedure for plaquing in PCK cells. The appropriate dilutions for this passage are 10<sup>-1</sup> and 10<sup>-2</sup>; 2) pick several plaques following the procedure previously described. At this time you should know which are potential 6/2's and non-candidates can be discarded. One plaque will be amplified in SPAFAS eggs at 33°C and the others should be frozen at -70°C as backup material; 3) genotype the 6/2 candidates to confirm that 3PI passages have the 6/2 gene configuration; and 4) characterize the phenotypic profile of the 6/2 vaccine candidates at 25°C, 33°C and 39°C to confirm the presence of the *ca* and *ts* markers.

## B. TYPE B REASSORTANTS

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The following is a procedure for developing Type B 6/2 cold-adapted influenza virus vaccine (CAIV) reassortants.

## **Materials**

*Media.* The media used in part B of this example were prepared as described in part A above.

Inocula. Inocula of the ca master strain parent and wild type are diluted to 10<sup>-2</sup>

Cells. SPAFAS primary chick kidney (SPF-PCK) were grown as described in part A above.

## **Passages**

SPF-CK1 Passage. SPF-CK1 passages were performed as follows: 1) remove growth media from ten SPF-PCK tubes; 2) wash SPF-PCK tubes with 1 ml of HBSS media; 3) inoculate with 0.3 ml of inoculum per tube; 4) adsorb at room temperature for 90 min while continuously rocking at low speed; 5) remove inoculum; 6) wash SPF-PCK tubes with 1 ml of HBSS media; 7) add 0.3 ml of ferret antisera against B/AA/1/66 CL 4-1-7Pl treated by the trypsin-periodate method to remove nonspecific inhibitors and filter sterilized  $(0.22\mu)$ . Use a 1:56 final dilution of sera (note that the treated sera is a 1:8 dilution); 8) adsorb at room temperature for 15 min while continuously rocking at low speed; 9) add 1 ml of 2 x Eagle's media and incubate at 25°C; and 10) observe

cells daily for cytopathic effect (CPE). When CPE is >75%, pass the tubes to CK2 (usually 72-96 hr).

SPF-CK2 Passage. SPF-CK2 passages were performed as follows: 1) remove growth media from the SPF-PCK tubes; 2) wash SPF-PCK tubes with 1 ml of HBSS media; 3) serially pass the CK1 passage with 0.3 ml of inoculum per tube; 4) adsorb at room temperature for 90 min while continuously rocking at low speed; 5) remove inoculum; 6) wash SPF-PCK tubes with 1 ml of HBSS media; 7) add 0.3 ml of ferret antisera against B/AA/1/66 CL 4-1-7Pl treated by the trypsin-periodate method to remove nonspecific inhibitors which has been filter sterilized (0.22μ). Use a 1:56 final dilution of sera (note that the treated sera is a 1:8 dilution); 8) adsorb at room temperature for 15 min while continuously rocking at low speed; 9) add 1 ml of 2 x Eagle's media and incubate at 33°C; and 10) observe cells daily for CPE. When CPE is >75%, pass the tubes to CK3 (usually 48-72 hr).

## Plaque Purification/Genotype Screening

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1PI (1st) Plaque Purification. First plaque purifications and genotype screening were performed as follows: 1) serially dilute the CK2 passage in 2 x Eagle's media through a 10<sup>-4</sup> dilution, one ml of each dilution is needed per flask infected; 2)plague the 10<sup>-3</sup> and 10<sup>-4</sup> dilution of each tube at 33°C following the procedure for plaquing in PCK cells; 3) pick several plaques for each tube. Using a sterile, cottonplugged Pasteur pipet which has been bent to a 90° angle, remove the agar and cells surrounding a well-isolated plaque. Draw a small volume of HBSS into the Pasteur pipet prior to picking the plaque to facilitate the expulsion of the plaque from the Pasteur pipet. Transfer the plaque material to a sterile capped tube containing 0.5 ml of 2 x Eagle's media. One plaque will be passed in SPAFAS eggs and the others should be frozen at -70°C as backup material; 4) pass one plaque in two SPAFAS eggs (0.2 ml of inoculum per egg) at 33°C for 72 hr. Refrigerate eggs at 4°C for at least one hr prior to harvesting the allantoic fluid. Determine the hemagglutinin titer(HA) of the egg pool to confirm the presence of virus and determine plaquing dilutions for the next purification. Two eggs will provide all the virus needed; and 5) genotype the 1PI egg material following the genotype procedure to identify potential 6/2 candidates.

**2PI (2nd) Plaque Purification.** Second plaque purification and genotype screening were performed as follows: 1) plaque the 1PI egg material in SPF-PCK cells at 33°C following the procedure for plaquing in PCK cells. Use the appropriate dilutions to obtain well-isolated plaques, such as the following:

**TABLE 7** 

HA Titers	Approximate Dilutions
< 1:32	10 <sup>-3</sup> and 10 <sup>-4</sup>
<u>&lt;</u> 1:128	10 <sup>-4</sup> and 10 <sup>-5</sup>
<u>&lt;</u> 1:512	10 <sup>-5</sup> and 10 <sup>-6</sup>
> 1:512	10 <sup>-5</sup> , 10 <sup>-6</sup> and 10 <sup>-7</sup>

2PI plaques should be derived from the same material which is genotyped since the egg passage may exert selective pressure on the plaques; and 2) pick several plaques following the procedure previously described. One plaque will be replaqued in SPF-PCK cells and the others should be frozen at -70°C as backup material.

*3PI (3rd) Plaque Purification.* Third plaque purification and genotype screening were performed as follows: 1) plaque the 2PI plaques in SPF-PCK cells at 33°C following the procedure for plaquing in PCK cells. The appropriate dilutions for this passage are 10<sup>-1</sup> and 10<sup>-2</sup>; 2) pick several plaques following the procedure previously described. At this time you should know which are potential 6/2's and non-candidates can be discarded. One plaque will be amplified in SPAFAS eggs at 33°C and the others should be frozen at -70°C as backup material; 3) genotype the 6/2 candidates to confirm that 3PI passages have the 6/2 gene configuration; and 4) characterize the phenotypic profile of the 6/2 vaccine candidates at 25°C, 33°C and 37°C to confirm the presence of the *ca* and *ts* markers.

### C. INFLUENZA VIRUS

A number of cold-adapted reassortants and cold-adapted influenza vaccines (CAIV) have been produced and clinically tested using the general scheme set forth above with modifications known to or easily devisable by those skilled in the art without undue experimentation. In addition, the cold-adopted influenza vaccines that have proven efficacious are set forth in Table 10. The following Table sets forth the Type A and Type B reassortants:

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## TABLE 8

	CA	AIV
5	TYPE A REASSORTANT	TYPE B REASSORTANT
	A/Victoria/75 (H3N2)	B/Tecumseh/63/80
	A/Victoria/75 (H3N2)	B/Texas/1/84
	A/Swine/New Jersey/8/76/ (H1N1)	B/Ann Arbor/1/86
	A/Alaska/6/77 (H3N2)	B/Yamagata/16/88
10	A/Alaska/6/77 (H3N2)	B/Bangkok/163/90
	A/USSR/90/77 (H1N1)	B/Panama/45/90
	A/Hong Kong/77 (H1N1)	B/Panama/45/90
	A/California/10/78 (H1N1)	
	A/Alaska/6/77 (H3N2)	
15	A/Peking/2/79 (H3N2)	
	A/Washington D.C./897/80 (H3N2)	
	A/Shanghai/31/80 (H3N2)	
	A/Korea/1/82 (H3N2)	
	A/Dunedin/6/83 (H1N1)	
20	A/Bethesda/1/85 (H3N2)	
	A/Texas/1/85 (H1N1)	
	A/Kawasaki/9/86 (H1N1)	
	A/Wyoming/1/87 (H3N2)	
	A/Los Angeles/2/87 (H3N2)	
25	A/Shanghai/11/87 (H3N2)	
	A/Shanghai/16/89 (H3N2)	
	A/Guizhou/54/89 (H3N2)	
	A/Chick/Germany/N/49 (H10N7)	
	A/Equine/Miami/1/63 (H3N8)	
30	A/Beijing/352/89 (H3N2)	
	A/Yamagata/32/89 (H1N1)	
	A/Texas/36/91 (H1N1)	
	A/Beijing/352/89 (H3N2)	
	A/Los Angeles/2/87 (H3N2)	

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## SPECIFIC EXAMPLE 4 - CA INFLUENZA VIRUS REASSORTANT Vaccine Pools

Facilities. The inoculation, harvesting, pooling, and filling operations were performed in a Biohazard Laminar Flow Hood (Type A/B3). All containers and equipment utilized were sterilized within 72 hr prior to use.

**Production Substrate.** Ten-day old incubated, specific pathogen free - complement fixation avian leukosis (SPF-COFAL) negative embryonated hens' eggs from SPAFAS, Inc. (Norwich, CT) were used. Quality Control Sheets for the flocks were obtained and retained to maintain traceability of eggs.

Cold-adapted Reassortant Vaccine Donor Strain. The cold-adapted reassortant vaccine donor strain passage will vary between SPF egg passage 1 (SE1) and SE4. These passages (SE1-SE4) of the donor virus were produced as follows: The virus was thawed and diluted 1:100 to 1:10,000 (strain-dependent) in HBSS. Ten-day old embryonated eggs were inoculated via the allantoic route with 0.1 ml of the indicated diluent. All eggs were incubated at 33°C for 40-72 hr (strain-dependent) at which time they were chilled at 4°C for 1-2 hr prior to the harvesting of the allantoic fluid. This material was passed once to prepare the seed lot.

## A. Virus Seed Production

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The virus seed lot was used as the seed for the production of all vaccine pools. All work was done in production facilities.

Inoculation. The seed virus was thawed and diluted 1:100 to 1:10,000 (strain-dependent) in HBSS containing 1% of 10 x SPG (sucrose, 2.18M;  $K_2PO_4$ , 0.038M;  $K_2HPO_4$ , 0.072M; potassium glutamate, 0.049M). The ten-day old embryonated eggs were inoculated via the allantoic route with 0.1 ml of the indicated diluent. All eggs were incubated at 33°C for 40-72 hr (strain-dependent) at which time they were candled and any dead embryo was discarded. All live eggs were chilled overnight at 4°C prior to the harvesting of the allantoic fluid.

Harvest and Clarification. Allantoic fluids were harvested and pooled in approximately 180 ml amounts. The harvested allantoic fluid was incubated at 37°C (water bath) for 60 min to elute any virus adsorbed to red blood cells. Each bottle was then clarified by centrifugation at 1400 g for 15 min. 10 x SPG was added to each harvest to achieve a 10% v/v suspension for virus stabilization. The harvest bottles were pooled. Sterility assays were carried out on the pool (dual sterility tests in both fluid thioglycollate and tryptone soya broth at 33°C and 22°C.). The seed pool was assayed for hemagglutinin activity and aliquotted in the appropriate volumes needed for vaccine production.

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## B. Virus Pool Production

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Inoculation. The seed virus was thawed and diluted 1:100 to 1:10,000 (strain-dependent) in HBSS containing 1% of 10 x SPG. The ten-day old embryonated eggs were inoculated via the allantoic route with 0.1 ml of the indicated diluent. For negative controls, approximately 30 eggs were inoculated via the allantoic route with 0.1 ml of the indicated diluent. All eggs were incubated at 33°C for 40-72 hr (strain-dependent) at which time they were candled and dead embryos were discarded. All live eggs were chilled overnight at 4°C prior to the harvesting of the allantoic fluid.

Harvest and Clarification. Allantoic fluids were harvested and pooled in approximately 180 ml amounts. The harvested allantoic fluid was incubated at 37°C (water bath) for 60 min to elute any virus adsorbed to red blood cells. Each bottle (control and infected) was then clarified by centrifugation at 1400 g for 15 min. 10 x SPG was added to each harvest to achieve a 10% v/v suspension for virus stabilization. Aliquots were removed from each harvest bottle to form a sample master pool. Sterility assays were carried out on each individual bottle and on the sample master pool; dual sterility tests in both fluid thioglycollate and tryptone soya broth at 33°C and 22°C were conducted. The master pool was assayed for hemagglutinin activity and virus characterization (phenotype and genotype assays).

**Pooling, Treatment and Dispensation.** When the preliminary tests (sterility and virus characterization) proved satisfactory, the sterile harvests were thawed and pooled. Fluids were passed through sterile gauze pads to remove any membranous material that may be present. Antibiotics were added to the final pools to achieve the following concentrations: neomycin 100 mcg/ml, amphotericin B (I.V.) 5 mcg/ml.

Control Fluids: This pool was distributed into the appropriate aliquots needed for subsequent testing for adventitious agents. During dispensation the fluid was kept chilled in an ice-water bath. The fluids were stored at < -75°C in a mechanical freezer.

Virus-Infected Fluids: This pool was distributed into the appropriate aliquots needed for subsequent safety testing. The remainder of the fluid was distributed into aliquots for use as a live cold-adapted influenza virus vaccine. During dispensation the fluid was kept chilled in an ice-water bath. The fluids were stored at < -75°C in a mechanical freezer.

Tests for Adventitious Agents. The following are microbial sterility tests: 1) pre-antibiotic testing for bacteria with fluid thioglycollate at 22°C and 33°C, and tryptone soya broth media at 22°C and 33°C; and 2) post-antibiotic testing for bacteria in Lowenstein-Jensen egg medium, and for mycoplasma and brucella.

Identity in tissue culture is tested using serum-neutralization in Primary African Green Monkey Kidney (AGMK) cells.

Tissue culture tests for adventitious agents are performed using: 1) Primary African Green Monkey Kidney (AGMK) cells; 2) Primary Bovine Embryonic Kidney (BEK) cells; 3) Primary Human Amnion (PHA) cells; 4) Primary Rabbit Kidney (PRK) cells; 5) Human Diploid Fibroblast (MRC-5) cells; and 6) Human Carcinoma of the Cervix (HeLa) cells.

Animal tests for adventitious agents are performed using: 1) adult mice (ICR); 2) suckling mice (CD-1); and 3) adult guinea pigs. Guinea pig tests are conducted for *M. tuberculosis*, Q-fever and *B. abortus* antibodies.

A test for reverse transcriptase by assaying for the detection of RNA-dependent DNA-polymerase activity is also performed.

Final container/pool testing is performed by the following tests: microbial sterility is tested with fluid thioglycollate at 22°C and 33°C and fluid soybean-casein digest; COFAL testing is performed to test for avian leukosis virus; general safety testing using mice and guinea pigs; virus characterization including infectivity with TCID<sub>50</sub> in Madin-Darby Canine Kidney (MDCK) cells, plaquing efficiency with Madin-Darby Canine Kidney (MDCK) cells with Plaque Forming Unit (PFU) determination at 34, 36, 37, 38 and 39°C, and SPF derived Primary Chick Kidney (SPCK) cells with Plaque Forming Unit (pfu) determination at 25, 33 and 39°C for confirmation of phenotypic markers; antigenic analyses using hemagglutinin inhibition assay and neuraminidase inhibition assay; reactogenicity in ferrets; hemagglutinin activity; and passage level, wherein the final passage of the vaccine will vary between SPF Egg Passage 3 (SE3) and SE6.

## SPECIFIC EXAMPLE 5 - CHARACTERIZATION OF CA VACCINES

## A. CA VACCINE EVALUATION

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Production lots of cold-adapted influenza vaccines were evaluated prior to distribution to certify that they were identical to the seed strains from which they were produced. The production lots underwent three different tests to certify that they were identical to the seed strains: phenotypic evaluation, genotypic evaluation and ferret reactogenicity studies.

Phenotypic Evaluation of Cold-adapted Influenza Vaccines. Cold-adapted influenza vaccines contain two stable phenotypic markers, the cold-adapted (ca) marker and the temperature-sensitive (ts) marker. Presence of the ca marker is confirmed by comparable viral growth at 25°C and 33°C. The ts marker is confirmed by a minimum 100-fold decrease in viral growth at 39°C as compared to 33°C for the

Type A cold-adapted influenza vaccine. Viral growth is quantified as plaque-forming units/milliliter (pfu/ml) in primary chick kidney cells. Production lots are checked to certify that they have both of the phenotypic markers.

Genotypic Evaluation of Cold-adapted Influenza Vaccines. Influenza viruses are negative-stranded RNA viruses with eight unique strands of RNA, each of which corresponds to an individual gene. As described above, the cold-adapted influenza vaccine is a 6/2 reassortant which contains the six attenuated internal genes of the master strain parent with the two genes coding for the surface antigens of the wild type parent. Since the genes have different electrophoretic mobilities, they can be differentiated via polyacrylamide gel electrophoresis. Production lots are checked to certify that they have the 6/2 gene constellation of the seed strain.

Ferret Reactogenicity Studies. The ferret is the animal model of choice for testing the potential virulence of influenza viruses. The cold-adapted influenza vaccine is attenuated in ferrets and is characterized by an asymptomatic infection with viral growth restricted to the nasal turbinates. In this study, a ferret was infected with a high multiplicity of infection dose and monitored twice daily for symptoms of influenza. On day 3, the peak day for viral replication, the ferret was euthanized and the turbinate and lung were checked for viral growth. Production lots were checked to confirm that they are attenuated in the ferret model.

## 20 B. MATERIALS AND METHODS

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## **Preparation of PCK Cells**

Media and Materials. The media used in this example are prepared with the following components: a) 199 with 10% FBS - 450 ml sterile Type I deionized water; 50 ml Fetal Bovine Sera - heat inactivated; 50 ml 10 x 199 (GIBCO #330-1181); 10 ml L-glutamine (GIBCO 320-5030); 0.5 ml gentamicin sulfate (50mg/ml) (M.A. Bioproducts 17-518); and 16 ml 1.4% NaHCO<sub>3</sub>, pH to 6.8 with 0.5N NaOH. HBSS w/P&S - 500 ml HBSS (M.A. Bioproducts 10-508); and 0.5 ml gentamicin sulfate (50mg/ml) (M.A. Bioproducts 17-518); b) 0.25% trypsin - 1 L HBSS (M.A. Bioproducts 10-508); and 2.5 g trypsin 1:250 (Difco 0152-15-9). Dissolve in HBSS by stirring at room temperature, filter sterilize (0.22μ), pH to 7.6 with 0.5N NaOH after filtering; c) 0.5N NaOH - 2 g NaOH; and 100 ml Type I deionized water; sterilize by autoclaving 250°C for 15 min, liquid cycle; d) 1.4% NaHCO<sub>3</sub> - 100 ml Type I deionized water; 1.4 g NaHCO<sub>3</sub>; and 0.1 ml 4% Phenol Red. Sterilize - autoclave 250°C for 15 min, liquid cycle; e) 4% Phenol Red - 2 g Phenol Red (Difco 0203-11-2); 39 ml Type I deionized water; and 11 ml 0.5N NaOH; sterilize by autoclaving 250°C for 15 min, liquid cycle.

The following materials are also used: sterile instruments; sterile cotton balls; sterile gauze; sterile Petri dish; sterile 50 ml centrifuge tubes; ether jar and diethyl ether; dissecting boards and pins; and 70% ethanol.

Procedure. The following procedures are performed: 1) sacrifice 1 to 3-day old chicks with ether; 2) place chicks on dissecting board (backs against board) and pin the wings and feet; 3) wash chick with 70% ethanol; 4) cut away skin starting at throat to totally expose chest and abdomen using one set of sterile instruments; 5) using a second set of sterile instruments, cut along each side of the rib cage, peel down rib cage and omentum to expose internal organs; 6) with new sterile instruments cut the esophagus and trachea, peel down internal organs to expose the kidneys; 7) swab the body cavity with sterile cotton balls to remove blood; 8) with new sterile instruments remove kidneys and place in a Petri dish with HBSS; 9) with new sterile instruments remove connective tissue from the kidneys; 10) transfer kidneys to a 50 ml centrifuge tube. Keep the kidneys near the top for mincing; 11) mince the kidneys with a new set of instruments, using recurved scissors; 12) wash the kidneys three times with HBSS (10 ml per wash) and discard all washes; 13) add 5 ml of 0.25% trypsin per chick and incubate at 35°C for ten min with occasional shaking; 14) shake vigorously by hand for three minutes. (The trypsinization times can vary with the activity of each lot of trypsin used); 15) centrifuge for 10 min at 1000-1200 RPM; 16) pour off supernatant and resuspend cells in 10 ml of 199 w/ 10% FBS; 17) filter through sterile gauze into 20 ml per chick of 199 w/ 10% FBS, and dispense into culture flasks, tubes or plates and incubate at 35°C; 18) feed 100% with 199 w/ 10% FBS after 72 hr; and 19) incubate at 35°C, cells should be usable 96 hr after seeding.

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## Plaquing PCK Cells

*Media.* The following media are used: a) Kilbourne - 350 ml sterile Type I deionized water; 100 ml 10 X 199 (GIBCO #330-1181); 20 ml MEM amino acids (50X) (M.A. Bioproducts 13-606); 7.5 ml 5% NaHCO $_3$ ; 10 ml MEM vitamins (100X) (M.A. Bioproducts 13-607); 2.86 ml 35% Bovine Sera Albumin (SIGMA A-8918); and 0.5 ml gentamicin sulfate (50mg/ml) (M.A. Bioproducts 17-518) adjust pH to 7.0 using 0.5N NaOH; b) 2 x Eagle's - 500 ml HBSS (M.A. Bioproducts 10-508); 10 ml BME amino acids (GIBCO 320-1051); 10 ml BME vitamins (GIBCO 320-1040); 10 ml L-glutamine (GIBCO 320-5030); and 0.5 ml gentamicin sulfate 50 mg/ml (M.A. Bioproducts 17-518) adjust pH to 7.0 using 0.5N NaOH; c) 1% DEAE dextran - 1 g DEAE dextran (Pharmacia 17-0350-01); and 100 ml sterile Type I deionized water (Filter Sterilize (0.22μ filter)); d) 1% Neutral Red - 1 g Neutral Red (DIFCO Bacto Neutral Red 0208-13); 100 ml sterile Type I deionized water; 1) dissolve in H<sub>2</sub>0 by stirring at room

temperature for several hours; 2) filter through Whatman #1 filter paper to remove undissolved particulates; 3) aliquot into light-proof bottles and autoclave to sterilize (15 psi for 15min); and 4) store at room temperature (works best when the stain has aged; unlimited shelf life); e) HBSS - 500 ml HBSS (M.A. Bioproducts 10-508); and 0.5 ml gentamicin sulfate 50 mg/ml (M.A. Bioproducts 17-518), adjust pH to 7.0 using 0.5N NaOH); f) 0.5N NaOH - 2 g NaOH; and 100 ml Type I deionized water. Sterilize by autoclaving 250°C for 15 min, liquid cycle; g) 1.6% purified agar - 1.6 g BBL agar purified (Becton Dickison 11853); and 100 ml sterile Type I deionized water. Autoclave to sterilize and prepare while virus is adsorbing - make volume needed for overlay.

**Procedure.** The following procedure was used: 1) set up water bath to keep media and agar at proper temperature (39-41°C); 2) make serial dilutions of the virus in 2 x Eagle's (1 ml of diluted virus per 25 cm² tissue culture flask); 3) remove media from tissue culture flasks and wash once with HBSS, 2 ml per 25 cm² flask; 4) add 1 ml of diluted virus per 25 cm² flask; 5) adsorb virus at room temperature for 1 hr with gentle rocking; 6) remove virus inoculum from flask; 7) overlay with a 1:1 mixture as described below, 5 ml per 25 cm² flask (1st Overlay - see below); 8) cool bottles until agar gels at room temperature, approximately 10 min; 9) incubate at desired temperatures (Type A Influenza - Phenotype 25°, 33° and 39°C; Type B Influenza - Phenotype 25°, 33° and 37°C); 10) after appropriate incubation overlay with 1:1 mixture as described below, 4 ml per 25 cm² flask (2nd Overlay - see below);

**TABLE 9** 

Temperature	Incubation until 2nd overlay
25°C	96 hr
33°,37°,39°C	48 hr

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11) cool bottles until agar gels at room temperature, approximately 10 min; 12) incubate at desired temperature; and 13) check daily for plaques. At 33°, 37° and 39°C, all plaques should be visible within 48 hr after the second overlay. At 25°C, it can take up to 168 hr (7 days) after the second overlay for all plaques to be visible.

The 1st Overlay is prepared by a 1:1 mixture of the following media mixture with 1.6% purified agar: 100 ml Kilbourne media and 3 ml 1% DEAE dextran. The

amount of DEAE dextran needed will vary with the batch of purified agar. This concentration should work for most batches.

The 2nd Overlay - Neutral Red is prepared by a 1:1 mixture of the following media mixture with 1.6% purified agar: 100 ml Kilbourne media; 3 ml 1% DEAE dextran. The amount of DEAE dextran needed will vary with the batch of purified agar.

This concentration should work for most batches; and 2 ml 1% Neutral Red. The amount of Neutral Red needed can vary with the batch. For long-term consistency, enough Neutral Red can be made at one time to last several years.

## RNA Labelling

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Media and Solutions. The following media and solutions are used: a) HBSS -500 ml HBSS (M.A. Bioproducts 10-508); and 0.5 ml gentamicin sulfate 50 mg/ml (M.A. Bioproducts 17-518) (adjust pH to 7.0 using 0.5N NaOH); b) 2 x Eagle's -500 ml HBSS (M.A. Bioproducts 10-508); 10 ml BME amino acids (GIBCO 320-1051); 10 ml BME vitamins (GIBCO 320-1040); 10 ml L-glutamine (GIBCO 320-5030); 0.5 ml gentamicin sulfate 50 mg/ml (M.A. Bioproducts 17-518) (adjust pH to 7.0 using 0.5N NaOH); c) <sup>3</sup>H-uridine - [5,6-<sup>3</sup>H] Uridine - 1.0 mCi/ml (Amersham, Inc. TRK 410); d) 5 M NaCl - 146.1 g NaCl (Bring the volume to 500 ml with Type I deionized water); e) 1 M Tris-HCI (pH 7.4) - 60.55 g Trizma Base (Sigma T-1503); 400 ml Type I deionized water; 35 ml concentrated HCl; and 0.5 ml diethylpyrocarbonate (Sigma D-5758); Allow solution to cool to room temperature. Adjust pH to 7.4 with HCl. Bring the volume up to 500 ml with Type I deionized water. Sterilize by autoclaving 250°C for 15 min, liquid cycle; e) 0.5 M EDTA - 186.1 g disodium EDTA (Sigma ED2SS); 800 ml Type I deionized water; and 20 g NaOH. Mix and adjust the pH to 7.4 with NaOH, sterilize by autoclaving 250°C for 15 min, liquid cycle; f) 30% sucrose - 150 g sucrose (Sigma S-9378); 10 ml 5 M NaCl; 5 ml 1 M Tris-HCl, pH 7.4; and 1 ml 0.5 M disodium EDTA (ethylenediaminetetraacetic acid) (Sigma ED2SS). Bring up to 500 ml with Type I deionized water; g) STE (Sodium-Tris-EDTA) - 1 ml 0.5 M disodium EDTA (Sigma ED2SS); 10 ml 5.0 M NaCl; and 5 ml of 1 M Tris-HCl, pH 7.4 (Trizma Base) (Sigma T-1503). Add 484 ml of Type I deionized water; h) proteinase-K - proteinase-K 20 mg/ml (Beckman-340321). Dilute 100 mg in 5 ml of sterile Type I deionized water; i) SDS - sodium dodecyl sulfate (Sigma L-5750), 10% (w/v) in Type I deionized water; j) 1/10 x TBE loading buffer - 0.5 ml 10 x TBE; 0.5 ml 10% SDS; 1.0 g ficoll (Sigma F-4375); 2.5 ml glycerol (Baker 2140-03); and 0.125 g Bromophenol Blue (Bio-Rad 161-0404). Bring up to 50 ml with Type I deionized water.

**Protocol.** The following protocol is used:

Day 1: 1) Use 2 - 25 cm<sup>2</sup> flasks of primary chick kidney cells; 2) remove media and wash with HBSS, 2 ml/flask; 3) infect cells with virus - 2 ml virus diluted 1:2 in 2 x Eagle's; 4) rock cells gently for 1 hr at room temperature; 5) remove inoculum; 6) add label, use 0.2 mCi - 0.25 mCi <sup>3</sup>H-uridine/flask. Diluted in 2 x Eagle's, total volume 1.5 ml/flask; 7) place in 33°C incubator for 4 hr; 8) after 4 hr, add 3.5 ml 2 x Eagle's to each flask; and 9) incubate at 33°C for 48 hr.

Day 3: 1) Transfer fluid from the 2 flasks into a 15 ml centrifuge tube; 2) CAKRIDGE (tal. 10 centrifuge at 500 g for 15 min at 4°C; 3) pour supernatant into 30 ml Oakridge tubes; 4) underlay supernate with 7.5 ml 30% sucrose; 5) balance tubes with STE; 6) spin at 22,500 rpm for 2-1/2 hr in a Beckman type 30 rotor; 7) pour fluid from tubes into beaker (³H aqueous waste - discard); 8) let tubes sit on paper inverted for 5-10 min; (e.g., a K/MW)(FE)

9) mark pellet - dry tube with Kimwipe; 10) resuspend each pellet in 200 μl STE, place suspension in a 1.5 ml centrifuge tube; 11) add 8 μl proteinase K (0.16 mg) to each tube, mix and incubate at 37°C for 10 min; 12) add 10 μl of 10% SDS. Mix and incubate at 37°C for 10 min; and 13) add 0.65 ml of 95% EtOH. Mix and place at -20°C overnight.

Day 4: 1) Pellet the RNA in a microcentrifuge for 15 min at 4°C; 2) empty EtOH SPEED VAC-TYPE into beaker - drain tubes upside down for several min; 3) dry the tubes in a Speedvac concentrator for approximately 10-20 min; 4) resuspend pellet in 32 μl of 1/10 x TBE loading buffer; 5) heat at 56°C for 2-3 min; 6) remove 2 μl sample and mix with 2 ml of liquid scintillation fluid; 7) count on Channel 1 for 0.5 min in liquid scintillation counter to get CPM (counts per min); 8) freeze sample until used at -70°C; 9) heat at 56°C for 2-3 min before loading; and 10) load 150,000 - 200,000 CPM.

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## Mixed Agarose-PAGE

Reagents. The following reagents were employed: a) 30% acrylamide, 1.5% bis-acrylamide - 30 g acrylamide (Bio-Rad 115009B); and 1.5 g bis-acrylamide (Bio-Rad 41936B). Bring up to 100 ml with Type I deionized water; b) 10 x TBE Buffer - 54 g Trizma Base (0.89 M) (Sigma T-1503); 27.5 g boric acid (0.89 M) (Mallinckrodt CAS10043-35-3); 4.65 g EDTA disodium salt (20 mM); (ethylenediaminetetraacetic acid) (Sigma ED2SS). Bring up to 500 ml with Type I deionized water; c) 10% w/v SDS - 10 g sodium dodecyl sulfate (Sigma L-5750). Bring up to 100 ml with Type I deionized water; d) diethylpyrocarbonate - diethyl pyrocarbonate 50 ml in 100 ml deionized water (Sigma D-5758); e) 1 x TBE running buffer - 216 g Trizma Base (89 mM) (Sigma T-1503); 110g boric acid (0.89 M) (Mallinckrodt CAS10043-35-3); 18.6 g EDTA disodium salt (20 mM) (Sigma ED2SS); (ethylenediaminetetraacetic acid); and 20 g sodium dodecyl sulfate (SDS) (0.1%) (Sigma L-5750). Bring up to 20 liters with Type I deionized water and mix well; f) 10% ammonium persulfate - 0.3 g ammonium persulfate (Bio-Rad M3992); bring up to 3.0 ml. Stable for 7 days at 4°C; g) TEMED tetramethylethylenediamine (Bio-Rad 161-0801); h) agarose - Type V - high gelling temperature (SIGMA A-3768); i) salicylic acid - 0.3 g salicylic acid (Sigma S-3007); 36 g hexadecyltrimethylammonium bromide (Sigma H-5882); and 300 ml Type I deionized water.

*Procedure.* The following procedure is used for mixed acrylamide/agarose gel (3.0% acrylamide/0.6% agarose): Note that for proper polymerization of the gel, it must be at 56°C for 20 min after pouring. The standard procedure is to place the plates vertically in a 56°C water bath such that the water is within 1 inch of the plate tops 1) Combine and mix for 15 min: 0.6 g agarose Type V high gelling temperature, 92 ml Type I deionized water, and 50  $\mu$ I diethylpyrocarbonate; 2) boil until volume is below 79 ml; 3) measure in graduated cylinder, bring volume to 79 ml with sterile Type I deionized water, allow to cool slightly; 4) add: 10 ml of 10 x TBE, 10 ml of 30% Acrylamide/1.5% bis acrylamide, 1 ml of 10% SDS, 0.3 ml of 10% ammonium persulfate; and 30  $\mu$ I TEMED; and 5) gently mix and pour the gel immediately. After the gels have polymerized (20 min at 56°C), they are stored overnight in running buffer prior to use.

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The gels are run at a constant temperature in a circulating buffer system. Since the gels are run for extended periods (17 to 21 hr) the circulation of the running buffer is critical. The gels are run at temperatures ranging from 26°C to 40°C, and at either 230 or 240 volts (constant voltage) for 17 to 24 hr. The following are general guidelines for genotyping cold-adapted influenza vaccines: Type A: 30°C and 37°C (two gels run) at 230 volts for 17 hr. Type B: 26°C and 36°C (two gels run) at 240 volts for 21 hr.

After gels are run they are enhanced in salicylic acid for 45 min and then dried. The dried gels are placed in cassettes with X-ray film at -70°C and exposed for 24 to 72 hr. The film is developed and genotypes are read.

## **Ferret Reactogenicity Testing**

Media and Materials. The following media and materials are used: a) 2 x Eagle's - 500 ml HBSS (M.A. Bioproducts 10-508); 10 ml BME amino acids (GIBCO 320-1051); 10 ml BME vitamins (GIBCO 320-1040); 10 ml L-glutamine (GIBCO 320-5030); and 0.5 ml gentamicin sulfate 50 mg/ml (M.A. Bioproducts 17-518) adjust pH to 7.0 using 0.5N NaOH. b) sodium pentobarbital - sodium pentobarbital injection (65 mg/ml) Anthony Products Co.; c) alundum - 60 mesh norton alundum "RR" (Fisher Scientific Co. A-620); sterilize by autoclaving at 250°C for 15 min, dry cycle. Ferrets - 8 to 10-week old ferrets, male, castrated, and vaccinated against distemper (Marshall Research Animals). If the ferrets are not barrier-raised, they may have had an influenza infection during the influenza season. The animals will thus need to be treated with Penicillin G (30,000 units/day) for 7 days prior to use. (Durapen TM combination antibiotic (Vedco); and Penicillin G Benzathine and Penicillin G Procaine, 300,000 units/ml.) Miscellaneous - sterile instruments; sterile scalpel; diethyl ether for

Lysol-type disinfectant - 36 -

anesthesia; Neel; sterile Petri dishes; sterile mortar and pestle; and digital thermometer Model 8110-20 (Cole Parmer Instrument Company).

**Protocol.** The following protocol is used:

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Day 1: 1) Dilute the stock virus 10<sup>-1</sup> in 2 x Eagle's; 2) lightly anesthetize the ferret with diethyl ether. Inoculate ferret intranasally with 1 ml of the 10<sup>-1</sup> dilution of stock virus (0.5 ml in each nostril); 3) determine the EID<sub>50</sub>/ml (Egg Infectious Dose-50%/ml) titer of the inoculum; serially dilute the inoculum in 2 x Eagle's; inoculate 9-11 day old embryonated chicken eggs with dilutions 10<sup>-5</sup> through 10<sup>-8</sup>, four eggs per dilution (0.1 ml per egg); incubate the eggs at 33°C to 35°C for 72 hr; after 72 hr cool the eggs for several hr at 4°C; remove 1 ml of allantoic fluid from each egg and place in individual Kahn tubes; add 0.5 ml of 0.5% chicken red blood cells to each tube and mix; allow the blood to precipitate for 45 min and determine which tubes are positive for hemagglutinin activity. Calculate the EID<sub>50</sub> titer using the Reed-Meunch method; and 4) take rectal temperatures twice a day for 3 days.

Day 3: 1) The ferret is euthanized via heart puncture with sodium pentobarbital (130 mg/ferret); 2) place ferret on its back and clamp feet to immobilize; 3) wash abdomen with Lysol®; 4) using sterile forceps and scalpel make a 4-5 inch incision lengthwise down the sternum and pull skin back; 5) with new set of sterile forceps and scissors cut the ribs to make an opening large enough to remove the left lower lobe of lung; remove and place in a sterile Petri dish; 6) cut a section of the left lobe into small pieces and place into a freezable storage tube; 7) turn ferret over and wash head with Lysol®; 8) with scalpel and forceps remove the skin from the end of nose to below eyes; 9) cut off snout at the base of the septum; 10) cut the nasal bone on both sides of the septum - approximately 1/8 to 1/4 inch with sterile bone cutter; 11) scrape out the turbinate using sterile currette and place in freezable storage tube; 12) weigh the tubes containing the lung and turbinate samples and record; 13) place the tissues in sterile mortars and weigh the empty tube. The difference in the weight is the weight of tissues; 14) add sterile alundum to the mortars and grind (homogenize) the tissues with a sterile pestle; 15) dilute tissue with 2 x Eagle's to make 10% weight/volume suspension; 16) centrifuge the homogenate at 500 x g for 10 min at 4°C; 17) remove supernatant and freeze at -70°C; 18) thaw the supernatant and determine the EID<sub>50</sub>/ml as previously described. A general range for inoculation is: 3-day turbinate dilutions of 10<sup>-3</sup> to 10<sup>-6</sup> dilution, 3-day lung dilutions of 10<sup>-1</sup> to 10<sup>-4</sup> dilution; and 19) harvest the eggs from the inoculum and calculate the EID<sub>50</sub> as described previously.

Day 6: 1) Harvest the eggs from the 3-day turbinate and lung and calculate the EID<sub>50</sub>'s as described previously.

**Ferret Serum Collection** 

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Materials. The following materials are used: B-D Vacutainer brand Winged Collection Set, 19 gauge needle, with luer; adapter and 12-inch tubing (B-D #4919); B-D Vacutainer brand needle holder for 16mm tube (B-D #364888); B-D Vacutainer brand SST (Serum Separation Tube) 16 x 125mm (B-D #6512); diethyl ether for anesthesia; and 70% ethanol.

Procedure. The following protocol is employed: 1) assemble collection set and needle holder; 2) lightly anesthetize the ferret with diethyl ether; 3) place ferret on its back and hold firmly; 4) wash chest with 70% ethanol; 5) palpate for heartbeat (left side, between 3rd and 4th rib from base of sternum; 6) insert needle into ferret's heart; when blood is seen entering the collection tube insert the SST tube onto needle; collect the desired amount for test procedures; 3-4 ml of blood will provide 1-2 ml of serum; 7) allow blood to completely clot at room temperature (approx. 30 min); 8) centrifuge tube at room temperature for 10 min at 1000 - 1300 g; and 9) collect serum, aliquot, and store at -70°C. Note that ferret serum should be treated using the trypsin-periodate method described below to remove nonspecific inhibitors prior to use.

#### **Trypsin-Periodate Treatment for Ferret Sera**

Materials. The following materials are used: a) phosphate buffer for trypsin; Solution A consists of NaH<sub>2</sub>PO<sub>4</sub>•H<sub>2</sub>O (MW 138.01); 6.99 g NaH<sub>2</sub>PO4•H<sub>2</sub>O; and 500 ml sterile Type I deionized water. Solution B consists of Na<sub>2</sub>HPO<sub>4</sub> (MW 141.97); 7.1 g Na<sub>2</sub>HPO<sub>4</sub>; and 500 ml sterile Type I deionized water; b) working buffer consists of 1 volume of Solution A + 31 volumes of Solution B (pH=8.2); c) Trypsin solution - 0.4g trypsin 1:250 (DIFCO 0152-13-1); and 100 ml phosphate buffer. Solution is stable when frozen at -20°C; d) potassium periodate solution - 0.255 g KIO<sub>4</sub> (MW 230.02); and 100 ml sterile Type I deionized water. Store in a light-proof bottle. Stable at room temperature for one month; e) 1% glycerol saline - 1 ml glycerol; and 99 ml phosphate buffered saline (PBS) (M.A. Bioproducts 17-516).

Sera Treatment - 1) mix 1 volume of serum + 1 volume of trypsin solution; 2) heat immediately to 56°C for 30 min; 3) cool to room temperature; 4) add 3 volumes of potassium periodate solution; 5) mix and incubate at room temperature for 15 min; and 6) add 3 volumes of 1% glycerol saline; serum is a 1:8 dilution and is ready to use for HI tests. If serum is going to be used for making reassortants it needs to be filter sterilized through a 0.22µ filter (low protein binding).

#### Hemagglutinin Inhibition Screening of Ferret Sera

**Procedure.** The ferrets are screened prior to use to certify that they are immunologically naive to influenza virus. Follow the hemagglutinin inhibition procedure as described in: "Concepts and Procedures for Laboratory-Based Influenza Surveillance", U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control (July 1982).

Ferrets are screened for exposure to influenza strains which have circulated in the last 12 months and/or strains which are presently circulating. The ferret sera should always be screened against a Type A H1N1 strain, a Type A H3N2 strain, and the most recent Type B strain.

#### **SPECIFIC EXAMPLE 6 - CLINICAL RESULTS**

Since 1976 the clinical development of the cold-adapted influenza virus vaccines has included the testing of multiple reassortant vaccines in over 20,000 people between the ages of 4 months to over 80 years. A summary of the cold-adapted influenza vaccines tested in the United States is set forth in Table 10. These studies have consistently demonstrated the *ca* vaccines to be genetically stable, and non-transmissible in all populations tested. More recently, studies on the *ca* vaccine have focused in three broad areas: 1) evaluating the range and extent of the immunologic response; 2) determining the protective efficacy of the vaccine in the overall population as well as in targeted subsets; and 3) evaluating the immunologic and efficacious consequences of administrating divalent/trivalent *ca* influenza virus vaccines.

The following is a standard procedure for the clinical evaluation of and collection of specimens from volunteers in attenuated influenza vaccine studies.

#### 25 A. Clinical Observations

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Two observers should independently evaluate the clinical status of the volunteer. Optimally, each evaluator should see the patient daily before and during the first four days after virus administration.

Categories of Illness. 1) Fever - Oral temperature of greater than 37.7°C (100°F) will be considered a febrile reaction. Any temperature should be confirmed using a second thermometer, 5 minutes after the first measurement. If positive, measurement should be repeated every four hours. 2) Systemic Illness - Occurrence of myalgias, and/or chills and sweats are required for the assignment of systemic illness to a volunteer. Additional information should be gathered concerning feverishness, malaise, headache, anorexia, etc. It is recognized that these observations are subjective. 3) Pharyngitis - Sore, painful throat observed in 2

consecutive days. All volunteers reporting this symptom should receive appropriate bacterial diagnostic workups. 4) Rhinitis - Occurrence of rhinorrhea on two consecutive days. Presence of nasal obstruction and sneezing are supporting of this illness designation. 5) Lower Respiratory Tract Illness - A symptom complex consisting of substernal pain, cough (paroxysmal), sputum production.

Administration of Virus to Volunteers. An appropriate therapeutic dose, *i.e.* 0.25 ml, is administered to each nostril of a supine volunteer who should remain supine for at least ten minutes. Preferably the vaccine should be administered to all volunteers by the same individual.

# 10 B. Clinical Specimens

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1) For virus isolation, nasal wash (NW) consisting of 5 ml of veal infusion broth, containing no antibiotics, is administered to each nostril. 0.25 ml of this wash should be inoculated into each of 4 tubes of an appropriate tissue culture (RMK or MDCK). The remaining NW should be divided into three aliquots and stored at -70C. 2) At least 20 ml of blood should be collected before immunization and at 21 to 28 days after immunization. An alternative method is the use of a nasopharyngeal swab and 2 ml of veal infusion broth with antibiotics for viral isolation. 3) Nasal wash for local antibody determination - 5 ml of a physiologic salt solution is instilled into each nostril and collected. A second specimen is collected at least 30 minutes later. These two collections are pooled. The timing of the pre- and post- immunization collections is the same as for serum. The specimens should be concentrated approximately 10 fold.

#### C. Determination of Serum and Nasal Wash Antibody Levels

The tests and antigens for screening the volunteers and evaluating serum and nasal wash antibodies is as follows: Screening of volunteers - All volunteers should be HI and NI negative to the influenza subtypes that are being evaluated in the study. The antigens to be used are the A/Denver/57 and A/USSR/90/77 (Parke Davis vaccine). NI antibody determinations are performed on the specimens.

TABLE 10
Summary of Cold-adapted (ca) Influenza Vaccines
Tested in the United States

			Results		
ca Vaccine		Attenuated	Antigenic	Genetic Stability	Efficacy
B/Hong Kong/73, CR-7	Adults	+	+	+	+
	Children	QN	QN	Q	QN ·
A/Victoria/75, (H3N2) CR-22	Adults	+	+	+	+
	Children	+	+	+	+
A/Alaska/77, (H3N2) CR-29	Adults	+	+	+	+
	Children	+	+	+	+
A/Hong Kong/77, (H1N1) CR-35	Adults	+	+	+	#
	Children	+	+	+	++
A/California/78, (H1N1) CR-37	Adults	+	+	+	+
	Children	+	+	+	+
A/Washington/80, (H3N2) CR-48	Adults	+	+	+	+
	Children	+	+	+	+
A/Korea/82, (H3N2) CR-59	Adults	+	+	+	#
	Children	+	+	+	#
A/Dunedin/83, (H1N1) CR-64	Adults	+	+	+	++
	Children	+	+	+	Q
B/Texas/84, CRB-87	Adults	+	+	+	+
	Children	+	+	+	Q
A/Bethesda/85, (H3N2) CR-90	Adults	+	+	+	+
	Children	+	+	+	+

			Results		
ca Vaccine		Attenuated	Antigenic	Genetic Stability	Efficacy
A/Texas/85, (H1N1) CR-98	Adults	+	+	+	+
	Children	+	+	+	+
A/Kawasaki/86, (H1N1) CR-125	Adults	+	+	+	+
	Children	+	+	+	+
B/Ann Arbor/86, CRB-117	Adults	+	+	+	2
	Children	+	+	+	2
A/Los Angeles/87, (H3N2) CR-149	Adults	+	+	+	+
	Children	+	+	+	+
B/Yamagata/88	Adults	+	+	+	2
	Children	Q	2	Q	Q

ND = not done

#### **SPECIFIC EXAMPLE 7 -**

#### SIMULTANEOUS ADMINISTRATION WITH OTHER VACCINES

One of the pressing needs for the development of the ca vaccine is to determine if protective immunogenicity is compromised when a bivalent or trivalent preparation is administered, and if so, if this interference can be overcome. Previous studies comparing monovalent and bivalent ca A vaccine (H1N1 and H3N2) administration in seronegative children demonstrated that the frequency of seroconversion was higher when vaccines were administered individually rather than simultaneously. Wright, P.F. et al., J. Infect. Dis. 146:71-79 (1982); Wright, P.F. et al., Vaccine 3:305-308 (1985). Using simultaneous administration of 10<sup>5</sup> tissue culture infectious doses (TCID<sub>50</sub>) of each of three ca vaccines (H1N1, H3N2 and B), (less than 10 human infectious doses {HID<sub>50</sub>}/vaccine component) Belshe and coworkers evaluated the question of trivalent vaccine interference in infants. Belshe, R.B. et al., J. Infect. Dis. 165:727-732 (1992). Among the seropositive children, few children shed vaccine virus and few increases in antibody to any of the three vaccine components was observed. Within the triply seronegative infant group, 47% shed all three ca vaccine viruses and 75% of these infants had a significant antibody rise to all three ca vaccine components. Of those that showed either shedding or antibody rise to two of the three ca vaccine components, no strain pair preference was observed. These results suggest that in infants and children not previously exposed to influenza, it may be possible to identify an appropriate dose (e.g. 100 HID<sub>50</sub>/vaccine component) which could stimulate antibody response to all three components.

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The question of serological and/or protective interference in the adult population has been raised in relationship to the bivalent *ca* A vaccine efficacy studies. Edwards, K.M. et al. "A Randomized Controlled Trial of Cold-Adapted and Inactivated Vaccines for the Prevention of Influenza A Disease" (submitted for publication); Clover, R.D. et al., *J. Infect. Dis.* 163:300-304 (1991). Trivalent vaccine administration has recently been evaluated in adults having low antibody levels to all three components. In the adult population significant interference with virus shedding and a trend toward lower antibody responses, particularly against the *ca* B vaccine component, was observed in vaccinees receiving the trivalent *ca* vaccine when compared to either a bivalent A or monovalent B controls. Keitel, W.A. et al., "Trivalent Live Cold-adapted Influenza Virus Vaccine: Evidence for Virus Interference in Susceptible Adults." Manuscript in preparation). These results suggest that appropriate formulation may need to be developed to enhance the maximal response

of each influenza vaccine component. Thus, the present invention contemplates the use of such appropriate formulations which may be made by those skilled in the art.

#### SPECIFIC EXAMPLE 8 - OTHER GENETICALLY-ENGINEERED VACCINES

More recent techniques, such as recombinant DNA cloning and the transfection of in vitro mutagenized gene segments can be employed for the production of live virus vaccines. For example, the gene coding for the HA protein has been cloned into vaccinia virus and is expressed on the virus surface. Attenuated recombinant vaccinia viruses have been shown to provide protection to homologous wt virus challenge in hamsters. Smith, G.L. et al., PNAS (USA) 80:7155-7159 (1983). If necessary, other influenza genes cloned into the vaccinia virus carrier are also employed at the same time. Alternatively, master strains are comprised of a number of selected genes with specific mutations, including deletions to confer stability. Chanock, R.M. et al., Prospects for Stabilization of Attenuation in 'The Molecular Virology and Epidemiology of Influenza", Stuart-Harris et al. (eds.) Academic Press, NY (1984). CR43-3 virus is a cold reassortant whose genome contains an NS gene with a deletion in the NS1 protein coding region and is restricted for growth in both Madin-Darby canine kidney cells and in ferrets. Buonagurio, D.A. et al., J. Virol. 49:418-425 (1984); Maassab, H.F. et al., Virology 130:342-350 (1983). Because the remaining non-(HA and NA) genes are derived from the ca master strain A/Ann Arbor/6/60 virus, CR43-3 may have the potential to be used as a new master strain.

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Deletions are also generated through site specific mutagenesis in recombinant cDNA clones. The ability to introduce RNA transcripts of specifically mutagenized cDNA clones into the influenza viruses as stable parts of the genome has opened new areas of research into vaccine development. Enami, M. et al., *J. Virol.* 65:2711-2713 (1991); Enami, M. et al., *PNAS (USA)* 87:3802-3805 (1990). It is now thus possible to produce "tailor-made" influenza vaccines engineered for specific purposes in accordance with the principles of the present invention.

In particular, the *ca* A/Leningrad/47 virus is used as a model for the introduction of mutations. Klimov, A.I. et al., *Virol.* 186:795-797 (1992). The *ca* A/Leningrad/47 virus has been chosen as a model because 1) differences between the *wt* A/Leningrad, A/Leningrad/17, and A/Leningrad/47 viruses are published knowledge and they are one of the few H2N2 viruses sequenced and listed in GenBank; 2) these differences will not be lethal mutations; 3) these differences probably will not interfere with growth; 4) one or several of them may introduce another temperature sensitive (*ts*) lesion into the *ca* A/AA/6/60 genome. Since the PA, M, and NS genes of the ca and the *wt* 2(3) A/AA/6/60 viruses are identical, those three

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genes have been targeted for mutation. The *ca* A/Leningrad/47 PA gene has three differences from the *wt* A/Leningrad virus; the M gene has two differences and a *ts* lesion; and the NS gene has one difference and a *ts* lesion. The *ca* A/AA/6/60 virus has the nucleotides at these positions of the *wt* A/Leningrad virus, with the exception of 969 in the matrix gene. Because a helper virus is available which will facilitate the selection of clones bearing a mutated NS gene, that gene is mutated first and rescued using the techniques of reverse genetics known to those in the art. Nucleotide 798 of the *ca* NS gene will be mutated from guanine to adenine, coding for methionine to isoleucine in NS2. Although this nucleotide has not been definitively identified as responsible for the *ts* lesion residing on the NS gene of *ca* Leningrad, it is the only difference from the *wt* Leningrad sequence. After the mutation has been successfully rescued, the mutated *ca* A/AA/6/60 virus is evaluated for the retention of the *ca* and *ts* markers and for retention of antigenicity, as described above.

#### **SPECIFIC EXAMPLE 9 - VIRAL VECTORS**

The viruses of the present invention are also useful as vectors for foreign proteins. For example, the use of either the HA or NA genes as vectors for foreign viral proteins has been suggested. Li, S. et al. *J. Virol.* 66(1):399-404 (1992) and Castrucci, M.A. et al., *J. Virol.* 67(2):759-764 (1993). H3N2 amino acids and H2N2 amino acids were introduced into the HA of an H1N1 virus, thus constructing a chimeric HA influenza molecule. Li, S. et al., *J. Virol.* 66(1):399-404 (1992). Although foreign viral amino acids or additional amino acids were not introduced into the HA, a chimeric HA can be constructed with antigenic sites important for the current H1N1 and current H3N2 viruses in the same virus. Thus, one virus with a chimeric HA could be given instead of giving a divalent vaccine.

It has been shown that insertion of 28 amino acids into the neuraminidase stalk does not interfere with growth of the virus in eggs; in fact, the longer the stalk, the better it grew. This suggests use of the influenza virus as a vaccine vector to immunize against other unrelated infectious agents. Since the NA is a glycoprotein on the surface of the virus and is one of the two major antigenic proteins for the influenza virus, it may be an excellent site for presentation of a foreign antigenic epitope. Likewise, the *ca* A/AA/6/60 virus may also be used as a vaccine vector, Castrucci, M.A. et al., Abstract 15-4; ASV 12th Annual Meeting, July 10-14 (1993), *i.e.* a vector for the human immunodeficiency virus, HIV.

# SPECIFIC EXAMPLE 10 - CLINICAL STUDIES

As previously stated, many clinical studies have been performed using coldadapted vaccines. In this study, a live attenuated trivalent combination of vaccines

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was evaluated to see if a single intranasal administration of  $\leq$  10 TCID<sub>50</sub> of each vaccine virus could successfully immunize triply seronegative children. A detailed description of this study is also set forth in Belshe, R.B. et al., *J. Infect. Dis.* 165:727-732 (1992).

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Materials and Methods. The cold-recombinant (CR) influenza A vaccines and the CR influenza B vaccine included in the trivalent vaccine were derived from coldadapted parent strains of influenza using methods previously described. Maassab, H.F., J. Immunol. 102:728-732 (1969); Cox, N.J. et al., Virol. 97:190-194 (1979); Maassab, H.F. et al., Virol. 130:342-350 (1983); Maassab, H.F. et al., J. Infect. Dis. 146:780-790 (1982); Donabedian, A.M. et al., Microb. Pathog. 3:97-108 (1987). Influenza A/Kawasaki/9/86 (H1N1) and influenza A/Korea/1/82 (H3N2) were derived from the cold-adapted influenza A/Ann Arbor/6/60 parent virus, while influenza B/Texas/1/84 was produced from influenza B/Ann Arbor/1/66 cold-adapted parent virus. The vaccine viruses, designated CR125 (H1N1), CR59 (H3N2), and CRB-87, possessed the six internal genes of their parent cold-adapted virus, A/Ann Arbor/6/60 or B/Ann Arbor/1/66, and the hemagglutinin and neuraminidase genes of their respective wild type strains. Vaccinees received 0.5 ml of the cold-adapted trivalent influenza vaccine consisting of a mixture of CR125 and CRB-87, each diluted 1:100, and CR59 diluted 1:50. To ensure that an equal titer of each viral strain was incorporated into the trivalent vaccine, each of the three vaccines was diluted separately on the day of vaccination. Subsequently, an equal volume of each was pooled to make the vaccine for administration to the volunteers. Assays were done on an aliquot of each component of the trivalent vaccine to assess the titer of each of the influenza strains incorporated into the vaccine. Titering of vaccine on each of six vaccination dates revealed H1 vaccine to contain a mean of  $10^{5.0}~{\rm TCID_{50}},~{\rm H3}$ vaccine to contain a mean of 10<sup>4.9</sup> TCID<sub>50</sub>, and B vaccine to contain a mean of 10<sup>5.5</sup>  $\mathsf{TCID}_{50}$  per half mil of a vaccine stock before being combined into trivalent vaccine. Thus the final concentration was one-third of the above (H1, 10<sup>4.5</sup>; H3, 10<sup>4.4</sup>; and B, 10<sup>5.0</sup> TCID<sub>50</sub>/0.5-ml dose of vaccine).

Vaccination and Clinical Observations. Healthy infants and children aged 6 months to 13 years were recruited to join the study. Volunteers were randomized to receive vaccine or vaccine diluent as placebo in a double-blinded way. One of every three to four children received placebo.

Children were placed in a supine position and 0.5 ml of vaccine was instilled into the nose as previously described. Belshe, R.B. et al. *J. Infect. Dis.* 149:735-740 (1984); Anderson, E.L. et al., *J. Clin. Microbiol.* 27: 909-914 (1989). After vaccination,

the children were observed in their homes for 11 days by the vaccine center nursing staff with daily sampling by nasopharyngeal swabbing for isolation of influenza virus. Serum for antibody determinations was obtained on days 0 and 28-31. One post-vaccine serum sample was obtained on day 60.

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Potential adverse reactions were defined as: (1) fever, rectal temperature > 38.3°C (infants and young children) or oral temperature > 37.8°C (older children); (2) cough, two or more episodes noted during examination visits on 2 consecutive days; (3) rhinorrhea, fluid or mucus exiting nostrils on 2 consecutive days; (4) wheeze, sustained musical sound during expiration and confirmed by a physician investigator; (5) otitis media, red, immovable ear drum diagnosed by a physician using pneumootoscopy; (6) rhonchi, continuous low-pitched sound heard by auscultation of lung fields; (7) rales, discontinuous, interrupted explosive sounds, fine or coarse crackles heard by auscultation of lung fields and confirmed by a physician; and (8) pneumonia, a new alveolar consolidation seen radiographically.

Laboratory Studies. Serologic tests for antibody to each vaccine strain were assayed by hemagglutination inhibition (HAI) and ELISA. HAI assays used homologous, tissue-culture-grown antigen for each of the vaccine strains in the trivalent vaccine as previously described. World Health Organization, "The hemagglutination inhibition test for influenza virus." U.S. Department of Health, Education and Welfare Procedure Manual, Atlanta: Center for Disease Control (1975). Prevaccination immune status of the vaccinees was based on HAI titers; a titer <1:4 was considered seronegative. Purified hemagglutinin from heterologous influenza strains, consisting of influenza Taiwan (A/H1N1), influenza Shanghai (A/H3N2), and influenza B/Yamagata (Connaught Laboratories, Swiftwater, PA), was used for the ELISA. Briefly, microtiter plates (Dynatech, Chantilly, VA) were coated with antigen (1 μg/ml) overnight at 4°C. The remaining steps of the ELISA procedure were done the next day as follows: (1) antigen was removed but the plates were not washed; (2) plates were blocked with 0.1% bovine serum albumin in PBS and washed with PBS-Tween; (3) four-fold dilutions of test samples were added to the plates and the plates were incubated at 37°C for 2 hr; (4) after plates were washed with PBS-Tween, goat anti-human IgG was added for a 2 hr incubation at 37°C; and (5) plates were washed, developed using a phosphatase substrate kit (Kirkegaard & Perry, Gaithersburg, MD), and read in a microtiter plate reader after 30 min for IgG and 90 min for IgA. An antibody response was defined as a seroconversion by HAI or ELISA (<1:4 to ≥1:8 by HAI; <1:20 to  $\ge 1:20$  by ELISA) or as a four-fold increase in titer.

Viral shedding was monitored by isolation in cell-culture tubes of primary rhesus monkey kidney (RhMK) cells as previously described. Belshe, R.B. et al., *J. Infect. Dis.* 150:834-840 (1984). Cell cultures were incubated at 32°C for 14 days. Hemadsorption of monolayers with 0.4% guinea pig erythrocytes was done on days 5, 9 and 14. In addition, some specimens were inoculated into RhMK tubes containing combinations of polyvalent antiserum specific for two of the three subtypes to permit selective growth of the third subtype. Viral subtype was identified by HAI or by indirect immunofluorescence using monoclonal antibodies (see below). Harmon, N.W. et al., *Influenza Viruses* in "Diagnostic Procedures for Viral Rickettsial and Chlamydial Infections." Schmidt, N.J. et al. (eds.) Washington, D.C.: American Public Health Association 651-653 (1989); Riggs, R.L., *Immunofluorescence Staining* in "Diagnostic Procedures for Viral Rickettsial and Chlamydial Infections." Schmidt, N.J. et al. (eds.) Washington, D.C.: American Public Health Association 651-653 (1989).

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To enumerate the viral subtypes shed by each vaccinee, plaque assays were done using subtype-specific monoclonal antibodies in an immunoperoxidase-staining procedure. Confluent monolayers of RhMK cells in 24-well plates were rinsed with sterile PBS, pH 7.2, and then infected in triplicate with 0.2 ml/well of specimen. After absorption for 1 h at 33°C, each well was overlaid with L-15 medium (Whittaker M.A. Bioproducts, Walkersville, MD) containing 1% agarose (SeaKem; FMC Bioproducts, Rockland, ME), 200 mM L-glutamine (Whittaker M.A. Products), and 50 µg/ml gentamicin. Infected plates were incubated at 33°C for 3 days. Subsequently, plates were fixed, the agarose overlay was removed, and the plates were stained by a modification of an immunoperoxidase procedure developed by William Gruber (Department of Pediatrics, Vanderbilt Unitersity, Nashville, TN). Infected monolayers were first fixed sequentially with 80% and 100% methanol for 15 min at 4°C, and then were overlaid with 5% skim milk (Difco, Detroit) in PBS for 30 min at 37°C. After removal of the skim milk, each well was overlaid with 0.2 ml of subtype-specific monoclonal antibody diluted 1:2000 (v/v, in PBS for 1 hr at 37°C. Monoclonal antibodies designated as (B/AA/1/86 [B/AA]1/2; A/Mem/2/85 [H3 M2-7]; A/Baylor/11515/82 [H1 AB/28] were provided by Robert Webster, St. Jude Children's Research Hospital (Memphis). After two washes with 5% skim milk, 0.2 ml of peroxidase-conjugated rabbit anti-mouse antibody (1:35, Dako, Carpinteria, CA) was added to each well for 30 min at 37°C. Plates were washed twice with 5% skim milk after which each well was overlaid with 0.2 ml of peroxidase-conjugated swine antirabbit antibody (1:90; Dako) for 30 min at 37°C. After two 5% skim milk washes, each well was overlaid with 0.2 ml of AEC substrate (Dako) prepared according to manufacturer's instructions. Plates were incubated at room temperature until positive control wells showed satisfactory color development (~5 min.). Plates were washed with distilled water and read under a dissecting microscope for the presence of red-stained plaques. Uninfected wells were stained in parallel to control for background staining.

**Results.** The clinical and serologic response of vaccinees is summarized in Table 11. As in other trials, some background mild respiratory illness was seen in both vaccinees and controls and was more frequent among children < 12 months old. There was no suggestion of influenza-like symptoms or temporal clustering to suggest that illness was related to vaccine.

The majority of triply seronegative vaccinees exhibited an antibody response to each vaccine component by HAI; fewer antibody rises to H3 and B hemagglutinins (heterologous antigens were used, see Materials and Methods) were detected by ELISA than HAI (Table 11). Of 17 triply seronegative vaccinees, 8 (47%) developed an antibody response to all three strains of the vaccine by HAI or ELISA. Mean postvaccination serum HAI titers were significantly higher for the H3 component than for the other two vaccine strains (Table 11). In contrast to seronegative children, ELISA was more sensitive than HAI at detecting antibody increases in seropositive children (Table 11). Of the 15 seropositive children, by ELISA 4 (27%) had antibody increases to H1, 4 (27%) to H3, and 5 (33%) to B hemagglutinin.

TABLE 11

Clinical and Serologic Responses After Intranasal Vaccination
With Cold-Adapted Trivalent Influenza Vaccine

		Group	
Finding	Seronegative <sup>b</sup> (n = 17)	Seropositive <sup>b</sup> (n = 15)	Control (n = 17)
AGE RANGE, MONTHS	7-23	10-116	6-60
NO. WITH ILLNESS <sup>a</sup>			
Fever	0	2	2
Upper respiratory illness (RI)	12°	5	8
Lower RI	0	0 _	1 .
Otitis media	2	4	1
SEROLOGIC RESPONSES TO VACCINE <sup>d</sup>			
H1N1/Kawasaki			

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Before vaccination	<2	5.3	1.2
After vaccination	2.7 <sup>e,f</sup>	5.3	1.2
No. with HAI response	10	0	0
No. with ELISA response	10	4	NT
H3N2/Korea			
Before vaccination	<2	5.5	1
After vaccination	4.1 <sup>e,f</sup>	6.1	1.2
No. with HAI response	12	2	0
No. with ELISA response	9	4	NT
B/Texas			
Before vaccination	<2	3.4	1
After vaccination	2.5 <sup>f</sup>	4.2	1.2
No. with HAI response	8	4	0
No. with ELISA response	6	5	NT

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HAI=hemagglutination inhibition assay; NT=not tested.

- a Fever, rectal temperature >38.3°C; upper Rl, ≥2 consecutive days with rhinnorhea or pharyngitis; lower Rl, wheezing or pneumonia; otitis media was diagnosed by a pediatrician.
  - <sup>b</sup> Seronegative (HAI <1:4) or seropositive (HAI ≥ 1:4) to all three strains of virus. Two children were vaccinated and were doubly or singly seronegative; they are not included in the analysis.

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- <sup>c</sup> Significantly more rhinorrhea than seropositive vaccinees (12 of 17 vs. 5 of 17, x 2 = 5.8; P < 0.05) but not significant when compared to controls (Fisher's exact test, P = 0.14).
- 30 d Antibody response defined as four-fold increase; for negative volunteers a titer rise from <1:4 to ≥1:8 by HAI or ≥1:20 by ELISA.</p>
  - <sup>e</sup> P <0.03, Student's *t* test.
- 35 f P < 0.02, Student's *t* test.

As shown in Table 12, viral shedding was observed in most seronegative volunteers and occurred significantly more often in seronegative recipients than in seropositive recipients ( $P \le 0.02$  for all comparisons between seronegatives and seropositives stratified by viral subtype). Sixteen of seventeen seronegative vaccinees shed at least one strain of virus; one vaccinee who failed to shed vaccine was infected with coxsackie B2 virus. Shedding of H1 and H3 was first observed 1 day after

vaccination while type B shedding began on day 2. The number of children shedding vaccine virus peaked on day 4 for H1, on day 6 for H3, and day 5 for B.

#### **TABLE 12**

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# Viral Shedding After Intranasal Vaccination With Cold-Adapted Trivalent Influenza Vaccine

	Subj	ects
Vaccine	Seronegative <sup>a</sup>	Seropositive <sup>a</sup>
H1N1/Kawasaki		
No. shedding/no. infected with vaccine virus <sup>b</sup>	10/12	2/5
Mean duration (days)	7.8	9
Mean peak titer (pfu/ml)	12	NT
H3N2/Korea		
No. shedding/no. infected with vaccine virus <sup>b</sup>	13/13	2/4
Mean duration (days)	8.8	6.5
Mean peak titer (pfu/ml)	74	NT
B/Texas		
No. shedding/no. infected with vaccine virus <sup>b</sup>	11/13	2/6
Mean duration (days)	9.4	3.5
Mean peak titer (pfu/ml)	41	NT

25

35

40

Eleven seronegative subjects were infected with all three vaccine viruses; NT = not tested.

Plaque assays to quantitate each subtype shed by seronegative vaccinees were done on samples from 15 of 17 volunteers (Table 11). The minimum titer detectable by plaque assay was 5 pfu/ml. Specimens positive by tube culture but negative by plaque assay were considered to have a titer < 5 pfu/ml. The highest mean viral titer was observed for H3 (74 pfu/ml), which was significantly higher than that of H1 (12 pfu/ml; p < 0.02, Student's t test). The highest titers of H1 were shed

a Hemagglutination inhibition assay seronegative and seropositive values, respectively,
 30 were <1:4 or ≥1:4.</li>

<sup>&</sup>lt;sup>b</sup> Indicated by viral shedding or antibody response by hemagglutination inhibition assay or by ELISA.

early, on days 3 and 4 after vaccination. Peak H3 and B titers were found on days 7 and 4 after vaccination, respectively.

Overall, 12 (71%), 13 (76%), and 13 (76%) of seronegative children were infected by H1N1, H3N2, or B vaccine viruses, respectively, as indicated by viral shedding or by HAI or ELISA antibody responses (Table 12). Eleven (65%) were infected by all three strains. Among seropositive children five (33%), four (27%), and six (40%) were infected by H1N1, H3N2, or B vaccine viral strains, respectively, as indicated by viral shedding or by HAI or ELISA antibody responses. None of the seropositive children was infected by all three vaccine viruses.

Those skilled in the art can now appreciate from the foregoing description that the broad teachings of the present invention can be implemented in a variety of forms. Therefore, while this invention has been described in connection with particular examples thereof, the true scope of the invention should not be so limited since other modifications will become apparent to the skilled practitioner upon a study of the drawings, specification and following claims.

10

15

All applications and publications cited herein are incorporated by reference.





# (1) GENERAL INFORMATION:

- (i) APPLICANT: Maassab, Hunein F Herlocher, Martha L
- (ii) TITLE OF INVENTION: Cold-adapted Influenza Virus
- (iii) NUMBER OF SEQUENCES: 40
- (iv) CORRESPONDENCE ADDRESS:
  - (A) ADDRESSEE: Anna M Lewak
  - (B) STREET: 5445 Corporate Drive
  - (C) CITY: Troy
  - (D) STATE: MI
  - (E) COUNTRY: USA
  - (F) ZIP: 48098
  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER: US
  - (B) FILING DATE:
  - (C) CLASSIFICATION:

#### (viii) ATTORNEY/AGENT INFORMATION:

- (A) NAME: Lewak, Anna M
- (B) REGISTRATION NUMBER: 33006
- (C) REFERENCE/DOCKET NUMBER: 2115-00257

# (ix) TELECOMMUNICATION INFORMATION:

- (A) TELEPHONE: 313-641-1600
- (B) TELEFAX: 313-641-0270

# (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 890 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)

#### (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Influenza virus
- (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)

# (vii) IMMEDIATE SOURCE:

(B) CLONE: NS

# (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 27..56
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2" /citation= ([1][2])

# (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(483, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 529..861
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2" /citation= ([1][2])

# (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(813, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)"
  /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: join(27..56, 529..861)
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 27..677

(D) OTHER INFORMATION: /product= "nonstructural protein NS1" /gene= "NS" /note= "nonstructural protein NS1" /citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G

- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:1: FROM 1 TO 890

# (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J Kitame, F Kendal, A P Maassab. H F Naeve. C

- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60(H2N2)
  - (C) JOURNAL: Virology
  - (D) VOLUME: 167
  - (F) PAGES: 554-567
  - (G) DATE: 1988
  - (K) RELEVANT RESIDUES IN SEQ ID NO:1: FROM 1 TO 890
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AGCAAAAGCA GGGUGACAAA GACAUA AUG GAU CCU AAC ACU GUG UCA AGC UUU Met Asp Pro Asn Thr Val Ser Ser Phe 1 5

CAG GUA Gln Val 10															101
GAA CUA Glu Lei															149
UCC CUA Ser Lei											_			_	197
ACC CGL Thr Arg															245
GAU GAG Asp Glu 75	ı Ala														293
CUA ACU Leu Thr 90															341
AUG CCC Met Pro															389
GCA AUG Ala Ile															437
UUU GAC Phe Asp															485
GGA GCA Gly Ala 155	ı Ile														533
ACU AAL Thr Asr 170															581
GAA UGG Glu Trp															629
GCU UGG Ala Trp															677
UAGAAA	GGA .	AAAU(	GGCG/	AG A	ACAAl	JUAG	G UC/	4444(	GUUC	GAA	GAAAl	JAA (	GAUG	GCUGAU	737
UGAAGAA	AGUG .	AGAC	4CAA	AU U	GAAGA	AUAA(	C AG	4GAAl	JAGU	UUU	GAGC/	444 1	JAAC	AUUUAU	797

GCAAGCCUUA	CAGCUGCUAU	UUGAAGUGGA	ACAAGAGAUA	AGAACUUUCU	CGUUUCAGCU
UAUUUAAUGA	UAAAAAACAC	CCUUGUUUCU	ACU		

857

890

# (2) INFORMATION FOR SEQ ID NO:2:

#### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 217 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp

5 10 15

His Val Arg Lys Gln Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 25 30

Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser 35 40 45

Thr Leu Gly Leu Asn Ile Glu Thr Ala Thr Arg Val Gly Lys Gln Ile 50 55 60

Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 65 70 75 80

Met Ala Ser Ala Pro Ala Ser Arg Tyr Leu Thr Asp Met Thr Ile Glu 85 90 95

Glu Met Ser Arg Asp Trp Phe Met Leu Met Pro Lys Gln Lys Val Ala 100 105 110

Gly Pro Leu Cys Ile Arg Met Asp Gln Ala Ile Met Asp Lys Asn Ile 115 120 125

Ile Leu Lys Ala Asn Phe Ser Val Ile Phe Asp Arg Leu Glu Thr Leu 130 135 140

Ile Leu Leu Arg Ala Phe Thr Glu Thr Gly Ala Ile Val Gly Glu Ile 145 150 155 160

Ser Pro Leu Pro Ser Leu Pro Gly His Thr Asn Glu Asp Val Lys Asn 165 170 175

Ala	Ile	Gly	Val 180	Leu	Ile	Gly	Gly	Leu 185	Glu	Trp	Asn	Asp	Asn 190	Thr	Val		
Arg	Val	Ser 195	Lys	Thr	Leu	Gln	Arg 200	Phe	Ala	Trp	Arg	Ser 205	Ser	Asp	Glu		
Asn	Gly 210	Arg	Pro	Pro	Leu	Thr 215	Pro	Lys					•				
(2)	INF	ORMA <sup>-</sup>	TION	FOR	SEQ	ID I	NO:3	•									
	(i	) SE(	QUENC	CE CH	HARA(	CTER:	ISTI	CS:									
		()	4) LE	ENGTH	H: 41	18 ba	ase p	pair:	S								
		(1	3) T	/PE:	nuc <sup>-</sup>	leic	acio	d									
		((	C) S	[RAN[	DEDNE	ESS:	sing	gle									
		([	) T(	)POL(	OGY:	line	ear										
	(ii	) MOI	_ECUL	E T	/PE:	RNA	(ger	nomi	c)								
	(ix	) FE	ATURI	:													
		()	4) NA	AME/k	KEY:	CDS							ć				
		(1	3) L(	CAT:	ON:	27.	. 389										
		[]	O) 0	ΓHER	INFO	ORMA <sup>-</sup>	TION		rodu ene=			stru	ctura	al pi	rotein	2"	
	(xi	) SE(	QUENC	CE DE	SCR1	[PT](	ON: S	SEQ	ID NO	0:3:							
AGC	4444	GCA (	GGGU(	GACA/	\A GA	ACAU/									C UÚU Phe		53
									AUG Met								101
GAG Glu	GAC Asp	UUG Leu	AAU Asn	GGA Gly 30	AUG Met	AUA Ile	ACA Thr	CAG Gln	UUC Phe 35	GAG Glu	UCU Ser	CUA Leu	AAA Lys	CUC Leu 40	UAC Tyr		149
AGA Arg	GAU Asp	UCG Ser	CUU Leu 45	GGA Gly	GAA Glu	GCA Ala	GUG Val	AUG Met 50	AGA Arg	AUG Met	GGA Gly	GAC Asp	CUC Leu 55	CAC His	UCA Ser		197

								-	59	-						
	CAA Gln															245
	GAA Glu 75															293
	GAG Glu															341
	UUU Phe															389
UAAl	UAAUGAUAAA AAACACCCUU GUUUCUACU													418		
(2)	INFO	ORMAT	TION	FOR	SEQ	ID 1	NO : 4									
	(	(i) S	SEQUE	ENCE	CHAF	RACT	ERIS	ΓICS	:							
			(A)	) LEI	NGTH	: 12	l am	ino a	acids	5						
			(B)	) TYI	PE: a	amino	o ac	i d								
			(D)	) TO	P0L0(	GY:	linea	ar								
	(1	ii) M	10LE(	CULE	TYP[	E: pi	rote <sup>.</sup>	in								
	,			LUCE	חבכו	-	ETON	C E /		NO.						

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Asp Ile Leu Met Arg Met Ser Lys Met Gln Leu Gly Ser Ser Ser Glu Asp Leu Asn Gly Met Ile 20 25 30

Thr Gln Phe Glu Ser Leu Lys Leu Tyr Arg Asp Ser Leu Gly Glu Ala

Val Met Arg Met Gly Asp Leu His Ser Leu Gln Asn Arg Asn Gly Lys 55

Trp Arg Glu Gln Leu Gly Gln Lys Phe Glu Glu Ile Arg Trp Leu Ile 65 70 75 80

Glu Glu Val Arg His Lys Leu Lys Ile Thr Glu Asn Ser Phe Glu Gln

Ile Thr Phe Met Gln Ala Leu Gln Leu Leu Phe Glu Val Glu Gln Glu 100 105 110

Ile Arg Thr Phe Ser Phe Gln Leu Ile

# (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1027 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: M
- (ix) FEATURE:
  - (A) NAME/KEY: exon
  - (B) LOCATION: 26..51
  - (D) OTHER INFORMATION: /product= "matrix protein M2" /gene= "M" /note= "matrix protein M2"

/citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 740..1004
- (D) OTHER INFORMATION: /product= "matrix protein M2" /gene= "M" /note= "matrix protein M2" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(969, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: join(26..51, 740..1004)
- (D) OTHER INFORMATION: /product= "matrix protein M2" /gene= "M" /note= "matrix protein M2" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 26..781

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus

(C) JOURNAL: Proceedings of the National Academy of Sciences of the USA

(G) DATE: 1993

(K) RELEVANT RESIDUES IN SEQ ID NO:5: FROM 1 TO 1027

# (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60(H2N2)
- (C) JOURNAL: Virology

(D) VOLUME: 167

(F) PAGES: 554-557

(G) DATE: 1988

(K) RELEVANT RESIDUES IN SEQ ID NO:5: FROM 1 TO 1027

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AGCA	<b>\</b> AAA(	GCA (	GGUA(	GAUAL	JU GA	\AAG				GUC Val		52
			UCU Ser									100
			GAA Glu								GCU Ala	148
			UGG Trp 45									196
			GGA Gly									244

CUG CAG CGU AGA CGC UUU Leu Gln Arg Arg Arg Phe 75			292
CCA AAU AAC AUG GAC AGA Pro Asn Asn Met Asp Arg 90 95	Ala Val Lys Leu T		340
GAG AUA ACA UUC CAU GGG Glu Ile Thr Phe His Gly 110			388
GGU GCA CUU GCC AGU UGU Gly Ala Leu Ala Ser Cys 125			436
GUG ACC ACU GAA GUG GUC Val Thr Thr Glu Val Val 140			484
AUU GCU GAC UCC CAG CAU Ile Ala Asp Ser Gln His 155			532
AAU CCA CUA AUA AGA CAU Asn Pro Leu Ile Arg His 170 175	Glu Asn Arg Met V		580
GCU AAG GCU AUG GAG CAA Ala Lys Ala Met Glu Gln 190			628
GCC AUG GAG GUU GCU AGU Ala Met Glu Val Ala Ser 205			676
GUU AUU GGG ACU CAU CCU Val Ile Gly Thr His Pro 220			724
CUU GAA AAU UUG CAG GCC Leu Glu Asn Leu Gln Ala 235			772
CGA UUC AAG UGACCCUCUU ( Arg Phe Lys 250	GUUGUUGCCG CGAGUAU	CAU UGGGAUCUUG	821
CACUUGAUAU UGUGGAUUCU U	GAUCAUCUU UUUUUCAA	AU GCAUUUAUCG CUUCUUUAAA	881
CACGGUCUGA AAAGAGGGCC UI	JCUACGGAA GGAGUACC	AG AGUCUAUGAG GGAAGAAUAU	941
CGAAAGGAAC AGCAGAGUGC U	GUGGAUUCU GACGAUAG	UC AUUUUGUCAG CAUAGAGCUG	1001
GAGUAAAAAA CUACCUUGUU U	CUACU	,	1027

# (2) INFORMATION FOR SEQ ID NO:6:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 252 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Ser Leu Leu Thr Glu Val Glu Thr Tyr Val Leu Ser Ile Ile Pro 1 5 10 15

Ser Gly Pro Leu Lys Ala Glu Ile Ala Gln Arg Leu Glu Asp Val Phe 20 25 30

Ala Gly Lys Asn Thr Asp Leu Glu Ala Leu Met Glu Trp Leu Lys Thr 35 40 45

Arg Pro Ile Leu Ser Pro Leu Thr Lys Gly Ile Leu Gly Phe Val Phe 50 55 60

Thr Leu Thr Val Pro Ser Glu Arg Gly Leu Gln Arg Arg Arg Phe Val 65 70 75 80

Gln Asn Ala Leu Asn Gly Asn Gly Asp Pro Asn Asn Met Asp Arg Ala 85 90 95

Val Lys Leu Tyr Arg Lys Leu Lys Arg Glu Ile Thr Phe His Gly Ala 100 105 110

Lys Glu Île Ala Leu Ser Tyr Ser Ala Gly Ala Leu Ala Ser Cys Met 115 120 125

Gly Leu Ile Tyr Asn Arg Met Gly Ala Val Thr Thr Glu Val Val Leu 130 135 140

Gly Leu Val Cys Ala Thr Cys Glu Gln Ile Ala Asp Ser Gln His Arg 145 150 155 160

Ser His Arg Gln Met Val Thr Thr Asn Pro Leu Ile Arg His Glu 165 170 175

Asn Arg Met Val Leu Ala Ser Thr Thr Ala Lys Ala Met Glu Gln Met 180 185 190

Ala Gly Ser Ser Glu Gln Ala Ala Glu Ala Met Glu Val Ala Ser Gln 195 200 205

Ala	Arg 210	Gln	Met	Val	Gln	Ala 215	Met	Arg	Val	Ile	Gly 220	Thr	His	Pro	Ser	
Ser 225	Ser	Ala	Gly	Leu	Lys 230	Asn	Asp	Leu	Leu	G1u 235	Asn	Leu	Gln	Ala	Tyr 240	
Gln	Lys	Arg	Met	Gly 245	Val	Gln	Met	Gln	Arg 250	Phe	Lys					
(2)	INF	ORMA"	TION	FOR	SEQ	ID N	10:7:	:								
	(i)	) SE(	QUENC	CE CH	IARA(	CTERI	STIC	CS:						,		
		()	4) LE	ENGTH	1: 33	39 ba	ase p	pairs	5							
		(1	3) T	/PE:	nuc]	leic	acio	t								
		((	C) S	[RAN[	DEDNE	ESS:	sing	gle								
		([	)) T(	POLO	OGY:	line	ear									
	(ii)	) MOI	ECUL	.E T\	/PE:	RNA	(ger	nomid	c)							
	(ix	) FE	ATURE	:						•						
		()	4) NA	AME/k	KEY:	CDS										
		(1	3) L(	CAT 1	ON:	26	316									
		([	0) 0	THER	INFO	)RMAT	TION:	: /pr	roduc	ct= '	'Matr	rix M	12"			
	(xi)	) SE(	QUEN(	CE DE	SCR1	[PT](	ON: S	SEO I	IÒ NO	D:7:						
AGCA			GGUAC								ACC	GAG	GUC	GAA	ACG	52
, 100,				., 10, 10	, o .	•••			Leu							
			AAC Asn													100
			GCC Ala													148
			CAU His 45													196

	AGA Arg											244
	CGA Arg											292
	AGC Ser			UAA	VAAA(	CUA (	CCUU	GUUU(	CU AC	CU		339

# (2) INFORMATION FOR SEQ ID NO:8:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 97 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly
1 5 10 15

Cys Arg Cys Asn Asp Ser Ser Asp Pro Leu Val Val Ala Ala Ser Ile 20 25 30

Ile Gly Ile Leu His Leu Ile Leu Trp Ile Leu Asp His Leu Phe Phe 35 40 45

Lys Cys Ile Tyr Arg Phe Phe Lys His Gly Leu Lys Arg Gly Pro Ser 50 60

Thr Glu Gly Val Pro Glu Ser Met Arg Glu Glu Tyr Arg Lys Glu Gln 65 70 75 80

Gln Ser Ala Val Asp Ser Asp Asp Ser His Phe Val Ser Ile Glu Leu 85 90 95

Glu

# (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1566 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: NP
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(113, "c")
  - (D) OTHER INFORMATION: /note= "c in ca "master" strain; a in wt2(3); a in 1988 reported ca vaccine strain (manuscript), but c reported in 1988 genbank" /citation= ([1][2])

#### . (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(146, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(627, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain and in wt2(3); a in 1988 reported ca vaccine strain"

/citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(909, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); c in 1988 reported ca vaccine strain"

/citation= ([1][2])

# (ix) FEATURÉ:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1550, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3)" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 46..1539
- (D) OTHER INFORMATION: /product= "Nucleoprotein" /gene= "NP" /note= "nucleoprotein" /citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R W
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus

(C) JOURNAL: Proceedings of the National Academy of Sciences of the USA

(G) DATE: 1993

(K) RELEVANT RESIDUES IN SEQ ID NO:9: FROM 1 TO 1566

### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox. N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

(B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60 (H2N2)

(C) JOURNAL: Virology

(D) VOLUME: 167

(F) PAGES: 554-567

(G) DATE: 1988

(K) RELEVANT RESIDUES IN SEQ ID NO:9: FROM 1 TO 1566

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGCA	AGCAAAAGCA GGGUAGAUAA UCACUCACUG AGUGACAUCA AAAUC AUG GCG UCC Met Ala Ser 1															. 54
														GAA Glu		102
														GGU Gly		150
														AGU Ser 50		198
UAU Tyr	GAG Glu	GGG Gly	CGG Arg 55	CUG Leu	AUC Ile	CAG Gln	AAC Asn	AGC Ser 60	UUA Leu	ACA Thr	AUA Ile	GAG Glu	AGA Arg 65	AUG Met	GUG Val	246

			AGG Arg							294
			AAG Lys 90							342
			AGG Arg							390
			CAA Gln						·	438
			AUC Ile							486
			CUU Leu							534
			UCG Ser 170							582
			GUU Val							630
			AAU Asn							678
			GCU Ala	Tyr						726
			GCA Ala							774
			AAU Asn 250						·	822
			UUG Leu							870
			GGA Gly							918

	AAA Lys												966
	AAC Asn												1014
	AAG Lys 325									Ala			1062
	CUA Leu												1110
	AAA Lys												1158
	ACU Thr												1206
	AGG Arg												1254
	CAA Gln 405												1302
	GAC Asp												1350
	ACA Thr												1398
	CCA Pro												1446
	GAA Glu												1494
	GGA Gly 485												1539
UAA	GGAAA	AAA A	AUAC(	CCUU	GU UI	JCUA(	CU						1566

# (2) INFORMATION FOR SEQ ID NO:10:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 498 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Ala Ser Gln Gly Thr Lys Arg Ser Tyr Glu Gln Met Glu Thr Asp 1 5 10 15

Gly Glu Arg Gln Asn Ala Thr Glu Ile Arg Ala Ser Val Gly Lys Met 20 25 30

Ile Gly Gly Ile Gly Arg Phe Tyr Ile Gln Met Cys Thr Glu Leu Lys 35 40 45

Leu Ser Asp Tyr Glu Gly Arg Leu Ile Gln Asn Ser Leu Thr Ile Glu 50 55 60

Arg Met Val Leu Ser Ala Phe Asp Glu Arg Arg Asn Lys Tyr Leu Glu 65 70 75 80

Glu His Pro Ser Ala Gly Lys Asp Pro Lys Lys Thr Gly Gly Pro Ile 85 90 95

Tyr Lys Arg Val Asp Gly Lys Trp Met Arg Glu Leu Val Leu Tyr Asp 100 105 110

Lys Glu Glu Ile Arg Arg Ile Trp Arg Gln Ala Asn Asn Gly Asp Asp 115 120 125

Ala Thr Ala Gly Leu Thr His Met Met Ile Trp His Ser Asn Leu Asn 130 135 140

Asp Thr Thr Tyr Gln Arg Thr Arg Ala Leu Val Arg Thr Gly Met Asp 145 150 155 160

Pro Arg Met Cys Ser Leu Met Gln Gly Ser Thr Leu Pro Arg Arg Ser 165 170 175

Gly Ala Ala Gly Ala Ala Val Lys Gly Val Gly Thr Met Val Met Glu 180 185 190

Leu Ile Arg Met Ile Lys Arg Gly Ile Asn Asp Arg Asn Phe Trp Arg 195 200 205

Gly Glu Asn Gly Arg Lys Thr Arg Asn Ala Tyr Glu Arg Met Cys Asn 215 Ile Leu Lys Gly Lys Phe Gln Thr Ala Ala Gln Arg Ala Met Met Asp 235 Gln Val Arg Glu Ser Arg Asn Pro Gly Asn Ala Glu Ile Glu Asp Leu Ile Phe Leu Ala Arg Ser Ala Leu Ile Leu Arg Gly Ser Val Ala His 270 Ser Cys Leu Pro Ala Cys Val Tyr Gly Pro Ala Val Ala Ser Gly 275 280 285 Tyr Asp Phe Glu Lys Glu Gly Tyr Ser Leu Val Gly Ile Asp Pro Phe 295 Lys Leu Leu Gln Asn Ser Gln Val Tyr Ser Leu Ile Arg Pro Asn Glu 305 310 315 320 Asn Pro Ala His Lys Ser Gln Leu Val Trp Met Ala Cys Asn Ser'Ala 325 330 335 Ala Phe Glu Asp Leu Arg Val Ser Ser Phe Ile Arg Gly Thr Lys Val 345 Ile Pro Arg Gly Lys Leu Ser Thr Arg Gly Val Gln Ile Ala Ser Asn 355 365 Glu Asn Met Asp Thr Met Gly Ser Ser Thr Leu Glu Leu Arg Ser Arg 370 375 Tyr Trp Ala Ile Arg Thr Arg Ser Gly Gly Asn Thr Asn Gln Gln Arg 390 395 Ala Ser Ala Gly Gln Ile Ser Val Gln Pro Thr Phe Ser Val Gln Arg 405 410 415 Asn Leu Pro Phe Asp Lys Pro Thr Ile Met Ala Ala Phe Thr Gly Asn 425 Ala Glu Gly Arg Thr Ser Asp Met Arg Ala Glu Ile Ile Arg Met Met 440 Glu Gly Ala Lys Pro Glu Glu Val Ser Phe Gln Gly Arg Gly Val Phe 450 455 460 Glu Leu Ser Asp Glu Lys Ala Thr Asn Pro Ile Val Pro Ser Phe Asp 470 475 465 Met Ser Asn Glu Gly Ser Tyr Phe Phe Gly Asp Asn Ala Glu Glu Tyr 490

#### (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2233 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: PA
- (ix) FEATURE:
  - (A) NAME/KEY: conflict
  - (B) LOCATION: replace(20, "c")
    - (D) OTHER INFORMATION: /note= "c in ca "master" strain and in wt2(3)" /citation= ([1][2])
- (ix) FEATURE:
  - (A) NAME/KEY: conflict
  - (B) LOCATION: replace(75, "g")
  - (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); u in 1988 reported ca vaccine strain" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1861, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2167..2168, "cc")
- (D) OTHER INFORMATION: /note= "cc in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 25..2172
- (D) OTHER INFORMATION: /product= "polymerase acidic protein" /gene= "PA" /note= "polymerase acidic protein" /citation= ([1][2])

# (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:11: FROM 1 TO 2233

# (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Cox, N J Kitame, F Kendal, A P Maassab, H F Naeve, C
- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza strain, A/Ann Arbor/6/60(H2N2)
- (C) JOURNAL: Virology
- (D) VOLUME: 167
- (F) PAGES: 554-567
- (G) DATE: 1988
- (K) RELEVANT RESIDUES IN SEQ ID NO:11: FROM 1 TO 2233

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AGCGAAAGCA 0	GGUACUGAUC CGAA		UU GUG CGA CAA UGC U he Val Arg Gln Cys P 5	
			GCA AUG AAA GAG UAU Ala Met Lys Glu Tyr 20	
			GCA GCA AUA UGC ACU Ala Ala Ile Cys Thr 40	
			CAU UUC AUC AAU GAG His Phe Ile Asn Glu 55	
			CCA AAU GCA CUU UUG Pro Asn Ala Leu Leu 70	
		Gly Arg Asp A	CGC ACA AUG GCC UGG Arg Thr Met Ala Trp 85	
		n Thr Thr Gly A	GCU GAG AAA CCG AAG Ala Glu Lys Pro Lys 100	

CUG Leu	CCA Pro	GAU Asp	UUG Leu	UAU Tyr 110	GAU Asp	UAC Tyr	AAG Lys	GAG Glu	AAU Asn 115	AGA Arg	UUC Phe	AUC Ile	GAG Glu	AUU Ile 120	GGA Gly	387
						CAC His										435
AUU Ile	AAA Lys	UCU Ser 140	GAG Glu	AAG Lys	ACA Thr	CAC His	AUC Ile 145	CAC His	AUU Ile	UUC Phe	UCA Ser	UUC Phe 150	ACU Thr	GGG Gly	GAA Glu	483
GAA Glu	AUG Met 155	GCC Ala	ACA Thr	AAG Lys	GCC Ala	GAC Asp 160	UAC Tyr	ACU Thr	CUC Leu	GAU Asp	GAG Glu 165	GAA Glu	AGC Ser	AGG Arg	GCU Ala	531
						UUC Phe										579
GGC Gly	CUC Leu	UGG Trp	GAU Asp	UCC Ser- 190	UUU Phe	CAU His	CAG Gln	UCC Ser	GAA Glu 195	AGA Arg	GGC Gly	GAA Glu	GAA Glu	ACA Thr 200	AUU Ile	627
GAA G1u	GAA Glu	AGA Arg	UUU Phe 205	GAA G1u	AUC	ACA Thr	GGG Gly	ACA Thr 210	AUG Met	CGC Arg	AGG Arg	CUC Leu	GCC Ala 215	GAC Asp	CAA Gln	675
						UCC Ser										723
GAU Asp	GGA Gly 235	UUC Phe	GAA Glu	CCG Pro	AAC Asn	GGC Gly 240	UAC Tyr	AUU Ile	GAG Glu	GGC Gly	AAG Lys 245	CUU Leu	UCU Ser	CAA Gln	AUG Met	771
						AAA Lys										819
AGA Arg	CCA Pro	AUU Ile	AGA Arg	CUU Leu 270	CCG Pro	GAU Asp	GGG Gly	CCU Pro	CCU Pro 275	UGU Cys	UCU Ser	CAG Gln	CGG Arg	UCC Ser 280	AAA Lys	867
						UUA Leu										915
						CUA Leu										963
UUC Phe	UUU Phe 315	GGA Gly	UGG Trp	AAA Lys	GAA Glu	CCC Pro 320	UAU Tyr	GUU Val	GUU Val	AAA Lys	CCA Pro 325	CAC His	GAA Glu	AAG Lys	GGA Gly	1011

								-	78	-							
		CCA Pro															1059
		AUU Ile															1107
		AGU Ser															1155
		GAC Asp 380															1203
		GAU Asp															1251
GAG Glu 410	UUC Phe	AAC Asn	AAG Lys	GCA Ala	UGC Cys 415	GAG Glu	CUG Leu	ACC Thr	GAU Asp	UCA Ser 420	AUC Ile	UGG Trp	AUA Ile	GAG G1u	CUC Leu 425		1299
		AUU Ile															1347
		AAU Asn															1395
		AUG Met 460														J	1443
		GCA Ala															1491
		AAA Lys															1539
		AGA Arg															1587
		GAG Glu															1635
		UAC Tyr 540															1683

			CAG Gln										Arg				1731
T			AAG Lys														1779
			UCA Ser														1827
G V	UC al	AAG Lys	GAG Glu	AAA Lys 605	GAC Asp	AUG Met	ACC Thr	AAA Lys	GAG Glu 610	UUU Phe	UUC Phe	GAG Glu	AAU Asn	AAA Lys 615	UCA Ser	GAA Glu	1875
A T	CA hr	UGG Trp	CCC Pro 620	AUU Ile	GGA Gly	GAG Glu	UCC Ser	CCC Pro 625	AAA Lys	GGA Gly	GUG Val	GAA Glu	GAA Glu 630	GGU Gly	UCC Ser	AUU Ile	1923
			GUC Val														1971
T			UCU Ser														2019
			GUC Val														2067
			GGG Gly														2115
			GUU Val 700														2163
		CCA Pro 715	AGA Arg	UAGL	JUGU(	GC A	\AUG(	CUACI	JA Ul	JUGCL	JAUC(	C AUA	ACUGI	JCCA			2212
Α	AA/	\AGU/	ACC (	JUGUL	JUCUA	AC U							•				2233

# (2) INFORMATION FOR SEQ ID NO:12:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 716 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Glu Asp Phe Val Arg Gln Cys Phe Asn Pro Met Ile Val Glu Leu Ala Glu Lys Ala Met Lys Glu Tyr Gly Glu Asp Leu Lys Ile Glu Thr 20 25 30 Asn Lys Phe Ala Ala Ile Cys Thr His Leu Glu Val Cys Phe Met Tyr Ser Asp Phe His Phe Ile Asn Glu Gln Gly Glu Ser Ile Ile Val Glu Leu Asp Asp Pro Asn Ala Leu Leu Lys His Arg Phe Glu Ile Ile Glu Gly Arg Asp Arg Thr Met Ala Trp Thr Val Val Asn Ser Ile Cys Asn
90
95 Thr Thr Gly Ala Glu Lys Pro Lys Phe Leu Pro Asp Leu Tyr Asp Tyr 105 100 Glu Asn Arg Phe Ile Glu Ile Gly Val Thr Arg Arg Glu Val His Ile Tyr Tyr Leu Glu Lys Ala Asn Lys Ile Lys Ser Glu Lys Thr His 130 135 140 Ile His Ile Phe Ser Phe Thr Gly Glu Glu Met Ala Thr Lys Ala Asp 145 Tyr Thr Leu Asp Glu Glu Ser Arg Ala Arg Ile Lys Thr Arg Leu Phe 165 Thr Ile Arg Gln Glu Met Ala Ser Arg Gly Leu Trp Asp Ser Phe His 180 185 190 180 Gln Ser Glu Arg Gly Glu Glu Thr Ile Glu Glu Arg Phe Glu Ile Thr 195 200 Gly Thr Met Arg Arg Leu Ala Asp Gln Ser Leu Pro Pro Asn Phe Ser Cys Leu Glu Asn Phe Arg Ala Tyr Val Asp Gly Phe Glu Pro Asn Gly 225 - 230 235 240 Tyr Ile Glu Gly Lys Leu Ser Gln Met Ser Lys Glu Val Asn Ala Lys 245

Ile Glu Pro Phe Leu Lys Thr Thr Pro Arg Pro Ile Arg Leu Pro Asp 260 265 Gly Pro Pro Cys Ser Gln Arg Ser Lys Phe Leu Leu Met Asp Ala Leu 280 Lys Leu Ser Ile Glu Asp Pro Ser His Glu Gly Glu Gly Ile Pro Leu Tyr Asp Ala Ile Lys Cys Met Arg Thr Phe Phe Gly Trp Lys Glu Pro 315 Tyr Val Val Lys Pro His Glu Lys Gly Ile Asn Pro Asn Tyr Leu Leu 330 Ser Trp Lys Gln Val Leu Ala Glu Leu Gln Asp Ile Glu Asn Glu Glu 345 Lys Ile Pro Arg Thr Lys Asn Met Lys Lys Thr Ser Gln Leu Lys Trp Ala Leu Gly Glu Asn Met Ala Pro Glu Lys Val Asp Phe Asp Asp Cys 370 375 Arg Asp Val Ser Asp Leu Lys Gln Tyr Asp Ser Asp Glu Pro Glu Leu 390 395 Arg Ser Leu Ser Ser Trp Ile Gln Asn Glu Phe Asn Lys Ala Cys Glu . 405 Leu Thr Asp Ser Ile Trp Ile Glu Leu Asp Glu Ile Gly Glu Asp Val 420 Ala Pro Ile Glu His Ile Ala Ser Met Arg Arg Asn Tyr Phe Thr Ala Glu Val Ser His Cys Arg Ala Thr Glu Tyr Ile Met Lys Gly Val Tyr 455 Ile Asn Thr Ala Leu Leu Asn Ala Ser Cys Ala Ala Met Asp Asp Phe 470 480 465 Gln Leu Ile Pro Met Ile Ser Lys Cys Arg Thr Lys Glu Gly Arg Arg 490 Lys Thr Asn Leu Tyr Gly Phe Ile Ile Lys Gly Arg Ser His Leu Arg Asn Asp Thr Asp Val Val Asn Phe Val Ser Met Glu Phe Ser Leu Thr 515 520 525 Asp Pro Arg Leu Glu Pro His Lys Trp Glu Lys Tyr Cys Val Leu Glu 535 Ile Gly Asp Met Leu Leu Arg Ser Ala Ile Gly Gln Val Ser Arg Pro

545 550 555 560 Met Phe Leu Tyr Val Arg Thr Asn Gly Thr Ser Lys Ile Lys Met Lys 565 Trp Gly Met Glu Met Arg Arg Cys Leu Leu Gln Ser Leu Gln Gln Ile 580 585 590 Glu Ser Met Ile Glu Ala Glu Ser Ser Val Lys Glu Lys Asp Met Ihr 600 Lys Glu Phe Phe Glu Asn Lys Ser Glu Thr Trp Pro Ile Gly Glu Ser Pro Lys Gly Val Glu Glu Gly Ser Ile Gly Lys Val Cys Arg Thr Leu 625 Leu Ala Lys Ser Val Phe Asn Ser Leu Tyr Ala Ser Pro Gin Leu Giu 645 655 Gly Phe Ser Ala Glu Ser Arg Lys Leu Leu Leu Val Val Gln Ala Leu 665 Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Leu Gly Gly Leu Tyr Glu 675

Ala Ile Glu Glu Cys Leu Ile Asn Asp Pro Trp Val Leu Leu Asn Ala

(2) INFORMATION FOR SEQ ID NO:13:

705

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2341 base pairs

695

Ser Trp Phe Asn Ser Phe Leu Thr His Ala Pro Arg

710

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)

#### (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Influenza virus
- (B) STRAIN: cold adapted "Master Strain" A/AA/6/60 7PI (H2N2)

### (vii) IMMEDIATE SOURCE:

(B) CLONE: PB1

# (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(123, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(486. "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1195, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(1276, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain; a in wt2(3); g in 1988 reported ca vaccine strain" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1395, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1766, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2005, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2019, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 25..2295

			-	85 -			
(x) F	PUBL :	ICATION II	NFORMATION:				
	(A)	AUTHORS:	Herlocher, M   Maassab, H F Webster, R G	-			
	(B)	TITLE: Mo	olecular and b aster strain A	iological /AA/6/60	changes ir (H2N2) infl	n the cold Luenza viru	adapted s
	(C)	JOURNAL:	Proceedings of the USA	f the Nat	ional Acade	emy of Scie	nces of
	(G)	DATE: 199	93				
	(K)	RELEVANT	RESIDUES IN SI	EQ ID NO:	13: FROM 1	TO 2341	
(x) F	PUBL :	CATION II	NFORMATION:				
	(A)	AUTHORS:	Cox, N J Kitame, F Kendal, A P Maassab, H F Naeve, C				
	(B)	TITLE: I	dentification ( ive attenuated	of sequenc influenza	ce changes a vaccine s	in the col strain	d-adapted
	(C)	JOURNAL:	Virology				
	(D)	VOLUME:	167				
	(F)	PAGES: 5	54-567				
	(G)	DATE: 198	38				
	(K)	RELEVANT	RESIDUES IN SI	EQ ID NO:	13: FROM 1	TO 2341	
(xi) S	SEQUE	ENCE DESCI	RIPTION: SEQ I	NO:13:			
AGCGAAAGCA	A GG(	CAAACCAU (	JUGA AUG GAU GI Met Asp Va 1		G ACC UUA ( c Thr Leu L		51
			A AAU GCC AUA n Asn Ala Ile S				99

GGA GAU CCU CCA UAC AGC CAU GGG ACA GGA ACA GGA UAC ACC AUG GAC Gly Asp Pro Pro Tyr Ser His Gly Thr Gly Thr Gly Tyr Thr Met Asp 30 . 35 40147

						CAA Gln										195
						CAC His										243
						AGU Ser 80										291
GAA Glu 90	GCA Ala	AUG Met	GCU Ala	UUC Phe	CUU Leu 95	GAA Glu	GAA Glu	UCC Ser	CAC His	CCA Pro 100	GGA Gly	AUC Ile	UUU Phe	GAA Glu	AAC Asn 105	339
						GAA Glu										38,7
						ACC Thr										435
						GCC Ala										483
						UCG Ser 160										531
						AAA Lys										579
						AGA Arg		Asn								627
						AAG. Lys										675
						UUG Leu										723
						GCA Ala 240										771
						GAA Glu										819

														GCU Ala 280		867
														ACA Thr		915
														AAU Asn		963
														AAU Asn		1011
CCU Pro 330	GAA Glu	UGG Trp	UUU Phe	AGA Arg	AAC Asn 335	GUC Val	CUG Leu	AGC Ser	AUC Ile	GCA Ala 340	CCU Pro	AUA Ile	AUG Met	UUC Phe	UCA Ser 345	1059
														AAG Lys 360		1107
														AUU Ile		1155
														AUA Ile		1203
														AUG Met		1251
														CUG Leu		1299
														GGA Gly 440		1347
														CAU His		1395
														CUA Leu	_	1443
														GGG Gly		1491

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UUU GA Phe G 490										1539
AGC AI Ser M										1587
GAU AI Asp Mo		er								1635
GAC CI Asp Le	eu G									1683
GAC UA Asp To 5										1731
ACA AG Thr Ag 570										1779
AAG GO										1827
CGG Av Arg As		eu								1875
GAA GA Glu As	sp T									1923
CAU AA His Ly 60										1971
GGU CO Gly Po 650										2019
UGG AU Trp I										2067
AUU CI Ile Le		lu .								2115
AAA UI Lys Pi	he Pl									2163

								-	89	-					
			GCC Ala												2211
			GGA Gly												2259
			ACC Thr								UAGL	JGAAL	JUU		2305
AGCl	JUGUC	CCU L	JCAU6	AAA/	\A Al	JGCCL	JUGUĹ	J UCL	JACU						2341
(2)	INFO	RMAT	ΓΙΟN	FOR	SEQ	ID N	10:14	<b>1</b> :							
	(	(i) S	SEQUE	ENCE	CHAF	RACTE	RIST	ICS:							
			(A)	LEN	IGTH :	757	'ami	no a	cids	5					

# (ii) MOLECULE TYPE: protein

(B) TYPE: amino acid

(D) TOPOLOGY: linear

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Asp Val Asn Pro Thr Leu Leu Phe Leu Lys Val Pro Ala Gln Asn 1 5 10 15

Ala Ile Ser Thr Thr Phe Pro Tyr Thr Gly Asp Pro Pro Tyr Ser His 20 25 30

Gly Thr Gly Tyr Thr Met Asp Thr Val Asn Arg Thr His Gln
35 40 45

Tyr Ser Glu Lys Gly Lys Trp Thr Thr Asn Thr Glu Thr Gly Ala His 50 55 60

Gln Leu Asn Pro Ile Asp Gly Pro Leu Pro Glu Asp Asn Glu Pro Ser 65 70 75 80

Gly Tyr Ala Gln Thr Asp Cys Val Leu Glu Ala Met Ala Phe Leu Glu 85 · 90 95

Glu Ser His Pro Gly Ile Phe Glu Asn Ser Cys Leu Glu Thr Met Glu 100 105 110

Val Ile Gln Gln Thr Arg Val Asp Lys Leu Thr Gln Gly Arg Gln Thr 115 120 125 Tyr Asp Trp Thr Leu Asn Arg Asn Gln Pro Ala Ala Thr Ala Leu Ala 135 140 Asn Thr Ile Glu Val Phe Arg Ser Asn Gly Leu Thr Ala Asn Glu Ser 145 150 155 Gly Arg Leu Ile Asp Phe Leu Lys Asp Val Ile Glu Ser Met Asp Lys Glu Glu Met Glu Ile Thr Thr His Phe Gln Arg Lys Arg Arg Val Arg 180 Asp Asn Met Thr Lys Lys Met Val Thr Gln Arg Thr Ile Gly Lys Lys Lys Gln Arg Leu Asn Lys Arg Ser Tyr Leu Ile Arg Ala Leu Thr Leu Asn Thr Met Thr Lys Asp Ala Glu Arg Gly Lys Leu Lys Arg Arg Ala 225 230 235 240 Ile Ala Thr Pro Gly Met Gln Ile Arg Gly Phe Val Tyr Phe Val Glu 245 250 255 Thr Leu Ala Arg Ser Ile Cys Glu Lys Leu Glu Gln Ser Gly Leu Pro Val Gly Gly Asn Glu Lys Lys Ala Lys Leu Ala Asn Val Val Arg Lys 275 280 285 Met Met Thr Asn Ser Gln Asp Thr Glu Leu Ser Phe Thr Ile Thr Gly 295 300 Asp Asn Thr Lys Trp Asn Glu Asn Gln Asn Pro Arg Met Phe Leu Ala Met Ile Thr Tyr Ile Thr Arg Asn Gln Pro Glu Trp Phe Arg Asn Val 335 325 330 Leu Ser Ile Ala Pro Ile Met Phe Ser Asn Lys Met Ala Arg Leu Gly Gly Tyr Met Phe Lys Ser Lys Ser Met Lys Leu Arg Thr Gln Ile Pro Ala Glu Met Leu Ala Ser Ile Asp Leu Lys Tyr Phe Asn Glu Ser 370 380 Thr Arg Lys Lys Ile Glu Glu Ile Arg Pro Leu Leu Ile Asp Gly Thr 385 390 395 Val Ser Leu Ser Pro Gly Met Met Met Gly Met Phe Asn Met Leu Ser 405 410

Thr Val Leu Gly Val Ser Ile Leu Asn Leu Gly Gln Lys Lys Tyr Thr Lys Thr Thr Tyr Trp Trp Asp Gly Leu Gln Ser Ser Asp Asp Phe Ala 435 440 445 Leu Ile Val Asn Ala Pro Asn His Asp Gly Ile Gln Ala Gly Val Asp 455 Arg Phe Tyr Arg Thr Cys Lys Leu Val Gly Ile Asn Met Ser Lys Lys 465 470 475 480 Lys Ser Tyr Ile Asn Arg Thr Gly Thr Phe Glu Phe Thr Ser Phe Phe Tyr Arg Tyr Gly Phe Val Ala Asn Phe Ser Met Glu Leu Pro Ser Phe 505 Gly Val Ser Gly Ile Asn Glu Ser Ala Asp Met Ser Ile Gly Val Thr Val Ile Lys Asn Asn Met Ile Asn Asn Asp Leu Gly Pro Ala Thr Ala Gln Leu Ala Leu Gln Leu Phe Ile Lys Asp Tyr Arg Tyr Thr Tyr Arg 545 Cys His Arg Gly Asp Thr Gln Ile Gln Thr Arg Arg Ser Phe Glu Leu 565 570 575 Lys Lys Leu Trp Gly Gln Thr Arg Ser Lys Ala Gly Leu Leu Val Ser Asp Gly Gly Pro Asn Leu Tyr Asn Ile Arg Asn Leu His Ile Pro Glu 600 605 Val Cys Leu Lys Trp Glu Leu Met Asp Glu Asp Tyr Gln Gly Arg Leu Cys Asn Pro Leu Asn Pro Phe Val Ser His Lys Glu Ile Glu Ser Val 640 Asn Asn Ala Val Val Met Pro Ala His Gly Pro Ala Lys Ser Met Glu 645 Tyr Asp Ala Val Thr Thr Thr His Ser Trp Ile Pro Lys Arg Asn Arg Ser Ile Leu Asn Thr Ser Gln Arg Gly Ile Leu Glu Asp Glu Gln Met 675 Tyr Gln Lys Cys Cys Asn Leu Phe Glu Lys Phe Phe Pro Ser Ser Ser 695

Tyr Arg Arg Pro Val Gly Ile Ser Ser Met Val Glu Ala Met Val Ser 705 710 715 720

Arg Ala Arg Ile Asp Ala Arg Ile Asp Phe Glu Ser Gly Arg Ile Lys 725 730 735

Lys Glu Glu Phe Ala Glu Ile Met Lys Ile Cys Ser Thr Ile Glu Glu 740 745 750

Leu Arg Arg Gln Lys 755

# (2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2341 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: PB2
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(141, "g")
  - (D) OTHER INFORMATION: /note= "g in ca "master" strain: a in wt2(3); g in 1988 reported ca vaccine strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(426, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(714, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); c in 1988 reported ca vaccine strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(821, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(963, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported ca vaccine strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1182, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1212, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1353, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1923, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3)" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(1933, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain; u in wt2(3); u in 1988 reported ca vaccine strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 28..2304
- (D) OTHER INFORMATION: /product= "polymerase basic 2" /gene= "PB2" /note= "polymerase basic 2" /citation= ([1][2])

Thr Phe Gly Pro Val His Phe Arg Asn Gln Val Lys Ile Arg Arg 135 Val Asp Ile Asn Pro Gly His Ala Asp Leu Ser Ala Lys Glu Ala Gln 160 Asp Val Ile Met Glu Val Val Phe Pro Asn Glu Val Gly Ala Arg Ile 165 Leu Thr Ser Glu Ser Gln Leu Thr Ile Thr Lys Glu Lys Lys Glu Glu 185 Leu Gln Asp Cys Lys Ile Ser Pro Leu Met Val Ala Tyr Met Leu Glu 195 205 Arg Glu Leu Val Arg Lys Thr Arg Phe Leu Pro Val Ala Gly Gly Thr 215 Ser Ser Val Tyr Ile Glu Val Leu His Leu Thr Gln Gly Thr Cys Trp 235 Glu Gln Met Tyr Thr Pro Gly Gly Glu Val Arg Asn Asp Asp Val Asp 245 Gln Ser Leu Ile Ile Ala Ala Arg Ser Ile Val Arg Arg Ala Ala Val 265 Ser Ala Asp Pro Leu Ala Ser Leu Leu Glu Met Cys His Ser Thr Gln 280 Ile Gly Gly Thr Arg Met Val Asp Ile Leù Arg Gln Asn Pro Thr Glu 290 295 300 Glu Gln Ala Val Glu Ile Cys Lys Ala Ala Met Gly Leu Arg Ile Ser 315 320 Ser Ser Phe Ser Phe Gly Gly Phe Thr Phe Lys Arg Thr Ser Gly Ser 325 330 335 Ser Val Lys Arg Glu Glu Glu Val Leu Thr Gly Asn Leu Gln Thr Leu 350 Lys Ile Arg Val His Glu Gly Tyr Glu Glu Phe Thr Met Val Gly Lys 355 360 365 Arg Ala Thr Ala Ile Leu Arg Lys Ala Thr Arg Arg Leu Ile Gln Leu 380 Ile Val Ser Gly Arg Asp Glu Gln Ser Ile Ala Glu Ala Ile Ile Val 385 390 395 400 Ala Met Val Phe Ser Gln Glu Asp Cys Met Ile Lys Ala Val Arg Gly 405 410

Asp Leu Asn Phe Val Asn Arg Ala Asn Gln Arg Leu Asn Pro Met His 425 Gln Leu Leu Arg His Phe Gln Lys Asp Ala Lys Val Leu Phe Gln Asn 435 445 Trp Gly Ile Glu His Ile Asp Asn Val Met Gly Met Ile Gly Val Leu Pro Asp Met Thr Pro Ser Thr Glu Met Ser Met Arg Gly Val Arg Val 475 Ser Lys Met Gly Val Asp Glu Tyr Ser Ser Ala Glu Arg Val Val Val 485 Ser Ile Asp Arg Phe Leu Arg Val Arg Asp Gln Arg Gly Asn Val Leu 505 Leu Ser Pro Glu Glu Val Ser Glu Thr Gln Gly Thr Glu Lys Leu Thr Ile Thr Tyr Ser Ser Ser Met Met Trp Glu Ile Asn Gly Pro Glu Ser 530 535 Val Leu Val Asn Thr Tyr Gln Trp Ile Ile Arg Asn Trp Glu Thr Val 550 555 560 545 Lys Ile Gln Trp Ser Gln Asn Pro Thr Met Leu Tyr Asn Lys Met Glu 565 5/0 Phe Glu Pro Phe Gln Ser Leu Val Pro Lys Ala Ile Arg Gly Gln Tyr 590 580 Ser Gly Phe Val Arg Thr Leu Phe Gln Gln Met Arg Asp Val Leu Gly 600 605 Thr Phe Asp Thr Thr Gln Ile Ile Lys Leu Leu Pro Phe Ala Ala Ala 615 Pro Pro Lys Gln Ser Arg Met Gln Phe Ser Ser Leu Thr Val Asn Val 640 625 630 Arg Gly Ser Gly Met Arg Ile Leu Val Arg Gly Asn Ser Pro Ile Phe 645 650 Asn Tyr Asn Lys Thr Thr Lys Arg Leu Thr Ile Leu Gly Lys Asp Ala 665 Gly Thr Leu Thr Glu Asp Pro Asp Glu Gly Thr Ser Gly Val Glu Ser 675 Ala Val Leu Arg Gly Phe Leu Ile Leu Gly Lys Glu Asp Arg Arg Tyr 695 690

Gly Pro Ala Leu Ser Ile Asn Glu Leu Ser Asn Leu Ala Lys Gly Glu 705 710 715 720

Lys Ala Asn Val Leu Ile Gly Gln Gly Asp Val Val Leu Val Met Lys 725 730 735

Arg Lys Arg Asn Ser Ser Ile Leu Thr Asp Ser Gln Thr Ala Thr Lys
740 745 750

Arg Ile Arg Met Ala Ile Asn 755

### (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1773 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: HA
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(144, "u")
  - (D) OTHER INFORMATION: /gene= "HA" /note= "u in ca "master" strain; a in w2(3)" /citation= ([1])

### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(455, "a")
- (D) OTHER INFORMATION: /gene= "HA"

/note= "a in ca "master" strain; g in

wt2(3)"

/citation= ([1])

#### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(729, "c")
- (D) OTHER INFORMATION: /gene= "HA"

/note= "c in ca "master" strain; a in

wt2(3)"

/citation= ([1])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 44..1729
- (D) OTHER INFORMATION: /product= "hemagglutinin"

/gene= "HA"

/note= "hemagglutinin protein"

/citation= ([1])

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Herlocher, M L

Maassab, H F Webster, R G

(B) TITLE: Molecular and biological changes in the cold adapted

master strain A/AA/6/60 (H2N2) influenza virus

(C) JOURNAL: Proceedings of the National Academy of Sciences of

the USA

- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:17: FROM 1 TO 1773

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# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

AGCAAAAGCA G	GGGUUAUAC C	AUAGACAAC CA	AAAGCAAA	GCC AUC Ala Ile	55
UAU CUC AUU Tyr Leu Ile 5					103
GGA UAC CAU Gly Tyr His					151
CGG AAC GUC Arg Asn Val			Asp Ile		199
AAC GGA AAG Asn Gly Lys . 55					247
GAC UGU AGC Asp Cys Ser 70					295
CUU CUA AGU Leu Leu Ser 85					343
AGA AAC GGU Arg Asn Gly					391
AAA CAU CUC Lys His Leu					439
CCC AAA GAU Pro Lys Asp 135					487
UGC GCG GUG Cys Ala Val			Phe Arg		535
ACA GAG GAA Thr Glu Glu 165					583
ACA AGC GGA Thr Ser Gly					631

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GAG Glu								679
GUA Va 1								727
AGG Arg 230								775
CUC Leu								823
AUU Ile								871
AUC Ile								919
ACU Thr								967
CCA Pro 310								1015
GUC Val								1063
UUG Leu								1111
GUU Val								1159
UAU Tyr								1207
AAC Asn 390								1255
GUU Val								1303

		AAG Lys														1351
		CUA Leu														1399
		GUC Val 455														1447
AAC Asn	GUC Val 470	AAA Lys	GAA Glu	CUA Leu	GGA Gly	AAU Asn 475	GGA Gly	UGU Cys	UUU Phe	GAA Glu	UUU Phe 480	UAU Tyr	CAC His	AAA Lys	UGU Cys	1495
		GAA Glu														1543
AAG Lys	UAU Tyr	GAA Glu	GAA G1u	GAG Glu 505	UCU Ser	AAA Lys	CUA Leu	AAU Asn	AGA Arg 510	AAU Asn	GAA Glu	AUU Ile	AAA Lys	GGG Gly 515	GUA Val	1591
		AGC Ser														1639
		GGU Gly 535														1687
		UGC Cys														1729
UGA	JUAUA	AAG l	JCAUL	JUUAl	JA AL	JUAAA	\AAC#	A CCO	CUUGL	JUUC	UACI	J				1773

# (2) INFORMATION FOR SEQ ID NO:18:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 562 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Ala Ile Ile Tyr Leu Ile Leu Leu Phe Thr Ala Val Arg Gly Asp Lys Ile Cys Ile Gly Tyr His Ala Asn Asn Ser Thr Glu Thr Val Asp Thr Ile Leu Glu Arg Asn Val Thr Val Thr His Ala Lys Asp Ile Leu 40 Glu Lys Thr His Asn Gly Lys Leu Cys Lys Leu Asn Gly Ile Pro Pro 50 55 60 Leu Glu Leu Gly Asp Cys Ser Ile Ala Gly Trp Leu Leu Gly Asn Pro 65 70 75 80 Glu Cys Asp Arg Leu Leu Ser Val Pro Glu Trp Ser Tyr Ile Met Glu Lys Glu Asn Pro Arg Asn Gly Leu Cys Tyr Pro Gly Asn Phe Asn Asp Tyr Glu Glu Leu Lys His Leu Leu Ser Ser Val Lys His Phe Glu Lys 125 Val Lys Ile Leu Pro Lys Asp Arg Trp Thr Gln His Thr Thr Gly 130 135 140 Gly Ser Gln Ala Cys Ala Val Ser Gly Asn Pro Ser Phe Phe Arg Asn 150 155 Met Val Trp Leu Thr Glu Glu Gly Ser Asn Tyr Pro Val Ala Lys Gly 170 165 Ser Tyr Asn Asn Thr Ser Gly Glu Gln Met Leu Ile Ile Trp Gly Val 180 185 His His Pro Ile Asp Glu Thr Glu Gln Arg Thr Leu Tyr Gln Asn Val 200 205 Gly Thr Tyr Val Ser Val Gly Thr Ser Thr Leu Asn Lys Arg Ser Thr 210 215 220 210 Pro Glu Ile Ala Thr Arg Pro Lys Val Asn Gly Leu Gly Ser Arg Met 235 225 Glu Phe Ser Trp Thr Leu Leu Asp Met Trp Asp Thr Ile Asn Phe Glu 245 Ser Thr Gly Asn Leu Ile Ala Pro Glu Tyr Gly Phe Lys Ile Ser Lys 260 Arg Gly Ser Ser Gly Ile Met Lys Thr Glu Gly Thr Leu Glu Asn Cys 275 280 285

Glu Thr Lys Cys Gln Thr Pro Leu Gly Ala Ile Asn Thr Thr Leu Pro 295 Phe His Asn Val His Pro Leu Thr Ile Gly Glu Cys Pro Lys Tyr Val 315 305 Lys Ser Glu Lys Leu Val Leu Ala Thr Gly Leu Arg Asn Val Pro Gln 330 325 Ile Glu Ser Arg Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Gly Gly Trp Gln Gly Met Val Asp Gly Trp Tyr Gly Tyr His His Ser Asn 355 Asp Gln Gly Ser Gly Tyr Ala Ala Asp Lys Glu Ser Thr Gln Lys Ala 375 380 Phe Asp Gly Ile Thr Asn Lys Val Asn Ser Val Ile Glu Lys Ile Asn 390 395 Thr Gln Phe Glu Ala Val Gly Lys Glu Phe Ser Asn Leu Glu Arg Arg Leu Glu Asn Leu Asn Lys Lys Met Glu Asp Gly Phe Leu Asp Val Trp 425 Thr Tyr Asn Ala Glu Leu Leu Val Leu Met Glu Asn Glu Arg Thr Leu 440 Asp Phe His Asp Ser Asn Val Lys Asn Leu Tyr Asp Lys Val Arg Met Gln Leu Arg Asp Asn Val Lys Glu Leu Gly Asn Gly Cys Phe Glu Phe Tyr His Lys Cys Asp Asp Glu Cys Met Asn Ser Val Lys Asn Gly Thr 490 Tyr Asp Tyr Pro Lys Tyr Glu Glu Glu Ser Lys Leu Asn Arg Asn Glu 500 505 Ile Lys Gly Val Lys Leu Ser Ser Met Gly Val Cys Arg Ile Leu Ala 520 Ile Tyr Ala Thr Val Ala Gly Ser Leu Ser Leu Ala Ile Met Met Ala Gly Ile Ser Phe Trp Met Cys Ser Asn Gly Ser Leu Gln Cys Arg Ile 560

Cys Ile

- (2) INFORMATION FOR SEQ ID NO:19:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1467 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: RNA (genomic)
    - (vi) ORIGINAL SOURCE:
      - (A) ORGANISM: Influenza virus
      - (B) STRAIN: cold-adapted "Master Strain" A/Ann Arbor/6/60 7PI (H2N2)
  - (vii) IMMEDIATE SOURCE:
    - (B) CLONE: NA
  - (ix) FEATURE:
    - (A) NAME/KEY: mutation
    - (B) LOCATION: replace(394, "u")
    - (D) OTHER INFORMATION: /product= "Neuraminidase"

/gene= "NA"

/note= "u in ca "master" strain; c in wt2(3)"

/citation=([1])

- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(604, "u")
  - (D) OTHER INFORMATION: /product= "Neuraminidase"

/gene= "NA" /note= "u in ca "master" strain; a in

wt2(3)"

/citation=([1])

(	ix	)	F	F۵	T	H	R	F	
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(A) NAME/KEY: CDS

(B) LOCATION: 20..1426

(D) OTHER INFORMATION: /product= "neuraminidase"

/gene= "NA"

/note= "neuraminidase protein"

/citation=([1])

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G

- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) Influenza Virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:19: FROM 1 TO 1467

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

AGCAAAA	GCA (	GGAGI	JGAA			n Glr			e Gly	52
UCU GUC Ser Val										100
AUC CUG Ile Leu										148
UCC CCC Ser Pro 45	Ala									196
AGG AAC Arg Asn 60										244
GAG AUU Glu Ile										292

	CAA Gln															340
	UCU Ser															388
	GAU Asp 125															436
	GAC. Asp															484
	ACC Thr															532
	CAA Gln															580
	UGG Trp															628
	UUC Phe 205															676
	AAU Asn															724
	UGC Cys															772
ACU Thr	AGA Arg	AUA Ile	CUA Leu 255	UUC Phe	AUU Ile	AAA Lys	GAG Glu	GGG Gly 260	AAA Lys	AUU Ile	GUC Val	CAU His	AUU Ile 265	GGC Gly	CCA Pro	820
	UCA Ser															868
	CCU Pro 285															916
	CCC Pro															964

UAU GUG UGC UCA GGG CUU GUU GGC GAC ACA CCC AGG AAC GAC GAC ACA Tyr Val Cys Ser Gly Leu Val Gly Asp Thr Pro Arg Asn Asp Asp 320 325 330											
UCU AGC AAU AGC AAU UGC AGG GAU CCU AAC AAU GAG AGA GGG AAU ( Ser Ser Asn Ser Asn Cys Arg Asp Pro Asn Asn Glu Arg Gly Asn B 335 340 345											
GGA GUG AAA GGC UGG GCC UUU GAC AAU GGA GAU GAU GUA UGG AUG (Gly Val Lys Gly Trp Ala Phe Asp Asn Gly Asp Asp Val Trp Met (350 350 360											
AGA ACA AUC AGC AAA GAU UUA CGC UCA GGU UAU GAA ACU UUC AAA (Arg Thr Ile Ser Lys Asp Leu Arg Ser Gly Tyr Glu Thr Phe Lys No. 365 370 375											
AUU GGU GGU UGG UCC ACA CCU AAU UCC AAA UCG CAG GUC AAU AGA (Ile Gly Gly Trp Ser Thr Pro Asn Ser Lys Ser Gln Val Asn Arg (380 390 390											
GUC AUA GUU GAC AAC AAU AAU UGG UCU GGU UAC UCU GGU AUU UUC UVAl Ile Val Asp Asn Asn Asn Trp Ser Gly Tyr Ser Gly Ile Phe 9400 405	UCU 1252 Ser										
GUU GAG GGC AAA AGC UGC AUC AAU AGG UGC UUU UAU GUG GAG UUG AVAI Glu Gly Lys Ser Cys Ile Asn Arg Cys Phe Tyr Val Glu Leu 1415 420 425											
AGG GGA AGG CCA CAG GAG ACU AGA GUA UGG UGG ACC UCA AAC AGU AAR Gly Arg Pro Gln Glu Thr Arg Val Trp Trp Thr Ser Asn Ser 430 435 440	AUU · 1348 Ile										
GUU GUA UUU UGU GGC ACU UCA GGU ACU UAU GGA ACA GGC UCA UGG (Val Val Phe Cys Gly Thr Ser Gly Thr Tyr Gly Thr Gly Ser Trp 4450 455											
GAU GGG GCG AAC AUC AAU UUC AUG CCU AUA UAACGUUUCG CAAUUUUAGAAsp Gly Ala Asn Ile Asn Phe Met Pro Ile	A 1446										
AAAAAACUCC UUGUUUCUAC U											

# (2) INFORMATION FOR SEQ ID NO:20:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 469 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Asn Pro Asn Gln Lys Thr Ile Thr Ile Gly Ser Val Ser Leu Thr Ile Ala Thr Val Cys Phe Leu Met Gln Ile Ala Ile Leu Ala Thr Thr 20 25 Val Thr Leu His Leu Lys Gln His Glu Cys Asp Ser Pro Ala Ser Asn Gln Val Met Pro Cys Glu Pro Ile Ile Ile Glu Arg Asn Ile Thr Glu Ile Val Tyr Leu Asn Asn Thr Thr Ile Glu Lys Glu Ile Cys Pro Glu 65 70 75 80 Val Val Gly Tyr Arg Asn Trp Ser Lys Pro Gln Cys Gln Ile Thr Gly Phe Ala Pro Phe Ser Lys Asp Asn Ser Ile Arg Leu Ser Ala Gly Gly 105 110 Asp Ile Trp Val Thr Arg Glu Pro Tyr Val Ser Cys Asp Pro Gly Lys 115 120 125 Cys Tyr Gln Phe Ala Leu Gly Gln Gly Thr Thr Leu Asp Asn Lys His 135 Ser Asn Gly Thr Ile His Asp Arg Ile Pro His Arg Thr Leu Leu Met 145 155 160 Asn Glu Leu Gly Val Pro Phe His Leu Gly Thr Lys Gln Val Cys Ala 165 Ala Trp Ser Ser Ser Ser Cys His Asp Gly Lys Ala Trp Leu His Val 185 Cys Val Thr Gly Asp Asp Arg Asn Ala Thr Ala Ser Phe Ile Tyr Asp 205 195 Gly Lys Leu Val Asp Ser Ile Gly Ser Trp Ser Gln Asn Val Leu Arg Thr Gln Glu Ser Glu Cys Val Cys Ile Asn Gly Thr Cys Thr Val Val 225 230 235 240 Met Thr Asp Gly Ser Ala Ser Gly Arg Ala Asp Thr Arg Ile Leu Phe 245 250 255 Ile Lys Glu Gly Lys Ile Val His Ile Gly Pro Leu Ser Gly Ser Ala 260 265 270

Gln His Val Glu Glu Cys Ser Cys Tyr Pro Arg Tyr Pro Asp Val Arg 275 280 285

Cys Ile Cys Arg Asp Asn Trp Lys Gly Ser Asn Arg Pro Val Ile Asp 290 295 300

Ile Asn Met Glu Asp Tyr Ser Ile Asp Ser Ser Tyr Val Cys Ser Gly 305 310 315

Leu Val Gly Asp Thr Pro Arg Asn Asp Ser Ser Ser Asn Ser Asn 325 330 335

Cys Arg Asp Pro Asn Asn Glu Arg Gly Asn Pro Gly Val Lys Gly Trp 340 345 350

Ala Phe Asp Asn Gly Asp Asp Val Trp Met Gly Arg Thr Ile Ser Lys 355 360 365

Asp Leu Arg Ser Gly Tyr Glu Thr Phe Lys Val Ile Gly Gly Trp Ser 370 380

Thr Pro Asn Ser Lys Ser Gln Val Asn Arg Gln Val Ile Val Asp Asn 385 390 - 395 400

Asn Asn Trp Ser Gly Tyr Ser Gly Ile Phe Ser Val Glu Gly Lys Ser 405 410 415

Cys Ile Asn Arg Cys Phe Tyr Val Glu Leu Ile Arg Gly Arg Pro Gln
420 425 430

Glu Thr Arg Val Trp Trp Thr Ser Asn Ser Ile Val Val Phe Cys Gly 435 440 445

Thr Ser Gly Thr Tyr Gly Thr Gly Ser Trp Pro Asp Gly Ala Asn Ile 450 455 460

Asn Phe Met Pro Ile 465

### (2) INFORMATION FOR SEQ ID NO:21:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 890 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)

# (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Influenza virus
- (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)

### (vii) IMMEDIATE SOURCE:

(B) CLONE: NS

#### (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 27..56
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2" /citation= ([1][2])

### (ix) FEATURE:.

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(483, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3); g in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 529..861
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(813, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in

wt2(3); a in 1988 reported wild type
E28-32 strain"
/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: join(27..56, 529..861)
- (D) OTHER INFORMATION: /product= "nonstructural protein NS2" /gene= "NS" /note= "nonstructural protein NS2"

/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 27..677
- (D) OTHER INFORMATION: /product= "nonstructural protein NS1" /gene= "NS" /note= "nonstructural protein NS1" /citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:21: FROM 1 TO 890

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox. N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

-. • • • . . ,

(B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60(H2N2)

(C) JOURNAL: Virology

(D) VOLUME: 167

(F) PAGES: 554-567

(G) DATE: 1988

(K) RELEVANT RESIDUES IN SEQ ID NO:21: FROM 1 TO 890

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

AGCAAAAGCA GGG	GUGACAAA GACAUA	A AUG GAU CCU AA Met Asp Pro As 1	C ACU GUG UCA AGC n Thr Val Ser Ser 5	UUU 53 Phe
			CAA GUU GCA GAC Gln Val Ala Asp	
			CGC CGA GAU CAG Arg Arg Asp Gln 40	
Ser Leu Arg G			AAC AUC GAA ACA Asn Ile Glu Thr 55	
			CUG AAG GAA GAA Leu Lys Glu Glu 70	
			CCU GCU UCG CGA Pro Ala Ser Arg 85	
			GAC UGG UUC AUG Asp Trp Phe Met	
AUG CCC AAG C Met Pro Lys G	AG AAA GUG GCA in Lys Val Ala 110	GGC CCU CUU UGU Gly Pro Leu Cys 115	AUC AGA AUG GAC Ile Arg Met Asp 120	CAG 389 Gln
Ala Ile Met A	AU AAG AAC AUC Asp Lys Asn Ile 25	AUA UUG AAA GCG Ile Leu Lys Ala 130	AAU UUC AGU GUG Asn Phe Ser Val 135	AUU 437 Ile

UUU Phe	Asp															485
GGA Gly																533
ACU Thr 170											Leu					581
GAA Glu	UGG Trp	AAU Asn	GAU Asp	AAC Asn 190	ACA Thr	GUU Val	CGA Arg	GUC Val	UCU Ser 195	AAA Lys	ACU Thr	CUA Leu	CAG Gln	AGA Arg 200	UUC Phe	629
GCU Ala															AAA Lys	677
UAGA	AACG	GA A	\AAU(	GCGA	AG AA	ACAAL	JUAGO	G UCA	<b>\</b> AAA(	SUUC	GAA	BAAAI	JAA (	GAUG(	GCUGAU	737
UGAA	GAAG	aug A	AGACA	ACAA/	AU UG	SAAGA	AUAA(	C AGA	AGAAL	JAGU	UUU	GAGC	4 <b>4</b> 4 l	JAACA	UAUUUAU	797
GCAA	GCCL	JUA (	CAGCU	JGCUA	AU UL	JGAAC	GUGGA	A ACA	\AGA(	AUA	AGA	ACUUI	JCU (	CGUU	JCAGCU	857
UAUU	UAAL	JGA l	JAAA	AAACA	AC CO	CUUGL	JUUCI	J ACI	J							890

# (2) INFORMATION FOR SEQ ID NO:22:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 217 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp  $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$ 

His Val Arg Lys Gln Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 25 30

Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser 35 40 45

Thr Leu Gly Leu Asn Ile Glu Thr Ala Thr Arg Val Gly Lys Gln Ile 50 55 60

Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 65 70 75 80

Met Ala Ser Ala Pro Ala Ser Arg Tyr Leu Thr Asp Met Thr Ile Glu 85 90 95

Glu Met Ser Arg Asp Trp Phe Met Leu Met Pro Lys Gln Lys Val Ala 100 105 110

Gly Pro Leu Cys Ile Arg Met Asp Gln Ala Ile Met Asp Lys Asn Ile 115 120 125

Ile Leu Lys Ala Asn Phe Ser Val Ile Phe Asp Arg Leu Glu Thr Leu 130 140

Ile Leu Leu Arg Ala Phe Thr Glu Thr Gly Ala Ile Val Gly Glu Ile 145 150 155 160

Ser Pro Leu Pro Ser Leu Pro Gly His Thr Asn Glu Asp Val Lys Asn 165 170 175

Ala Ile Gly Val Leu Ile Gly Gly Leu Glu Trp Asn Asp Asn Thr Val 180 185 190

Arg Val Ser Lys Thr Leu Gln Arg Phe Ala Trp Arg Ser Ser Asp Glu 195 200 205

Asn Gly Arg Pro Pro Leu Thr Pro Lys 210 215

### (2) INFORMATION FOR SEQ ID NO:23:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 418 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

#### (ii) MOLECULE TYPE: RNA (genomic)

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 27..389

(D) OTHER INFORMATION: /product= "Nonstructural protein 2" /gene= "NS2"

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

AGCAAAAGCA GGGUGACAAA GACAUA AUG GAU CCU AAC ACU GUG UCA AGC UUU Met Asp Pro Asn Thr Val Ser Ser Phe 1 5	53
CAG GAC AUA CUA AUG AGG AUG UCA AAA AUG CAA UUG GGG UCC UCA UCG Gln Asp Ile Leu Met Arg Met Ser Lys Met Gln Leu Gly Ser Ser Ser 10 20 25	101
GAG GAC UUG AAU GGA AUG AUA ACA CAG UUC GAG UCU CUA AAA CUC UAC Glu Asp Leu Asn Gly Met Ile Thr Gln Phe Glu Ser Leu Lys Leu Tyr 30 35 40	149
AGA GAU UCG CUU GGA GAA GCA GUG AUG AGA AUG GGA GAC CUC CAC UCA Arg Asp Ser Leu Gly Glu Ala Val Met Arg Met Gly Asp Leu His Ser 45	197
CUC CAA AAU AGA AAC GGA AAA UGG CGA GAA CAA UUA GGU CAA AAG UUC Leu Gln Asn Arg Asn Gly Lys Trp Arg Glu Gln Leu Gly Gln Lys Phe 60 65 70	245
GAA GAA AUA AGA UGG CUG AUU GAA GAA GUG AGA CAC AAA UUG AAG AUA Glu Glu Ile Arg Trp Leu Ile Glu Glu Val Arg His Lys Leu Lys Ile 75 80 85	293
ACA GAG AAU AGU UUU GAG CAA AUA ACA UUU AUG CAA GCC UUA CAG CUG Thr Glu Asn Ser Phe Glu Gln Ile Thr Phe Met Gln Ala Leu Gln Leu 90 95 100 105	341
CUA UUU GAA GUG GAA CAA GAG AUA AGA ACU UUC UCG UUU CAG CUU AUU Leu Phe Glu Val Glu Gln Glu Ile Arg Thr Phe Ser Phe Gln Leu Ile 110 115 120	389
UAAUGAUAAA AAACACCCUU GUUUCUACU	418

# (2) INFORMATION FOR SEQ ID NO:24:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 121 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Asp Ile Leu Met Arg Met 1 5 10 15

Ser Lys Met Gln Leu Gly Ser Ser Ser Glu Asp Leu Asn Gly Met Ile 20 25 30

Thr Gln Phe Glu Ser Leu Lys Leu Tyr Arg Asp Ser Leu Gly Glu Ala 35 40 45

Val Met Arg Met Gly Asp Leu His Ser Leu Gln Asn Arg Asn Gly Lys 50 55 60

Trp Arg Glu Gln Leu Gly Gln Lys Phe Glu Glu Ile Arg Trp Leu Ile
65 70 75 80

Glu Glu Val Arg His Lys Leu Lys Ile Thr Glu Asn Ser Phe Glu Gln 85 90 95

Ile Thr Phe Met Gln Ala Leu Gln Leu Leu Phe Glu Val Glu Gln Glu 100 105 110

Ile Arg Thr Phe Ser Phe Gln Leu Ile 115 120

# (2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1027 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)

#### (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Influenza virus
- (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)

### (vii) IMMEDIATE SOURCE:

(B) CLONE: M

- (A) NAME/KEY: exon
- (B) LOCATION: 26..51
- (D) OTHER INFORMATION: /product= "matrix protein M2"

/gene= "M"

/note= "matrix protein M2"

/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: exon
- (B) LOCATION: 740..1004
- (D) OTHER INFORMATION: /product= "matrix protein M2"

/gene= "M"

/note= "matrix protein M2"

/citation= ([1][2])

# (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(969, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in

wt2(3); g in 1988 reported wild type

E28-32 strain"

/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: join(26..51, 740..1004)
- (D) OTHER INFORMATION: /product= "matrix protein M2"

/gene= "M"

/note= "matrix protein M2"

/citation= ([1][2])

- (A) NAME/KEY: CDS
- (B) LOCATION: 26..781
- (D) OTHER INFORMATION: /product= "matrix protein M1" /gene= "M" /note= "matrix protein M1" /citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:25: FROM 1 TO 1027

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Cox. N J
  Kitame, F
  Kendal, A P
  Maassab, H F
  Naeve, C
- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60(H2N2)
- (C) JOURNAL: Virology
- (D) VOLUME: 167
- (F) PAGES: 554-557
- (G) DATE: 1988
- (K) RELEVANT RESIDUES IN SEQ ID NO:25: FROM 1 TO 1027.

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# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

AGCAAAAGCA GGUAGAUAUU GAAAG	AUG AGU ( Met Ser L	CUU CUA ACC Leu Leu Thr	GAG GUC GAA Glu Val Glu	ACG 52 Thr
UAC GUU CUC UCU AUC AUC CCG Tyr Val Leu Ser Ile Ile Pro 10 15				
CAG AGA CUU GAA GAU GUC UUU Gln Arg Leu Glu Asp Val Phe 30				
CUC AUG GAA UGG CUA AAG ACA Leu Met Glu Trp Leu Lys Thr 45				
GGG AUU UUG GGA UUU GUA UUC Gly Ile Leu Gly Phe Val Phe 60				
CUG CAG CGU AGA CGC UUU GUC Leu Gln Arg Arg Arg Phe Val 75 80	CAA AAU ( Gln Asn A	GCC CUC AAU Ala Leu Asn 85	GGG AAU GGG Gly Asn Gly	GAU 292 Asp
CCA AAU AAC AUG GAC AGA GCA Pro Asn Asn Met Asp Arg Ala 90 95				
GAG AUA ACA UUC CAU GGG GCC Glu Ile Thr Phe His Gly Ala 110	Lys Glu			
GGU GCA CUU GCC AGU UGU AUG Gly Ala Leu Ala Ser Cys Met 125				
GUG ACC ACU GAA GUG GUC UUA Val Thr Thr Glu Val Val Leu 140				
AUU GCU GAC UCC CAG CAU AGG Ile Ala Asp Ser Gln His Arg 155 160	Ser His /	AGG CAA AUG Arg Gln Met 165	GUG ACA ACA Val Thr Thr	ACC 532 Thr
AAU CCA CUA AUA AGA CAU GAG Asn Pro Leu Ile Arg His Glu 170 175	AAC AGA A Asn Arg I	AUG GUU CUG Met Val Leu ·180	GCC AGC ACU Ala Ser Thr	ACA 580 Thr 185
GCU AAG GCU AUG GAG CAA AUG Ala Lys Ala Met Glu Gln Met 190	Ala Gly	UCG AGU GAG Ser Ser Glu 195	CAA GCA GCA Gln Ala Ala 200	GAG 628 Glu

GCC Ala	AUG Met	GAG Glu	GUU Val 205	GCU Ala	AGU Ser	CAG Gln	GCC Ala	AGG Arg 210	CAA Gln	AUG Met	GUG Val	CAG Gln	GCA Ala 215	AUG Met	AGA Arg		676
GUU Val	AUU Ile	GGG Gly 220	ACU Thr	CAU His	CCU Pro	AGC Ser	UCC Ser 225	AGU Ser	GCU Ala	GGU Gly	CUA Leu	AAA Lys 230	AAU Asn	GAU Asp	CUU Leu		724
CUU Leu	GAA Glu 235	AAU Asn	UUG Leu	CAG Gln	GCC Ala	UAU Tyr 240	CAG Gln	AAA Lys	CGA Arg	AUG Met	GGG Gly 245	GUG Val	CAG Gln	AUG Met	CAA Gln		772
	Phe		UGA	CCCU(	CUU (	GUUGI	JUGC(	CG C(	GAGU/	AUCAL	J UG(	GGAU(	CUUG				821
CAC	UUGA	JAU I	UGUG	GAUU	CU U	GAUC/	AUCUI	J UUI	UUUC	<b>AAA</b> U	GCA	UUUA	UCG	CUUCI	JUUAA	А	881
CAC	GGUC	UGA .	AAAG,	AGGG	CC UI	JCUA(	CGGA	A GG/	AGUA	CCAG	AGU	CUAU	GAG	GGAA	GAAUA	.U	941
CGA	AAGG,	AAC .	AGCA	GAGU	GC U	GUGG/	AUUC	J GA	CGAU	AGUC	AUU	UUGU	CAG	CAUA	GAGCU	G :	1001
GAG	UAAA	AAA	CUAC	CUUG	UU U	CUAC	J									•	1027

# (2) INFORMATION FOR SEQ ID NO:26:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 252 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Ser Leu Leu Thr Glu Val Glu Thr Tyr Val Leu Ser Ile Ile Pro 1 5 10 15

Ser Gly Pro Leu Lys Ala Glu Ile Ala Gln Arg Leu Glu Asp Val Phe 20 25 30

Ala Gly Lys Asn Thr Asp Leu Glu Ala Leu Met Glu Trp Leu Lys Thr 35 40 45

Arg Pro Ile Leu Ser Pro Leu Thr Lys Gly Ile Leu Gly Phe Val Phe 50 55 60

Thr Leu Thr Val Pro Ser Glu Arg Gly Leu Gln Arg Arg Phe Val

65					70					75					80
Gln	Asn	Ala	Leu	Asn · 85	Gly	Asn	Gly	Asp	Pro 90	Asn	Asn	Met	Asp	Arg 95	A٦ā
Val	Lys	Leu	Tyr 100	Arg	Lys	Leu	Lys	Arg 105	Glu	Ile	Thr	Phe	His 110	Gly	Ala
Lys	Glu	Ile 115	Ala	Leu	Ser	Tyr	Ser 120	Ala	Gly	Ala	Leu	Ala 125	Ser	Cys	Met
Gly	Leu 130	Ile	Tyr	Asn	Arg	Met 135	Gly	Ala	Val	Thr	Thr 140	Glu	Val	Val	Leu
Gly 145	Leu	Val	Cys	Ala	Thr 150	Cys	Glu	Gln	Ile	Ala 155	Asp	Ser	Gln	His	Arg 160
Ser	His	Arg	Gln	Met 165	Val	Thr	Thr	Thr	Asn 170	Pro	Leu	Пe	Arg	His 175	G٦ι
Asn	Arg	Met	Val 180	Leu	Ala	Ser	Thr	Thr 185	Ala	Lys	Ala	Met	Glu 190	Gln	Met
Ala	Gly	Ser 195	Ser	Glu	Gln	Ala	Ala 200	Glu	Ala	Met	Glu	Val 205	Ala	Ser	Glr
Ala	Arg 210	Gln	Met	Val	Gln	Ala 215	Met	Arg	Val	Пe	Gly 220	Thr	His	Pro	Ser
Ser	Ser	Ala	Gly	Leu	Lys 230	Asn	Asp	Leu	Leu	G1u 235		Leu	Gln	Ala	Tyr 240

# (2) INFORMATION FOR SEQ ID NO:27:

245

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 339 base pairs

Gln Lys Arg Met Gly Val Gln Met Gln Arg Phe Lys

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: RNA (genomic)

- (A) NAME/KEY: CDS
- (B) LOCATION: 26..316
- (D) OTHER INFORMATION: /product= "Matrix M2"

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

AGCA	<b>1</b> AAA(	GCA (	GUA(	JAUAL	JU GA	\AAG	AUG Met 1							GAA G1u		52
							UGC Cys									100
							AUU Ile									148
							AAA Lys									196
							ACG Thr 65									244
							CAG Gln								_	292
	UUU Phe						GAG Glu	UAA	<b>4</b> AAA(	CUA (	CCUU	GUUU(	CU A	CU		339

# (2) INFORMATION FOR SEQ ID NO:28:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 97 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Met Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly 1 5 10 15

Cys Arg Cys Asn Asp Ser Ser Asp Pro Leu Val Val Ala Ala Ser Ile 20 25 30

Ile Gly Ile Leu His Leu Ile Leu Trp Ile Leu Asp His Leu Phe Phe 35 40 45

Lys Cys Ile Tyr Arg Phe Phe Lys His Gly Leu Lys Arg Gly Pro Ser 50 60

Thr Glu Gly Val Pro Glu Ser Met Arg Glu Glu Tyr Arg Lys Glu Gln 65 70 75 80

Gln Ser Ala Val Asp Ser Asp Asp Ser His Phe Val Ser Ile Glu Leu 85 90 95

Glu

### (2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2341 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) egg passage 2(3)

### (vii) IMMEDIATE SOURCE:

(B) CLONE: PB2

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(141, "a")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain; a in wt2(3); a in 1988 reported wild type

E28-32 strain"

/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(426, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain and in wt2(3); u in 1988 reported wild type

E28-32 strain"

/citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(714, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in

wt2(3); c in 1988 reported wild type

E28-32 strain"

/citation=([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(821, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type

E28-32 strain"

/citation=([1][2])

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(963, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1182, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1212, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); c in 1988 reported wild type E28-32 strain" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1353, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); u in 1988 reported wild type E28-32 strain" /citation= ([1][2])

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1923, "q")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in  $\underline{\text{wt2}(3)}$ ; a in 1988 reported wild type

E28-32 strain" /citation= ([1][2])

# (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(1933, "u")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain; u in wt2(3); u in 1988 reported wild type E28-32 strain" /citation=([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 28..2304
- (D) OTHER INFORMATION: /product= "polymerase basic 2" /gene= "PB2" /note= "polymerase basic 2"

/citation=([1][2])

## (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:29: FROM 1 TO 2341

# (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60(H2N2)
- (C) JOURNAL: Virology

(D) VOLUME: 167

(F) PAGES: 554-567

(G) DATE: 1988

(K) RELEVANT RESIDUES IN SEQ ID NO:29: FROM 1 TO 2341

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

AGCGAAAGCA GGUCAAUU	GCGAAAGCA GGUCAAUUAU AUUCAAU AUG GAA AGA AUA AAA GAA CUA CGG Met Glu Arg Ile Lys Glu Leu Arg 1 5												
		GAG AUA CUA ACA AAA ACC ACA Glu Ile Leu Thr Lys Thr Thr 20	99										
		UAC ACA UCA GGG AGA CAG GAA Tyr Thr Ser Gly Arg Gln Glu 35 40	147										
	Arg Met Lys Trp	AUG AUG GCA AUG AAA UAU CCG Met Met Ala Met Lys Tyr Pro 50 55	195										
		AUG AUU CCU GAG AGA AAU GAG Met Ile Pro Glu Arg Asn Glu 70	243										
CAA GGG CAA ACU CUA Gln Gly Gln Thr Leu 75	UGG AGU AAA AUG Trp Ser Lys Met 80	AGU GAU GCC GGA UCG GAU CGU Ser Asp Ala Gly Ser Asp Arg 85	291										
		UGG UGG AAU AGA AAU GGA CCA Trp Trp Asn Arg Asn Gly Pro 100	339										

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		AGU Ser														,	387
AAA Lys	GUC Val	GAA Glu	AGG Arg	UUA Leu 125	AAA Lys	CAU His	GGA Gly	ACC Thr	UUU Phe 130	GGC Gly	CCU Pro	GUC Val	CAU His	UUU Phe 135	AGA Arg		435
		GUC Val															483
		AGU Ser 155															531
		GAA Glu															579
		AAA Lys															627
		GUU Val															675
		CCA Pro														•	723
		ACU Thr 235															771
		AGG Arg															819
AGC Ser 265	AUA Ile	GUG Val	AGA Arg	AGA Arg	GCA Ala 270	GCA Ala	GUA Val	UCA Ser	GCA Ala	GAU Asp 275	CCA Pro	CUA Leu	GCA Ala	UCU Ser	UUA Leu 280		867
		AUG Met															915
		AGG Arg															963
		AUG Met 315															1011

								UCA Ser									1059
								AAA Lys									1107
								AGG Arg									1155
								AUU Ile 385									1203
								GCC Ala									1251
								GAU Asp									1299
AAU Asn 425	CAG Gln	CGA Arg	UUG Leu	AÁU Asn	CCC Pro 430	AUG Met	CAU His	CAA Gln	CUU Leu	UUA Leu 435	AGA Arg	CAU His	UUU Phe	CAG Gln	AAG Lys 440		1347
								UGG Trp									1395
								CCA Pro 465									1443
								AGC Ser								•	1491
								AGC Ser								-	1539
								CUA Leu							GAA Glu 520	•	1587
								AUA Ile									1635
								GUG Val 545									1683

AUC Ile	AUC Ile	AGA Arg 555	AAC Asn	UGG Trp	GAA Glu	ACU Thr	GUU Val 560	AAA Lys	AUU Ile	CAG Gln	UGG Trp	UCU Ser 565	CAG Gln	AAU Asn	CCU Pro	1731
		CUA Leu														1779
		GCC Ala														1827
CAA Gln	CAA Gln	AUG Met	AGG Arg	GAU Asp 605	GUA Val	CUU Leu	GGG Gly	ACA Thr	UUU Phe 610	GAU Asp	ACC Thr	ACC Thr	CAG Gln	AUA Ile 615	AUA Ile	1875
AAA Lys	CUU Leu	CUU Leu	CCC Pro 620	UUU Phe	GCA Ala	GCC Ala	GCC Ala	CCA Pro 625	CCA Pro	AAG Lys	CAA Gln	AGU Ser	AGA Arg 630	AUG Met	CAG Gln	1923
		UCA Ser 635														1971
		GGC Gly														2019
CUA Leu 665	ACA Thr	AUU Ile	CUC Leu	GGA Gly	AAG Lys 670	GAU Asp	GCU Ala	GGC Gly	ACU Thr	UUA Leu 675	ACU Thr	GAA Glu	GAC Asp	CCA Pro	GAU Asp 680	2067
GAA Glu	GGC Gly	ACA Thr	UCU Ser	GGA Gly 685	GUG Val	GAG Glu	UCC Ser	GCU Ala	GUU Va1 690	CUG Leu	AGA Arg	GGA Gly	UUC Phe	CUC Leu 695	AUU Ile	2115
CÜG Leu	GGC Gly	AAA Lys	GAA Glu 700	GAU Asp	AGG Arg	AGA Arg	UAU Tyr	GGA Gly 705	CCA Pro	GCA Ala	UUA Leu	AGC Ser	AUC Ile 710	AAU Asn	GAA Glu	2163
CUG Leu	AGU Ser	AAC Asn 715	CUU Leu	GCG Ala	AAA Lys	GGA Gly	GAA G1u 720	AAG Lys	GCU Ala	AAU Asn	GUA Val	CUA Leu 725	AUU Ile	GGG Gly	CAA Gln	2211
GGA Gly	GAC Asp 730	GUG Val	GUG Val	UUG Leu	GUA Val	AUG Met 735	AAA Lys	CGA Arg	AAA Lys	CGG Arg	AAC Asn 740	UCU Ser	AGC Ser	AUA Ile	CUU Leu	2259
		AGC Ser														2304
UAAl	JGUU(	GAA (	JAGUI	JUAA	<b>4</b> A A(	CGAC	CUUGI	J UU	CUACI	J						2341

# (2) INFORMATION FOR SEQ ID NO:30:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 759 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

# (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Met Glu Arg Ile Lys Glu Leu Arg Asn Leu Met Ser Gln Ser Arg Thr 1 5 10 15

Arg Glu Ile Leu Thr Lys Thr Thr Val Asp His Met Ala Ile Ile Lys 20 25 30

Lys Tyr Thr Ser Gly Arg Gln Glu Lys Asn Pro Ser Leu Arg Met Lys 35 40 45

Trp Met Met Ala Met Lys Tyr Pro Ile Thr Ala Asp Lys Arg Ile Thr 50 55 60

Glu Met Ile Pro Glu Arg Asn Glu Gln Gly Gln Thr Leu Trp Ser Lys 65 70 75 80

Met Ser Asp Ala Gly Ser Asp Arg Val Met Val Ser Pro Leu Ala Val 85 90 95

Thr Trp Trp Asn Arg Asn Gly Pro Met Thr Ser Thr Val His Tyr Pro 100 105 110

Lys Ile Tyr Lys Thr Tyr Phe Glu Lys Val Glu Arg Leu Lys His Gly 115 120 125

Thr Phe Gly Pro Val His Phe Arg Asn Gln Val Lys Ile Arg Arg Arg 130 135 140

Val Asp Ile Asn Pro Gly His Ala Asp Leu Ser Ala Lys Glu Ala Gln 145 150 155 160

Asp Val Ile Met Glu Val Val Phe Pro Asn Glu Val Gly Ala Arg Ile 165 170 175

Leu Thr Ser Glu Ser Gln Leu Thr Ile Thr Lys Glu Lys Lys Glu Glu 180 185 190

Leu Gln Asp Cys Lys Ile Ser Pro Leu Met Val Ala Tyr Met Leu Glu 195 200 205

Arg Glu Leu Val Arg Lys Thr Arg Phe Leu Pro Val Ala Gly Gly Thr 215 Ser Ser Val Tyr Ile Glu Val Leu His Leu Thr Gln Gly Thr Cys Trp 230 235 Glu Gln Met Tyr Thr Pro Gly Gly Glu Val Arg Asn Asp Asp Val Asp Gin Ser Leu Ile Ile Ala Ala Arg Ser Ile Val Arg Arg Ala Ala Val 265 Ser Ala Asp Pro Leu Ala Ser Leu Leu Glu Met Cys His Ser Thr Gln 285 280 Ile Gly Gly Thr Arg Met Val Asp Ile Leu Arg Gln Asn Pro Thr Glu 300 295 Glu Gln Ala Val Glu Ile Cys Lys Ala Ala Met Gly Leu Arg Ile Ser 305 Ser Ser Phe Ser Phe Gly Gly Phe Thr Phe Lys Arg Thr Ser Gly Ser 330 Ser Val Lys Arg Glu Glu Glu Val Leu Thr Gly Asn Leu Gln Thr Leu 345 Lys Ile Arg Val His Glu Gly Tyr Glu Glu Phe Thr Met Val Gly Lys Arg Ala Thr Ala Ile Leu Arg Lys Ala Thr Arg Arg Leu Ile Gln Leu 380 Ile Val Ser Gly Arg Asp Glu Gln Ser Ile Ala Glu Ala Ile Ile Val 395 Ala Met Val Phe Ser Gln Glu Asp Cys Met Ile Lys Ala Val Arg Gly 405 Asp Leu Asn Phe Val Asn Arg Ala Asn Gln Arg Leu Asn Pro Met His 420 Gln Leu Leu Arg His Phe Gln Lys Asp Ala Lys Val Leu Phe Gln Asn Trp Gly Ile Glu His Ile Asp Asn Val Met Gly Met Ile Gly Val Leu 455 460 Pro Asp Met Thr Pro Ser Thr Glu Met Ser Met Arg Gly Val Arg Val 470 475 480 465 Ser Lys Met Gly Val Asp Glu Tyr Ser Ser Ala Glu Arg Val Val Val 485 490

Ser Ile Asp Arg Phe Leu Arg Val Arg Asp Gln Arg Gly Asn Val Leu 500 505 510 Leu Ser Pro Glu Glu Val Ser Glu Thr Gln Gly Thr Glu Lys Leu Thr 520 Ile Thr Tyr Ser Ser Ser Met Met Trp Glu Ile Asn Gly Pro Glu Ser 535 Val Leu Val Asn Thr Tyr Gln Trp Ile Ile Arg Asn Trp Glu Thr Val 560 545 Lys Ile Gln Trp Ser Gln Asn Pro Thr Met Leu Tyr Asn Lys Met Glu 565 Phe Glu Pro Phe Gln Ser Leu Val Pro Lys Ala Ile Arg Gly Gln Tyr Ser Gly Phe Val Arg Thr Leu Phe Gln Gln Met Arg Asp Val Leu Gly 595 Thr Phe Asp Thr Thr Gln Ile Ile Lys Leu Leu Pro Phe Ala Ala Ala 615 Pro Pro Lys Gln Ser Arg Met Gln Phe Ser Ser Leu Thr Val Asn Val Arg Gly Ser Gly Met Arg Ile Leu Val Arg Gly Asn Ser Pro Ile Phe 645 Asn Tyr Asn Lys Thr Thr Lys Arg Leu Thr Ile Leu Gly Lys Asp Ala 665 Gly Thr Leu Thr Glu Asp Pro Asp Glu Gly Thr Ser Gly Val Glu Ser Ala Val Leu Arg Gly Phe Leu Ile Leu Gly Lys Glu Asp Arg Arg Tyr 690 Gly Pro Ala Leu Ser Ile Asn Glu Leu Ser Asn Leu Ala Lys Gly Glu Lys Ala Asn Val Leu Ile Gly Gln Gly Asp Val Val Leu Val Met Lys 730 Arg Lys Arg Asn Ser Ser Ile Leu Thr Asp Ser Gln Thr Ala Thr Lys 750 740 Arg Ile Arg Met Ala Ile Asn 755

### (2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2341 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: PB1
- (ix) FEATURE:
  - (A) NAME/KEY: conflict
  - (B) LOCATION: replace(123, "g")
  - (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2]).

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(486, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); c in 1988 reported wild type E28-32 strain" /citation= ([1][2])

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1195, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(1276, "a")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain; a in wt2(3); g in 1988 reported wild type E28-32 strain" /citation= ([1][2])

### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1395, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); g in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1766, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2])

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2005, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3); g in 1988 reported wild type

E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2019, "u")
- (D) OTHER INFORMATION: /note= "u in ca "master" strain and in wt2(3); c in 1988 reported wild type

E28-32 strain"

/citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 25..2295
- (D) OTHER INFORMATION: /product= "polymerase basic 1"

/gene= "PB1"

/note= "polymerase basic 1"

/citation= ([1][2])

# (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:31: FROM 1 TO 2341

# (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain
- (C) JOURNAL: Virology
- (D) VOLUME: 167
- (F) PAGES: 554-567
- (G) DATE: 1988
- (K) RELEVANT RESIDUES IN SEQ ID NO:31: FROM 1 TO 2341

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

AGC(	BAAA(	GCA (	GGCA	AACC/	AU UI	AUG ( Met / 1					€.	51
	AAA Lys											99
	GAU Asp											147
	GUC Val											195
	ACG Thr											243
	GAG Glu 75											291
	GCA Ala											339

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UCG Ser	UGU Cys	CUU Leu	GAA Glu	ACG Thr 110	AUG Met	GAA Glu	GUU Val	AUU Ile	CAA Gln 115	CAA Gln	ACA Thr	AGA Arg	GUG Val	GAC Asp 120	AAA Lys	3	87
														AAU Asn		4.	35
														UCG Ser		4	83
														AAG Lys		5	31
GUG Val 170	AUA Ile	GAA Glu	UCA Ser	AUG Met	GAU Asp 175	AAA Lys	GAG Glu	GAG Glu	AUG Met	GAA Glu 180	AUC Ile	ACA Thr	ACA Thr	CAC His	UUC Phe 185	5	79
CAA Gln	AGA Arg	AAA Lys	AGA Arg	AGA Arg 190	GUA Val	AGA Arg	GAC Asp	AAC Asn	AUG Met 195	ACC Thr	AAG Lys	AAA Lys	AUG Met	GUC Val 200	ACA Thr	6	27
														AGC Ser		6	75
														GAG Glu		7	23
														AUC Ile		7	71
														GAG Glu		. 8	19
														GCU Ala 280		8	67
														ACA Thr		. 9	15
														AAU Asn		. 9	63
AAU Asn	CCU Pro 315	CGG Arg	AUG Met	UUC Phe	CUG Leu	GCG Ala 320	AUG Met	AUA Ile	ACA Thr	UAC Tyr	AUC Ile 325	ACA Thr	AGA Arg	AAU Asn	CAA Gln	10	11



		UGG Trp														1059
		AUG Met													_	1107
		CUC Leu														1155
		UAC Tyr 380														1203
		CUA Leu														1251
		UUC Phe														1299
CUU Leu	GGA Gly	CAA Gln	AAG Lys	AAG Lys 430	UAC Tyr	ACC Thr	AAA Lys	ACA Thr	ACA Thr 435	UAC Tyr	UGG Trp	UGG Trp	GAC Asp	GGA Gly 440	CUC Leu	1347
CAA Gln	UCC Ser	UCU Ser	GAU Asp 445	GAC Asp	UUC Phe	GCC Ala	CUC Leu	AUA Ile 450	GUG Val	AAU Asn	GCA Ala	CCA Pro	AAU Asn 455	CÁU His	GAU Asp	1395
		CAA Gln 460														1443
		AAU Asn														1491
		UUC Phe														1539
		GAG Glu														1587
		AGC Ser														1635
		GGG Gly 540														1683

								-	145	-						
	UAC Tyr 555															1731
	AGG Arg															1779
	GCA Ala															1827
	AAU Asn															1875
	GAC Asp															1923
	AAG Lys 635															1971
	CCA Pro															2019
	AUC Ile															2067
	CUU Leu													_		2115
	UUC Phe															2163
	GUG Val 715															2211
	GAG G1u															2259
	UGU Cys										UAGI	JGAAI	JUÙ		,	2305
AGC	JUGU(	ccu u	JCAU(	SAAA	AA Al	JGCCI	JUGUL	J UCI	JACU	4						2341

# (2) INFORMATION FOR SEQ ID NO:32:

### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 757 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: protein

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Met Asp Val Asn Pro Thr Leu Leu Phe Leu Lys Val Pro Ala Gln Asn 1 5 10 15

Ala Ile Ser Thr Thr Phe Pro Tyr Thr Gly Asp Pro Pro Tyr Ser His 20 25 30

Gly Thr Gly Tyr Thr Met Asp Thr Val Asn Arg Thr His Gln 35 40 45.

Tyr Ser Glu Lys Gly Lys Trp Thr Thr Asn Thr Glu Thr Gly Ala His 50 55 60

Gln Leu Asn Pro Ile Asp Gly Pro Leu Pro Glu Asp Asn Glu Pro Ser 65 70 75 80

Gly Tyr Ala Gln Thr Asp Cys Val Leu Glu Ala Met Ala Phe Leu Glu 85 90 • 95

Glu Ser His Pro Gly Ile Phe Glu Asn Ser Cys Leu Glu Thr Met Glu 100 105 110

Val Ile Gln Gln Thr Arg Val Asp Lys Leu Thr Gln Gly Arg Gln Thr 115 120 125

Tyr Asp Trp Thr Leu Asn Arg Asn Gln Pro Ala Ala Thr Ala Leu Ala 130 135 140

Asn Thr Ile Glu Val Phe Arg Ser Asn Gly Leu Thr Ala Asn Glu Ser 145 150 155 160

Gly Arg Leu Ile Asp Phe Leu Lys Asp Val Ile Glu Ser Met Asp Lys 165 170 175

Glu Glu Met Glu Ile Thr Thr His Phe Gln Arg Lys Arg Arg Val Arg 180 185 190

Asp Asn Met Thr Lys Lys Met Val Thr Gln Arg Thr Ile Gly Lys Lys 195 200 205 Lys Gln Arg Leu Asn Lys Arg Ser Tyr Leu Ile Arg Ala Leu Thr Leu Asn Thr Met Thr Lys Asp Ala Glu Arg Gly Lys Leu Lys Arg Ala 225 230 235 240 Ile Ala Thr Pro Gly Met Gln Ile Arg Gly Phe Val Tyr Phe Val Glu Thr Leu Ala Arg Ser Ile Cys Glu Lys Leu Glu Gln Ser Gly Leu Pro Val Gly Gly Asn Glu Lys Lys Ala Lys Leu Ala Asn Val Val Arg Lys Met Met Thr Asn Ser Gln Asp Thr Glu Leu Ser Phe Thr Ile Thr Gly 295 Asp Asn Thr Lys Trp Asn Glu Asn Gln Asn Pro Arg Met Phe Leu Ala Met Ile Thr Tyr Ile Thr Arg Asn Gln Pro Glu Trp Phe Arg Asn Val 330 Leu Ser Ile Ala Pro Ile Met Phe Ser Asn Lys Met Ala Arg Leu Gly 340 Lys Gly Tyr Met Phe Lys Ser Lys Ser Met Lys Leu Arg Thr Gln Ile Pro Ala Glu Met Leu Ala Ser Ile Asp Leu Lys Tyr Phe Asn Glu Ser Thr Arg Lys Lys Ile Glu Glu Ile Arg Pro Leu Leu Ile Asp Gly Thr 390 385 Val Ser Leu Ser Pro Gly Met Met Gly Met Phe Asn Met Leu Ser Thr Ile Leu Gly Val Ser Ile Leu Asn Leu Gly Gln Lys Lys Tyr Thr 420 425 430 Lys Thr Thr Tyr Trp Trp Asp Gly Leu Gln Ser Ser Asp Asp Phe Ala 435 Leu Ile Val Asn Ala Pro Asn His Asp Gly Ile Gln Ala Gly Val Asp 455 Arg Phe Tyr Arg Thr Cys Lys Leu Val Gly Ile Asn Met Ser Lys Lys 465 Lys Ser Tyr Ile Asn Arg Thr Gly Thr Phe Glu Phe Thr Ser Phe Phe

Tyr Arg Tyr Gly Phe Val Ala Asn Phe Ser Met Glu Leu Pro Ser Phe 505 Gly Val Ser Gly Ile Asn Glu Ser Ala Asp Met Ser Ile Gly Val Thr Val Ile Lys Asn Asn Met Ile Asn Asn Asp Leu Gly Pro Ala Thr Ala Gln Leu Ala Leu Gln Leu Phe Ile Lys Asp Tyr Arg Tyr Thr Tyr Arg 545 550 555 560 Cys His Arg Gly Asp Thr Gln Ile Gln Thr Arg Arg Ser Phe Glu Leu Lys Lys Leu Trp Gly Gln Thr Arg Ser Lys Ala Gly Leu Leu Val Ser 580 585 590 Asp Gly Gly Pro Asn Leu Tyr Asn Ile Arg Asn Leu His Ile Pro Glu Val Cys Leu Lys Trp Glu Leu Met Asp Glu Asp Tyr Gln Gly Arg Leu Cys Asn Pro Leu Asn Pro Phe Val Ser His Lys Glu Ile Glu Ser Val Asn Asn Ala Val Val Met Pro Ala His Gly Pro Ala Lys Ser Met Glu Tyr Asp Ala Val Thr Thr Thr His Ser Trp Ile Pro Lys Arg Asn Arg Ser Ile Leu Asn Thr Ser Gln Arg Gly Ile Leu Glu Asp Glu Gln Met 675 Tyr Gln Lys Cys Cys Asn Leu Phe Glu Lys Phe Phe Pro Ser Ser Ser 695 Tyr Arg Arg Pro Val Gly Ile Ser Ser Met Val Glu Ala Met Val Ser 705 710 715 Arg Ala Arg Ile Asp Ala Arg Ile Asp Phe Glu Ser Gly Arg Ile Lys 725 730 735 Lys Glu Glu Phe Ala Glu Ile Met Lys Ile Cys Ser Thr Ile Glu Glu 740 Leu Arg Arg Gln Lys 755

## (2) INFORMATION FOR SEQ ID NO:33:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2233 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Influenza virus
- (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg. Passage 2(3)

#### (vii) IMMEDIATE SOURCE:

(B) CLONE: PA

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(20, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain and in wt2(3); u in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(75, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); u in 1988 reported wild type E28-32 strain" /citation= ([1][2])

## (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1861, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); a in 1988 reported wild type E28-32 strain" /citation= ([1][2])

## (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(2167..2168, "cc")
- (D) OTHER INFORMATION: /note= "cc in ca "master" strain and in wt2(3); uu in 1988 reported wild type E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 25..2172
- (D) OTHER INFORMATION: /product= "polymerase acidic protein" /gene= "PA" /note= "polymerase acidic protein" /citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R G
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:33: FROM 1 TO 2233

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J
Kitame, F
Kendal, A P
Maassab, H F
Naeve, C

(B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza strain, A/Ann Arbor/6/60(H2N2)

(C) JOURNAL: Virology

(D) VOLUME: 167

(F) PAGES: 554-567

(G) DATE: 1988

(K) RELEVANT RESIDUES IN SEQ ID NO:33: FROM 1 TO 2233

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

AGC(	GAAA(	GCA (	GUA(	CUGA	TC C(				CAA ( Gln (		51
	CCG Pro										99
	GAU Asp										147
	GAA Glu										195
	GAG Glu										243
	AGA Arg 75										291
	GUA Val										339

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				UAU Tyr 110												387
				GAA Glu												435
				AAG Lys												483
				AAG Lys												531
				AGA Arg												579
				UCC Ser 190												627
				GAA Glu												675
				AAC Asn												723
				CCG Pro												771
UCC Ser 250	AAA Lys	GAA Glu	GUA Val	AAU Asn	GCU Ala 255	AAA Lys	AUU Ile	GAA Glu	CCU Pro	UUU Phe 260	CUG Leu	AAA Lys	ACA Thr	ACA Thr	CCA Pro 265	819
				CUU Leu 270												867
				GAU Asp												. 915
				AUA Ile												963
				AAA Lys												1011

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		CCA Pro														1059
		AUU Ile														1107
		AGU Ser														1155
AAG Lys	GUA Val	GAC Asp 380	UUU Phe	GAC Asp	GAC Asp	UGU Cys	AGA Arg 385	GAU Asp	GUA Val	AGC Ser	GAU Asp	UUG Leu 390	AAG Lys	CAA Gln	UAU Tyr	1203
		GAU Asp														1251
		AAC Asn														1299
		AUU Ile														1347
AGA Arg	AGG Arg	AAU Asn	UAC Tyr 445	UUC Phe	ACA Thr	GCA Ala	GAG G1u	GUG Val 450	UCU Ser	CAU His	UGC Cys	AGA Arg	GCC Ala 455	ACA Thr	GAA Glu	1395
		AUG Met 460														1443
		GCA Ala														1491
AGA Arg 490	ACU Thr	AAA Lys	GAG G1u	GGA Gly	AGG Arg 495	CGA Arg	AAG Lys	ACC Thr	AAU Asn	UUA Leu 500	UAU Tyr	GGU Gly	UUC Phe	AUC Ile	AUA Ile 505	1539
AAA Lys	GGA Gly	AGA Arg	UCU Ser	CAC His 510	UUA Leu	AGG Arg	AAU Asn	GAC Asp	ACC Thr 515	GAC Asp	GUG Val	GUA Val	AAC Asn	UUU Phe 520	GUG Val	1587
		GAG Glu														1635
GAG Glu	AAG Lys	UAC Tyr 540	Cys	GUU Val	CUU Leu	GAG Glu	AUA Ile 545	GGA Gly	GAU Asp	AUG Met	CUA Leu	CUA Leu 550	AGA Arg	AGU Ser	GCC Ala	1683

		CAG Gln														1731
ACA Thr 570	UCA Ser	AAG Lys	AUU Ile	AAA Lys	AUG Met 575	AAA Lys	UGG Trp	GGA Gly	AUG Met	GAG G1u 580	AUG Met	AGG Arg	CGU Arg	UGC Cys	CUC Leu 585	1779
CUU Leu	CAG Gln	UCA Ser	CUC Leu	CAA Gln 590	CAA Gln	AUC Ile	GAG Glu	AGU Ser	AUG Met 595	AUU Ile	GAA Glu	GCC Ala	GAG Glu	UCC Ser 600	UCU Ser	1827
GUC Val	AAG Lys	GAG Glu	AAA Lys 605	GAC Asp	AUG Met	ACC Thr	AAA Lys	GAG Glu 610	UUU Phe	UUC Phe	GAG Glu	AAU Asn	AAA Lys 615	UCA Ser	GAA Glu	1875
ACA Thr	UGG Trp	CCC Pro 620	AUU Ile	GGA Gly	GAG Glu	UCC Ser	CCC Pro 625	AAA Lys	GGA Gly	GUG Val	GAA Glu	GAA Glu 630	GGU Gly	UCC Ser	AUU Ile	1923
GGG Gly	AAG Lys 635	GUC Val	UGC Cys	AGG Arg	ACU Thr	UUA Leu 640	UUA Leu	GCC Ala	AAG Lys	UCG Ser	GUA Val 645	UUC Phe	AAU Asn	AGC Ser	CUG Leu	1971
		UCU Ser														2019
		GUC Val														2067
		GGG Gly														2115
		GUU Val 700														2163
	CCA Pro 715	AGA Arg	UAGI	JUGU(	GC A	AAUG(	CUACL	JA Ul	JUGCI	JAUC(	C AUA	ACUGI	JCCA			2212
AAA	\AGUA	ACC L	JUGUL	JUCUA	AC U											2233

## (2) INFORMATION FOR SEQ ID NO:34:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 716 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Met Glu Asp Phe Val Arg Gln Cys Phe Asn Pro Met Ile Val Glu Leu
1 5 10 15

Ala Glu Lys Ala Met Lys Glu Tyr Gly Glu Asp Leu Lys Ile Glu Thr 20 25 30

Asn Lys Phe Ala Ala Ile Cys Thr His Leu Glu Val Cys Phe Met Tyr 35 40 45

Ser Asp Phe His Phe Ile Asn Glu Gln Gly Glu Ser Ile Ile Val Glu 50 55 60

Leu Asp Asp Pro Asn Ala Leu Leu Lys His Arg Phe Glu Ile Ile Glu 65 70 75 80

Gly Arg Asp Arg Thr Met Ala Trp Thr Val Val Asn Ser Ile Cys Asn 85 90 95

Thr Thr Gly Ala Glu Lys Pro Lys Phe Leu Pro Asp Leu Tyr Asp Tyr 100 105 110

Lys Glu Asn Arg Phe Ile Glu Ile Gly Val Thr Arg Arg Glu Val His 115 120 125

Ile Tyr Tyr Leu Glu Lys Ala Asn Lys Ile Lys Ser Glu Lys Thr His 130 135 140

Ile His Ile Phe Ser Phe Thr Gly Glu Glu Met Ala Thr Lys Ala Asp 145 150 155 160

Tyr Thr Leu Asp Glu Glu Ser Arg Ala Arg Ile Lys Thr Arg Leu Phe 165 170 175

Thr Ile Arg Gln Glu Met Ala Ser Arg Gly Leu Trp Asp Ser Phe His 180 185 190

Gln Ser Glu Arg Gly Glu Glu Thr Ile Glu Glu Arg Phe Glu Ile Thr 195 200 205 Gly Thr Met Arg Arg Leu Ala Asp Gln Ser Leu Pro Pro Asn Phe Ser 215 Cys Leu Glu Asn Phe Arg Ala Tyr Val Asp Gly Phe Glu Pro Asn Gly 240 Tyr Ile Glu Gly Lys Leu Ser Gln Met Ser Lys Glu Val Asn Ala Lys Ile Glu Pro Phe Leu Lys Thr Thr Pro Arg Pro Ile Arg Leu Pro Asp Gly Pro Pro Cys Ser Gln Arg Ser Lys Phe Leu Leu Met Asp Ala Leu Lys Leu Ser Ile Glu Asp Pro Ser His Glu Gly Glu Gly Ile Pro Leu Tyr Asp Ala Ile Lys Cys Met Arg Thr Phe Phe Gly Trp Lys Glu Pro 315 Tyr Val Val Lys Pro His Glu Lys Gly Ile Asn Pro Asn Tyr Leu Leu Ser Trp Lys Gln Val Leu Ala Glu Leu Gln Asp Ile Glu Asn Glu Glu 345 Lys Ile Pro Arg Thr Lys Asn Met Lys Lys Thr Ser Gln Leu Lys Trp 360 Ala Leu Gly Glu Asn Met Ala Pro Glu Lys Val Asp Phe Asp Asp Cys 375 370 Arg Asp Val Ser Asp Leu Lys Gln Tyr Asp Ser Asp Glu Pro Glu Leu 395 Arg Ser Leu Ser Ser Trp Ile Gln Asn Glu Phe Asn Lys Ala Cys Glu 410 Leu Thr Asp Ser Ile Trp Ile Glu Leu Asp Glu Ile Gly Glu Asp Val 420 Ala Pro Ile Glu His Ile Ala Ser Met Arg Arg Asn Tyr Phe Thr Ala 440 435 Glu Val Ser His Cys Arg Ala Thr Glu Tyr Ile Met Lys Gly Val Tyr 455 Ile Asn Thr Ala Leu Leu Asn Ala Ser Cys Ala Ala Met Asp Asp Phe 465 Gln Leu Ile Pro Met Ile Ser Lys Cys Arg Thr Lys Glu Gly Arg Arg 485

Lys Thr Asn Leu Tyr Gly Phe Ile Ile Lys Gly Arg Ser His Leu Arg 500 505 510

Asn Asp Thr Asp Val Val Asn Phe Val Ser Met Glu Phe Ser Leu Thr 515 520 525

Asp Pro Arg Leu Glu Pro His Lys Trp Glu Lys Tyr Cys Val Leu Glu 530 540

Ile Gly Asp Met Leu Leu Arg Ser Ala Ile Gly Gln Val Ser Arg Pro 545 550 560

Met Phe Leu Tyr Val Arg Thr Asn Gly Thr Ser Lys Ile Lys Met Lys 565 570 575

Trp Gly Met Glu Met Arg Arg Cys Leu Leu Gln Ser Leu Gln Gln Ile 580 585 590

Glu Ser Met Ile Glu Ala Glu Ser Ser Val Lys Glu Lys Asp Met Thr 595 600 605

Lys Glu Phe Phe Glu Asn Lys Ser Glu Thr Trp Pro Ile Gly Glu Ser 610 615 620

Pro Lys Gly Val Glu Glu Gly Ser Ile Gly Lys Val Cys Arg Thr Leu 625 630 635 640

Leu Ala Lys Ser Val Phe Asn Ser Leu Tyr Ala Ser Pro Gln Leu.Glu 645 650 655

Gly Phe Ser Ala Glu Ser Arg Lys Leu Leu Leu Val Val Gln Ala Leu 660 665 670

Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Leu Gly Gly Leu Tyr Glu 675 680 685

Ala Ile Glu Glu Cys Leu Ile Asn Asp Pro Trp Val Leu Leu Asn Ala 690 695 700

Ser Trp Phe Asn Ser Phe Leu Thr His Ala Pro Arg 705 710 715

#### (2) INFORMATION FOR SEQ ID NO:35:

#### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1773 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: HA
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(144, "a")
  - (D) OTHER INFORMATION: /gene= "HA"

/note= "u in ca "master" strain; a in

wt2(3)"

/citation= ([1])

- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(455, "g")
  - (D) OTHER INFORMATION: /gene= "HA"

/note= "a in ca "master" strain; g in

wt2(3)"

/citation= ([1])

- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(729, "a")
  - (D) OTHER INFORMATION: /gene= "HA"

/note= "c in ca "master" strain; a in

wt2(3)"

/citation= ([1])

## (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 44..1729

(D) OTHER INFORMATION: /product= "hemagglutinin"

/gene= "HA"

/note= "hemagglutinin protein"

/citation=([1])

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Herlocher, M L

Maassab, H F Webster, R G

(B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus

(C) JOURNAL: Proceedings of the National Academy of Sciences of the USA

the ot

(G) DATE: 1993

(K) RELEVANT RESIDUES IN SEQ ID NO:35: FROM 1 TO 1773

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

AGCA	<b>1</b> 444(	GCA (	GGGGI	JUAUA	AC CA	AUAG/	ACAA(	C CAA	AAAG(	CAAA	ACA	GCC Ala	 	55·
		AUU Ile												103
		CAU His												151
		GUC Val												199
		AAG Lys 55												247
		AGC Ser												295

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		AGU Ser														343
AGA Arg	AAC Asn	GGU Gly	UUG Leu	UGU Cys 105	UAU Tyr	CCA Pro	GGC Gly	AAC Asn	UUC Phe 110	AAU Asn	GAU Asp	UAU Tyr	GAA Glu	GAA Glu 115	UUG Leu	391
		CUC Leu														439
CCC Pro	AAA Lys	GAU Asp 135	AGA Arg	UGG Trp	GCA Ala	CAG Gln	CAU His 140	ACA Thr	ACA Thr	ACU Thr	GGA Gly	GGU Gly 145	UCA Ser	CAG Gln	GCC Ala	487
		GUG Val													CUG Leu .	535
		GAA Glu														583
		GGA Gly														631
		ACA Thr														679
		GGC Gly 215														727
		CCU Pro														775
		UUG Leu														823
		GCA Ala														871
GGG Gly	AUC Ile	AUG Met	AAA Lys 280	ACA Thr	GAA Glu	GGA Gly	ACA Thr	CUU Leu 285	GAG G1u	AAC Asn	UGU Cys	GAG Glu	ACC Thr 290	AAA Lys	UGC Cys	919
CAA Gln	ACU Thr	CCU Pro 295	UUG Leu	GGA Gly	GCA Ala	AUA Ile	AAU Asn 300	ACA Thr	ACA Thr	UUG Leu	CCU Pro	UUU Phe 305	CAC His	AAU Asn	GUC Val	967

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		CUG Leu															1015
		UUA Leu														,	1063
		UUU Phe															1111
		GAU Asp															1159
		GCA Ala 375															1207
		AAG Lys															1255
		GGG Gly															1303
		AAG Lys													Ala		1351
		CUA Leu															1399
		GUC Val 455															1447
		AAA Lys															1495
		GAA Glu															1543
		GAA Glu															1591
AAA Lys	UUG Leu	AGC Ser	AGC Ser 520	AUG Met	GGG Gly	GUU Va 1	UGU Cys	CGG Arg 525	AUC Ile	CUU Leu	GCC Ala	AUU Ile	UAU Tyr 530	GCU Ala	ACA Thr		1639

								-	102	-						
			UCU Ser													1687
			UCC Ser													1729
UGA	JUAU	AAG (	JCAUL	JUUAl	JA AL	JUAA	VAAC/	A CC(	CUUGI	JUUC	UACI	J				1773
(2)	INF	ORMA <sup>-</sup>	TION	FOR	SEQ	ID N	NO : 36	5:								
	ĺ	(i) S	SEQUE	ENCE	CHAF	RACTE	ERIS	ΓICS	:							
			(A)	LEN	NGTH:	: 562	2 am	ino a	acids	5						
			(B)	YYT (	PE: a	amino	ac	id								
			(D)	) TOF	POLO	aY:	linea	ar								
	,		401.50	<b>-</b>	<b>**</b> \\D.	_										
	( -	ו (דו	10LEC	JULE	TYPE	:: pr	rote	ın								
	()	ki) S	SEQUE	ENCE	DESC	CRIP	TION:	: SE(	Q ID	NO:3	36:					
Met 1	Ala	Ile	Пе	Tyr 5	Leu	Ile	Leu	Leu	Phe 10	Thr	Ala	Val	Arg	Gly 15	Asp	
Lys	Ile	Cys	Ile 20	Gly	Tyr	His	Ala	Asn 25	Asn	Ser	Thr	Glu	Thr 30	Val	Asp	
Thr	Asn	Leu 35	Glu	Arg	Asn	Val	Thr 40	Val	Thr	His	Ala	Lys 45	Asp	Пe	Leu	
Glu	Lys 50	Thr	His	Asn	Gly	Lys 55	Leu	Cys	Lys	Leu	Asn 60	Gly	Ile	Pro	Pro	
Leu 65	Glu	Leu	Gly	Asp	Cys 70	Ser	IJе	Ala	Gly	Trp 75	Leu	Leu	Gly	Asn	Pro 80	
Glu	Cys	Asp	Arg	Leu 85	Leu	Ser	Val	Pro	G1u 90	Trp	Ser	Tyr	Пe	Met 95	Glu	
Lys	Glu	Asn	Pro 100	Arg	Asn	Gly	Leu	Cys 105	Tyr	Pro	Gly	Asn	Phe 110	Asn	Asp	
Tyr	Glu	Glu 115	Leu	Lys	His	Leu	Leu 120	Ser	Ser	Val	Lys	His 125	Phe	Glu	Lys	
Val	Lys 130	Ile	Leu	Pro	Lys	Asp 135	Arg	Trp	Ala	Gln	His 140	Thr	Thr	Thr	Gly	

Gly Ser Gln Ala Cys Ala Val Ser Gly Asn Pro Ser Phe Phe Arg Asn 145 155 160 Met Val Trp Leu Thr Glu Glu Gly Ser Asn Tyr Pro Val Ala Lys Gly 165 Ser Tyr Asn Asn Thr Ser Gly Glu Gln Met Leu Ile Ile Trp Gly Val 185 His His Pro Ile Asp Glu Thr Glu Gln Arg Thr Leu Tyr Gln Asn Val Gly Thr Tyr Val Ser Val Gly Thr Ser Thr Leu Asn Lys Arg Ser Thr 215 Pro Glu Ile Ala Lys Arg Pro Lys Val Asn Gly Leu Gly Ser Arg Met 225 Glu Phe Ser Trp Thr Leu Leu Asp Met Trp Asp Thr Ile Asn Phe Glu 245 Ser Thr Gly Asn Leu Ile Ala Pro Glu Tyr Gly Phe Lys Ile Ser Lys 260 265 Arg Gly Ser Ser Gly Ile Met Lys Thr Glu Gly Thr Leu Glu Asn Cys 275 280 285 Glu Thr Lys Cys Gln Thr Pro Leu Gly Ala Ile Asn Thr Thr Leu Pro Phe His Asn Val His Pro Leu Thr Ile Gly Glu Cys Pro Lys Tyr Val 310 305 Lys Ser Glu Lys Leu Val Leu Ala Thr Gly Leu Arg Asn Val Pro Gln 325 330 Ile Glu Ser Arg Gly Leu Phe Gly Ala Ile Ala Gly Phe Ile Glu Gly 345 Gly Trp Gln Gly Met Val Asp Gly Trp Tyr Gly Tyr His His Ser Asn 355 Asp Gln Gly Ser Gly Tyr Ala Ala Asp Lys Glu Ser Thr Gln Lys Ala 375 380 Phe Asp Gly Ile Thr Asn Lys Val Asn Ser Val Ile Glu Lys Ile Asn 390 395 400 Thr Gln Phe Glu Ala Val Gly Lys Glu Phe Ser Asn Leu Glu Arg Arg 405 Leu Glu Asn Leu Asn Lys Lys Met Glu Asp Gly Phe Leu Asp Val Trp 420 425

Thr Tyr Asn Ala Glu Leu Leu Val Leu Met Glu Asn Glu Arg Thr Leu 435 440 445

Asp Phe His Asp Ser Asn Val Lys Asn Leu Tyr Asp Lys Val Arg Met 450 460

Gln Leu Arg Asp Asn Val Lys Glu Leu Gly Asn Gly Cys Phe Glu Phe. 465 470 475 480

Tyr His Lys Cys Asp Asp Glu Cys Met Asn Ser Val Lys Asn Gly Thr 485 490 495

Tyr Asp Tyr Pro Lys Tyr Glu Glu Glu Ser Lys Leu Asn Arg Asn Glu 500 505 510

Ile Lys Gly Val Lys Leu Ser Ser Met Gly Val Cys Arg Ile Leu Ala 515 520 525

Ile Tyr Ala Thr Val Ala Gly Ser Leu Ser Leu Ala Ile Met Met Ala 530 535 540

Gly Ile Ser Phe Trp Met Cys Ser Asn Gly Ser Leu Gln Cys Arg Ile 545 550 560

Cys Ile

#### (2) INFORMATION FOR SEQ ID NO:37: ·

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1467 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)

#### (vii) IMMEDIATE SOURCE:

(B) CLONE: NA

#### (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(394, "c")
- (D) OTHER INFORMATION: /product= "Neuraminidase"

/gene= "NA"

/note= "u in ca·"master" strain; c in

wt2(3)"

/citation= ([1])

## (ix) FEATURE:

- (A) NAME/KEY: mutation
- (B) LOCATION: replace(604, "a")
- (D) OTHER INFORMATION: /product= "Neuraminidase"

/gene= "NA"

/note= "u in ca "master" strain; a in

wt2(3)"

/citation=([1])

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 20..1426
- (D) OTHER INFORMATION: /product= "neuraminidase"

/gene= "NA"

/note= "neuraminidase protein"

/citation= ([1])

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Herlocher, M L

Maassab, H F

Webster, R G

- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) Influenza Virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:37: FROM 1 TO 1467

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

AGCAAAAGCA	GGAGUGAAA AL Me	G AAU CCA AA t Asn Pro As 1	U CAA AAG ACA n Gln Lys Thr 5	A AUA ACA AUU 11e Thr I1e 10	e G1y
UCU GUC UCU Ser Val Ser	CUC ACC AUG Leu Thr Ile 15	GCA ACA GUA Ala Thr Val 20	UGC UUC CUC Cys Phe Leu	AUG CAG AUU Met Gln Ile 25	GCC 100 Ala
AUC CUG GCA Ile Leu Ala 30	Thr Thr Va	ACA UUG CAC Thr Leu His 35	CUU AAG CAA Leu Lys Gln	CAU GAG UGC His Glu Cys 40	GAC 148 Asp
UCC CCC GCG Ser Pro Ala 45	AGC AAC CA Ser Asn Gl	A GUA AUG CCA n Val Met Pro 50	UGU GAA CCA Cys Glu Pro 55	AUA AUA AUA Ile Ile Ile	GAA 196 Glu
AGG AAC AUA Arg Asn Ile 60	ACA GAG AU Thr Glu Ilo 6	e Val Tyr Leu	AAU AAC ACC AASN ASN Thr 70	ACC AUA GAG Thr Ile Glu	AAA 244 Lys 75
GAG AUU UGC Glu Ile Cys	CCC GAA GU Pro Glu Va 80	A GUG GGA UAC I Val Gly Tyr	AGA AAU UGG Arg Asn Trp 85	UCA AAG CCG Ser Lys Pro 90	CAA 292 Gln
UGU CAA AUU Cys Gln Ile	ACA GGA UU Thr Gly Ph 95	J GCA CCU UUL e Ala Pro Phe 100	J UCU AAG GAC e Ser Lys Asp )	AAU UCA AUC Asn Ser Ile 105	CGG 340 Arg
CUU UCU GCU Leu Ser Ala 110	Gly Gly As	C AUU UGG GUG D Ile Trp Va 115	G ACG AGA GAA I Thr Arg Glu	CCU UAU GUG Pro Tyr Val 120	UCA 388 Ser
UGC GAC CCU Cys Asp Pro 125	GGC AAG UG Gly Lys Cy	J UAU CAA UUU s Tyr Gln Phe 130	J GCA CUC GGG e Ala Leu Gly 135	CAG GGG ACC Gln Gly Thr	ACA 436 Thr
CUA GAC AAC Leu Asp Asr 140	AAA CAU UC Lys His Se 14	r Asn Gly Thi	A AUA CAU GAU ^ Ile His Asp 150	AGA AUC CCU Arg Ile Pro	CAU 484 His 155
CGA ACC CUA Arg Thr Leu	A UUA AUG AA 1 Leu Met As 160	J GAG UUG GGU n Glu Leu Gly	J GUU CCA UUU y Val Pro Phe 165	CAU UUA GGA His Leu Gly 170	ACC 532 Thr
AAA CAA GUG Lys Gln Val	G UGU GCA GC   Cys Ala Al   175	A UGG UCC AGG a Trp Ser Sei 18	C UCA AGU UGU r Ser Ser Cys O	CAC GAU GGA His Asp Gly 185	AAA 580 Lys
GCA UGG UUG Ala Trp Leu 190	ı His Val Cy	U GUC ACA GG s Val Thr Gl 195	G GAU GAU AGA y Asp Asp Arg	AAU GCA ACU Asn Ala Thr 200	GCU 628 Ala

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AGC Ser	UUC Phe 205	AUU Ile	UAU Tyr	GAC Asp	GGG Gly	AAG Lys 210	CUU Leu	GUG Val	GAC Asp	AGU Ser	AUU Ile 215	GGU Gly	UCA Ser	UGG Trp	UCU Ser	676
		GUC Val														724
		ACA Thr														772
		AUA Ile														820
		GGA Gly 270														868
		GAC Asp														916
		GUU Val														964
		UGC Cys														1012
		AAU Asn														1060
		AAA Lys 350														1108
		AUC Ile														1156
		GGU Gly														1204
		GUU Val														1252
		GGC Gly														1300

AGG GGA AGG CCA CAG GAG ACU AGA GUA UGG UGG ACC UCA AAC AGU AUU Arg Gly Arg Pro Gln Glu Thr Arg Val Trp Trp Thr Ser Asn Ser Ile 430 440	1348
GUU GUA UUU UGU GGC ACU UCA GGU ACU UAU GGA ACA GGC UCA UGG CCU Val Val Phe Cys Gly Thr Ser Gly Thr Tyr Gly Thr Gly Ser Trp Pro 445 450 455	1396
GAU GGG GCG AAC AUC AAU UUC AUG CCU AUA UAACGUUUCG CAAUUUUAGA Asp Gly Ala Asn Ile Asn Phe Met Pro Ile 460 465	1446
AAAAAACUCC UUGUUUCUAC U	1467

#### (2) INFORMATION FOR SEQ ID NO:38:

#### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 469 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

#### (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Met Asn Pro Asn Gln Lys Thr Ile Thr Ile Gly Ser Val Ser Leu Thr 1 5 10 15

Ile Ala Thr Val Cys Phe Leu Met Gln Ile Ala Ile Leu Ala Thr Thr 20 25 30

Val Thr Leu His Leu Lys Gln His Glu Cys Asp Ser Pro Ala Ser Asn 35 40 45

Gln Val Met Pro Cys Glu Pro Ile Ile Ile Glu Arg Asn Ile Thr Glu 50 55 60

Ile Val Tyr Leu Asn Asn Thr Thr Ile Glu Lys Glu Ile Cys Pro Glu 65 70 75 80

Val Val Gly Tyr Arg Asn Trp Ser Lys Pro Gln Cys Gln Ile Thr Gly 85 90 95

Phe Ala Pro Phe Ser Lys Asp Asn Ser Ile Arg Leu Ser Ala Gly Gly 100 105 110

Asp Ile Trp Val Thr Arg Glu Pro Tyr Val Ser Cys Asp Pro Gly Lys 115 120 125

Cys Tyr Gln Phe Ala Leu Gly Gln Gly Thr Thr Leu Asp Asn Lys His 130 135 Ser Asn Gly Thr Ile His Asp Arg Ile Pro His Arg Thr Leu Leu Met 150 155 Asn Glu Leu Gly Val Pro Phe His Leu Gly Thr Lys Gln Val Cys Ala Ala Trp Ser Ser Ser Cys His Asp Gly Lys Ala Trp Leu His Val Cys Val Thr Gly Asp Asp Arg Asn Ala Thr Ala Ser Phe Ile Tyr Asp 200 195 205 Gly Lys Leu Val Asp Ser Ile Gly Ser Trp Ser Gln Asn Val Leu Arg Thr Gln Glu Ser Glu Cys Val Cys Ile Asn Gly Thr Cys Thr Val Val 240 225 Met Thr Asp Gly Ser Ala Ser Gly Arg Ala Asp Thr Arg Ile Leu Phe Ile Lys Glu Gly Lys Ile Val His Ile Gly Pro Leu Ser Gly Ser Ala Gln His Val Glu Glu Cys Ser Cys Tyr Pro Arg Tyr Pro Asp Val Arg 275 280 285 Cys Ile Cys Arg Asp Asn Trp Lys Gly Ser Asn Arg Pro Val Ile Asp Ile Asn Met Glu Asp Tyr Ser Ile Asp Ser Ser Tyr Val Cys Ser Gly 305 310 315 320 Leu Val Gly Asp Thr Pro Arg Asn Asp Asp Ser Ser Ser Asn Ser Asn 335 325 Cys Arg Asp Pro Asn Asn Glu Arg Gly Asn Pro Gly Val Lys Gly Trp Ala Phe Asp Asn Gly Asp Asp Val Trp Met Gly Arg Thr Ile Ser Lys Asp Leu Arg Ser Gly Tyr Glu Thr Phe Lys Val Ile Gly Gly Trp Ser Thr Pro Asn Ser Lys Ser Gln Val Asn Arg Gln Val Ile Val Asp Asn 395 385 Asn Asn Trp Ser Gly Tyr Ser Gly Ile Phe Ser Val Glu Gly Lys Ser 410

Cys Ile Asn Arg Cys Phe Tyr Val Glu Leu Ile Arg Gly Arg Pro Gln 420 425 430

Glu Thr Arg Val Trp Trp Thr Ser Asn Ser Ile Val Val Phe Cys Gly
435 440 445

Thr Ser Gly Thr Tyr Gly Thr Gly Ser Trp Pro Asp Gly Ala Asn Ile 450 455 460

Asn Phe Met Pro Ile 465

## (2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1566 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: RNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Influenza virus
  - (B) STRAIN: wild type A/Ann Arbor/6/60 (H2N2) Egg Passage 2(3)
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: NP
- (ix) FEATURE:
  - (A) NAME/KEY: mutation
  - (B) LOCATION: replace(113, "a")
  - (D) OTHER INFORMATION: /note= "c in ca "master" strain; a in wt2(3); c in 1988 reported wild type E28-32 strain (manuscript) but a in 1988 reported wild type E28-32 strain /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(146, "g")
  - (D) OTHER INFORMATION: /note= "g in ca "master" strain and in

wt2(3); a in 1988 reported wild type

E28-32 strain" /citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(627, "c")
- (D) OTHER INFORMATION: /note= "c in ca "master" strain and in

wt2(3); a in 1988 reported wild type

E28-32 strain"

/citation= ([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(909, "g")
- (D) OTHER INFORMATION: /note= "g in ca "master" strain and in wt2(3); c in 1988 reported wild type

E28-32 strain"

/citation=([1][2])

#### (ix) FEATURE:

- (A) NAME/KEY: conflict
- (B) LOCATION: replace(1550, "a")
- (D) OTHER INFORMATION: /note= "a in ca "master" strain and in wt2(3); deletion in 1988 reported wild

type E28-32 strain" /citation= ([1][2])

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 46..1539
- (D) OTHER INFORMATION: /product= "Nucleoprotein" /gene= "NP"

/note= "nucleoprotein"
/citation= ([1][2])

#### (x) PUBLICATION INFORMATION:

- (A) AUTHORS: Herlocher, M L Maassab, H F Webster, R W
- (B) TITLE: Molecular and biological changes in the cold adapted master strain A/AA/6/60 (H2N2) influenza virus
- (C) JOURNAL: Proceedings of the National Academy of Sciences of the USA
- (G) DATE: 1993
- (K) RELEVANT RESIDUES IN SEQ ID NO:39: FROM 1 TO 1566

#### (x) PUBLICATION INFORMATION:

(A) AUTHORS: Cox, N J

Kitame, F Kendal, A P Maassab, H F Naeve, C

- (B) TITLE: Identification of sequence changes in the cold-adapted live attenuated influenza vaccine strain, A/Ann Arbor/6/60 (H2N2)
- (C) JOURNAL: Virology
- (D) VOLUME: 167
- (F) PAGES: 554-567
- (G) DATE: 1988
- (K) RELEVANT RESIDUES IN SEQ ID NO:39: FROM 1 TO 1566

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# (x1) SEQUENCE DESCRIPTION: SEQ ID NO:39:

AGCAA	<b>VAA</b> G	ica (	GGU/	AGAU#	<b>\</b> A U(	CACU(	CACU(	G AGU	JGACA	AUCA	AAAl		CG U( la Se	5	4
CAA G Gln G														10	2
CAG A Gln A 20														15	۰0
AUU G Ile G														19	8
UAU G Tyr G														. 24	.6
CUC U Leu S														29	14
AGC G Ser A														34	.2
GUA G Val A 100														39	01
AUA A Ile A														43	8
GGU C Gly L														48	6
UAC C Tyr G														53	4
UGC U Cys S 1														58	12
GGC G Gly A 180														63	10

1

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														GAG Glu 210		678
														CUC Leu		726
														GUG Val		774
GAA Glu	AGC Ser 245	CGG Arg	AAC Asn	CCA Pro	GGA Gly	AAU Asn 250	GCU Ala	GAG Glu	AUC Ile	GAA G1u	GAU Asp 255	CUC Leu	AUG Ile	UUU Phe	CUG Leu	822
GCA Ala 260	CGG Arg	UCU Ser	GCA Ala	CUC Leu	AUA Ile 265	UUG Leu	AGA Arg	GGG Gly	UCA Ser	GUU Val 270	GCU Ala	CAC His	AAA Lys	UCU Ser	UGU Cys 275	870
														GAC Asp 290		918
														CUG Leu		966
CAA Gln	AAC Asn	AGC Ser 310	CAA Gln	GUA Val	UAC Tyr	AGC Ser	CUA Leu 315	AUC Ile	AGA Arg	CCG Pro	AAU Asn	GAG G1u 320	AAU Asn	CCA Pro	GCA Ala	1014
														UUU Phe		1062
														CCA Pro		1110
														AAC Asn 370		1158
														UGG Trp		1206
														UCU Ser		1254
														CUC Leu		1302

	GAC Asp														1350
	ACA Thr														1398
	CCA Pro														1446
	GAA Glu														1494
	GGA Gly 485														1539
UAAGGAAAA AUACCCUUGU UUCUACU													1566		

## (2) INFORMATION FOR SEQ ID NO:40:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 498 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Met Ala Ser Gln Gly Thr Lys Arg Ser Tyr Glu Gln Met Glu Thr Asp 1 5 10 15

Gly Glu Arg Gln Asn Ala Asn Glu Ile Arg Ala Ser Val Gly Lys Met 20 25 30

The Gly Gly Ile Gly Arg Phe Tyr Ile Gln Met Cys Thr Glu Leu Lys 35 40 45

Leu Ser Asp Tyr Glu Gly Arg Leu Ile Gln Asn Ser Leu Thr Ile Glu 50 60

Arg Met Val Leu Ser Ala Phe Asp Glu Arg Arg Asn Lys Tyr Leu Glu 65 70 75 80

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Glu His Pro Ser Ala Gly Lys Asp Pro Lys Lys Thr Gly Gly Pro Ile Tyr Lys Arg Val Asp Gly Lys Trp Met Arg Glu Leu Val Leu Tyr Asp Lys Glu Glu Ile Arg Arg Ile Trp Arg Gln Ala Asn Asn Gly Asp Asp 115 Ala Thr Ala Gly Leu Thr His Met Met Ile Trp His Ser Asn Leu Asn 135 Asp Thr Thr Tyr Gln Arg Thr Arg Ala Leu Val Arg Thr Gly Met Asp Pro Arg Met Cys Ser Leu Met Gln Gly Ser Thr Leu Pro Arg Arg Ser 165 170 175 Gly Ala Ala Gly Ala Ala Val Lys Gly Val Gly Thr Met Val Met Glu Leu Ile Arg Met Ile Lys Arg Gly Ile Asn Asp Arg Asn Phe Trp Arg 195 200 205 Gly Glu Asn Gly Arg Lys Thr Arg Asn Ala Tyr Glu Arg Met Cys Asn 215 Ile Leu Lys Gly Lys Phe Gln Thr Ala Ala Gln Arg Ala Met Met Asp Gln Val Arg Glu Ser Arg Asn Pro Gly Asn Ala Glu Ile Glu Asp Leu Ile Phe Leu Ala Arg Ser Ala Leu Ile Leu Arg Gly Ser Val Ala His 265 Lys Ser Cys Leu Pro Ala Cys Val Tyr Gly Pro Ala Val Ala Ser Gly 275 280 285 Tyr Asp Phe Glu Lys Glu Gly Tyr Ser Leu Val Gly Ile Asp Pro Phe 290 295 300 Lys Leu Leu Gln Asn Ser Gln Val Tyr Ser Leu Ile Arg Pro Asn Glu 315 320 Asn Pro Ala His Lys Ser Gln Leu Val Trp Met Ala Cys Asn Ser Ala 325 330 335 Ala Phe Glu Asp Leu Arg Val Ser Ser Phe Ile Arg Gly Thr Lys Val 340 345 350 Ile Pro Arg Gly Lys Leu Ser Thr Arg Gly Val Gln Ile Ala Ser Asn 360 365

Glu Asn Met Asp Thr Met Gly Ser Ser Thr Leu Glu Leu Arg Ser Arg 370 375 380

Tyr Trp Ala Ile Arg Thr Arg Ser Gly Gly Asn Thr Asn Gln Gln Arg 385 390 395 400

Ala Ser Ala Gly Gln Ile Ser Val Gln Pro Thr Phe Ser Val Gln Arg 405 410 415

Asn Leu Pro Phe Asp Lys Pro Thr Ile Met Ala Ala Phe Thr Gly Asn 420 425 430

Ala Glu Gly Arg Thr Ser Asp Met Arg Ala Glu Ile Ile Arg Met Met 435 440 445

Glu Gly Ala Lys Pro Glu Glu Val Ser Phe Gln Gly Arg Gly Val Phe 450 455 460

Glu Leu Ser Asp Glu Lys Ala Thr Asn Pro Ile Val Pro Ser Phe Asp 465 470 475 480

Met Ser Asn Glu Gly Ser Tyr Phe Phe Gly Asp Asn Ala Glu Glu Tyr 485 490 495

Asp Asn